

Journal Pre-proof

Glycolysis in human cancers: Emphasis circRNA/glycolysis axis and nanoparticles in glycolysis regulation in cancer therapy

Ali G. Alkhathami, Ameer S. Sahib, Majed Saad Al Fayi, Ali Abdulhussain Fadhil, Mohammed Abed Jawad, Sahar Ahmad Shafik, Safwan Jassim Sultan, Abbas F. Almulla, Min Shen

PII: S0013-9351(23)00799-5

DOI: <https://doi.org/10.1016/j.envres.2023.116007>

Reference: YENRS 116007

To appear in: *Environmental Research*

Received Date: 26 February 2023

Revised Date: 24 April 2023

Accepted Date: 26 April 2023



Please cite this article as: Alkhathami, A.G., Sahib, A.S., Al Fayi, M.S., Fadhil, A.A., Jawad, M.A., Shafik, S.A., Sultan, S.J., Almulla, A.F., Shen, M., Glycolysis in human cancers: Emphasis circRNA/glycolysis axis and nanoparticles in glycolysis regulation in cancer therapy, *Environmental Research* (2023), doi: <https://doi.org/10.1016/j.envres.2023.116007>.

This is a PDF file of an article that has undergone enhancements after acceptance, such as the addition of a cover page and metadata, and formatting for readability, but it is not yet the definitive version of record. This version will undergo additional copyediting, typesetting and review before it is published in its final form, but we are providing this version to give early visibility of the article. Please note that, during the production process, errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

© 2023 Published by Elsevier Inc.

Glycolysis in human cancers: Emphasis circRNA/glycolysis axis and nanoparticles in glycolysis regulation in cancer therapy

Ali G. Alkhathami^{1*}, Ameer S. Sahib², Majed Saad Al Fayi³, Ali Abdulhussain Fadhil⁴,
Mohammed Abed Jawad⁵, Sahar Ahmad Shafik^{6,7}, **Safwan Jassim Sultan**⁸, Abbas F. Almulla⁹,
Min Shen^{10*}

¹ Department of Clinical Laboratory Sciences, College of Applied Medical Sciences, King Khalid University, Abha, Saudi Arabia

² Department of Pharmacy, Al- Mustaqbal University College, 51001 Hilla, Iraq

³ Department of Clinical Laboratory Sciences, College of Applied Medical Sciences, King Khalid University, Abha, Saudi Arabia

⁴ College of Medical Technology, Medical Lab techniques; Al-Farahidi University, Iraq

⁵ Department of Medical Laboratories Technology, Al-Nisour University College, Iraq

⁶ Professor of Community Health Nursing, Faculty of Nursing, Fayum University, Egypt

⁷ College of Nursing, National University of Science and Technology, Iraq

⁸ **Department of Dentistry, AlNoor University College, Nineveh, Iraq**

⁹ Medical Laboratory Technology Department, College of Medical Technology, The Islamic University, Najaf, Iraq

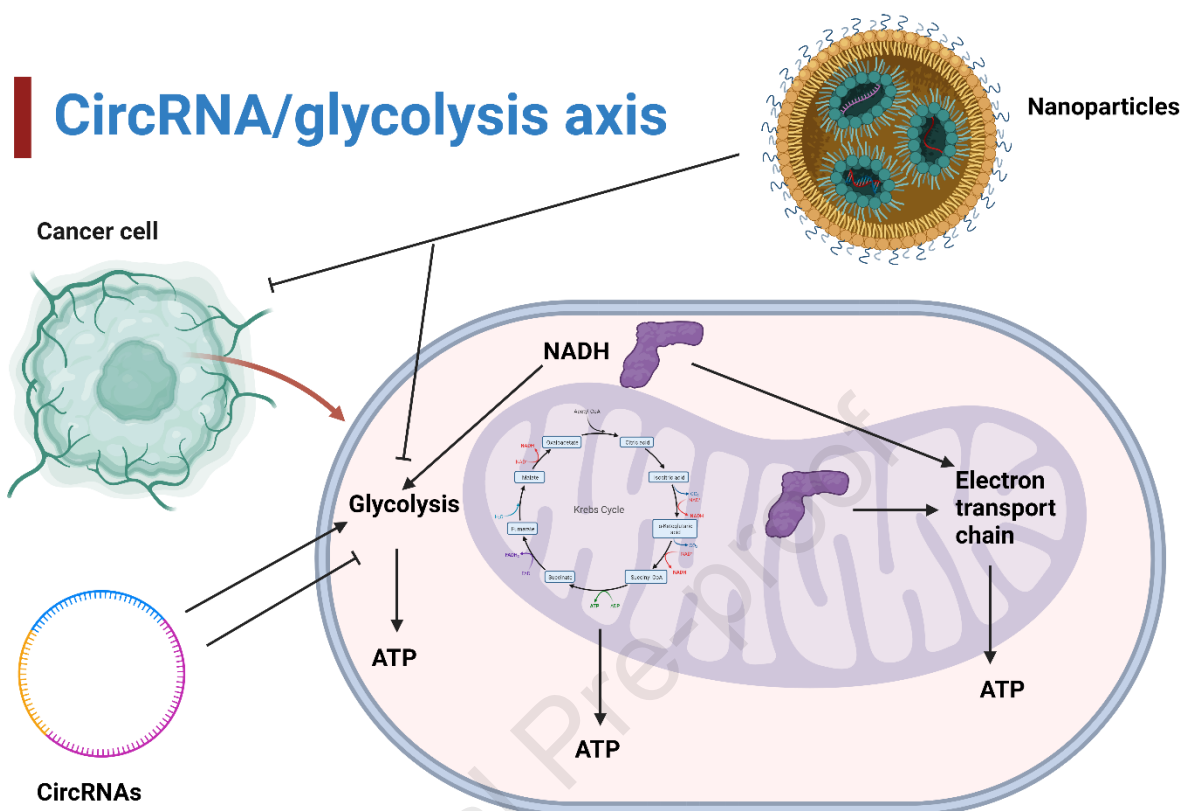
¹⁰ Department of Cardiology, Xijing Hospital, The Fourth Military Medical University, China

Corresponding author:

Ali G. Alkhathami, Department of Clinical Laboratory Sciences, College of Applied Medical Sciences, King Khalid University, Abha, Saudi Arabia; Email: gaithan@kku.edu.sa, aligalkhathami1212@gmail.com

Min Shen, Department of Cardiology, Xijing Hospital, The Fourth Military Medical University, China, Email: 31730734@qq.com

Graphical abstract:



Abstract:

The metabolism of cancer has been an interesting hallmark and metabolic reprogramming, especially the change from oxidative phosphorylation in mitochondria to glucose metabolism known as glycolysis occurs in cancer. The molecular profile of glycolysis, related molecular pathways and enzymes involved in this mechanism such as hexokinase have been fully understood. The glycolysis inhibition can significantly decrease tumorigenesis. On the other hand, circRNAs are new emerging non-coding RNA (ncRNA) molecules with potential biological functions and aberrant expression in cancer cells which have received high attention in recent years. CircRNAs have a unique covalently closed loop structure which makes them highly stable and reliable biomarkers in cancer. CircRNAs are regulators of molecular mechanisms including glycolysis. The enzymes involved in the glycolysis mechanism such as hexokinase are regulated by circRNAs to modulate tumor progression. Induction of glycolysis by circRNAs can significantly increase proliferation rate of cancer cells given access to energy and enhance metastasis. CircRNAs regulating glycolysis can influence drug resistance in cancers because of their impact on malignancy of tumor cells upon glycolysis induction. TRIM44, CDCA3, SKA2 and ROCK1 are among the downstream targets of circRNAs in regulating glycolysis in cancer. Additionally, microRNAs are key regulators of glycolysis mechanism in cancer cells and can affect related molecular pathways and enzymes. CircRNAs sponge miRNAs to regulate glycolysis as a main upstream mediator. Moreover, nanoparticles have been emerged as new tools in tumorigenesis suppression and in addition to drug and gene delivery, then mediate cancer immunotherapy and can be used for vaccine development. The nanoparticles can delivery circRNAs in cancer therapy and they are promising candidates in regulation of glycolysis, its suppression and inhibition of related pathways such as HIF-1 α . The stimuli-responsive nanoparticles and ligand-functionalized ones have been developed for selective targeting of glycolysis and cancer cells, and mediating carcinogenesis inhibition.

Keywords: Circular RNAs, glycolysis, cancer metabolism, chemoresistance, nanoparticles, gene delivery

Contents

1. Introduction.....	4
2. Glycolysis mechanism: An overview	4
3. Climate, environment and cancer: A brief discussion	5
4. Nanoparticles in cancer therapy: An overview	6
5. Nanoparticles in regulation of glycolysis in cancer	7
6. CircRNAs induce glycolysis mechanism.....	9
7. CircRNAs suppress glycolysis mechanism.....	10
8. CircRNAs regulate drug resistance via glycolysis regulation.....	11
9. CircRNA/miRNA axis and glycolysis	14
10. Anti-cancer agents regulating circRNA/glycolysis axis	20
11. Conclusion and remarks.....	21

1. Introduction

As endogenous ncRNAs, circRNAs were recognized in 1990s with one of their features being their scrambled exon order [1]. Attempts were made to understand the structure, mechanism of action, and their roles in diseases in recent years [2-5]. The conventional linear RNA molecules have 5'caps and 3'tails, while circRNAs have closed loop structures [6]. The highly conserved ncRNA family in mammals has other members such as miRNAs and lncRNAs, with circRNAs having higher stability because of their nuclease resistance characteristics [7, 8]. The expression of circRNAs has been mentioned in various species from humans and animals to plants [9-11]. Therefore, circRNAs are evolutionary conserved in a time-dependent, cell type and gene-specific manner. Because of advances in biology and high-throughput sequencing technologies, a high number of circRNAs have been identified with roles in cardiovascular diseases, neurological disorders, cancer and other diseases [12-14]. Since circRNAs are evolutionary conserved with high stability and expression in tissues and body fluids, they can be used for diagnosis and treatment of diseases [15, 16]. The biogenesis of circRNAs was reported by Jeck and colleagues in two models [17] including intron-pairing-driven circularization and lariat-driven circularization. Intron circRNA was discovered in recent years and consists of an 11-nucleotide C-rich element and a 7-nucleotide G-rich element in the parent gene which are mixed to generate a circular structure that can undergo splicing by spliceosome [18]. The spliceosome is a vital mechanism in the biogenesis of circRNAs and relies on trans-acting factors and cis-regulatory elements [19].

CircRNAs play various roles in cells that one of them is miRNA sponging. miRNAs have the capacity of reducing gene expression at mRNA levels [20]. The intracellular mobility of miRNAs determines their sponging ability and is featured as an intermittent active transport type [21]. Hanssen's lab was one of the leaders in studying the complexity of gene expression regulation and discovered circRNA and miRNA interactions [22]. Since circRNAs have unique structures, they can be protected from destabilization and degradation by miRNA-induced deadenylation [23]. Protein binding, transcription regulation, coding for proteins and peptides are other functions of circRNAs [24]. The role of circRNAs in cancer progression regulation has been of importance in recent years. In the oncology field, circRNAs are divided into two categories including oncogenic and onco-suppressor circRNAs [25, 26]. Circ-PTCD3 is a driver of breast cancer progression and by sponging miR-198, it increases growth and colony formation of tumor cells [27]. Overexpression of circ-SEPT9 in endometrial cancer is associated with induction of methylation of miR-186 to decrease its level and enhance cancer malignancy [28]. In lung cancer, circ-CPA4 promotes TIGF2 expression via miR-214-3p inhibition thereby increasing tumorigenesis [29]. On the other hand, onco-suppressor circRNAs induce apoptosis thereby reducing cancer progression [30]. Finally, circRNAs are cancer hallmark modulators which can interact with components of tumor microenvironment such as macrophages in regulating tumor progression [31-37]. The current review focuses on the role of circRNAs in regulating EMT mechanism in various cancers and provides new insights on their role in regulation of cancer metastasis.

2. Glycolysis mechanism: An overview

Among the various metabolic pathways in cancer, glycolysis or Warburg effect has been investigated in detail [38, 39]. Otto Warburg introduced a hypothesis as a fundamental basis for cancer in 1920s which explains a metabolic shift towards glycolysis [40]. Based on this hypothesis, cancer is a multifactorial malignancy and despite showing a high number of alterations, there is a major change in replacement of oxygen respiration with sugar fragmentation. This hypothesis is called Warburg effect in which cancers transform glucose into lactate, even at the presence of oxygen also called aerobic glycolysis. In contrast, anaerobic glycolysis occurs during hypoxia and acts as an energetic adaptation to hypoxia [41-43]. In normal cells during normal conditions when there is enough oxygen, glucose is utilized for generation of pyruvate. Glucose is then oxidated in mitochondria in the TCA cycle to carbon dioxide, known as oxidative phosphorylation [44]. On the other hand, the process of glycolysis occurs in cytoplasm where glucose is transformed into pyruvate and lactate is produced. HK, PFK and PK are other regulators of glycolysis [45, 46].

Remarkably, the induction of glycolysis increases progression of cancer. ENO1-IT1 upregulation by F. nucleatum can result in stimulation of glycolysis and increase in colorectal cancer growth [27]. The neutrophils present in tumor microenvironment can secrete extracellular vesicles and lead to an increase in mRNA levels of SPI1 which mediates glycolysis and enhances colon cancer progression [47]. ISLR inhibition suppresses JAK/STAT3 and impairment of glycolysis in lung cancer [48]. NEDD4L can enhance ubiquitination of ENO1 and inducing its degradation thereby inhibiting glycolysis in oral tumor [49]. SIRT6 stimulates glycolysis in lung cancer and in turn, α -hederin reduces SIRT6 expression to prevent glycolysis [50]. Penfluridol reduces PFKL expression to inhibit AMPK/FOXO3a axis, induce apoptosis, and interfere with glycolysis in esophageal cancer [51]. Interestingly, with glycolysis induction in cancer, tumor cells show more resistance to apoptosis [52] and this may be considered as a factor for development of therapy resistance. This idea has been confirmed by the function of POU2F1-ALDOA axis in glycolysis induction which leads to mediation of resistance to chemotherapy [53]. Therefore, interaction of molecular pathways can determine glycolysis status in cancer [54, 55] and the purpose of current review is to find interactions between circRNAs and glycolysis in cancers.

3. Climate, environment and cancer: A brief discussion

It is obvious that changes in climate can cause development of cancer. Now, there is no doubt that climate change can change health of humanity [56]. Currently, world is facing climate change and various international organization had predicted that disruption in planetary ecology can impact human health [57]. Researchers have shown that changes in climate can affect prevalence of infectious diseases and alter access to healthy water and food [58]. However, there is little focus on the cancer, since it is a chronic disease and does not bring suddenly after close temporal proximity to the exposures mediated by climate change. The exposure to tobacco products, ultraviolet irradiation, infection diseases, HPV, environmental toxics, chemicals and changes in lifestyle can lead to cancer development [59]. Based on these risk factors of cancer, when climate change occurs, it is obvious that increase in incidence rate of cancer is observed. The climate change can alter the plans of cancer control through air pollution, ultraviolet irradiation, food and water supply disruption, industrial toxics and chemicals and infection

disease-related cancer. Therefore, there is strong correlation between climate change, environmental factors and cancer development, and effective control of cancer requires an international law for controlling and preventing climate change.

4. Nanoparticles in cancer therapy: An overview

Nanomaterials are promising candidates in cancer therapy. The use of nanostructures is not limited only to delivery of drugs and progresses in field of nanoengineering and nanomedicine have resulted in development of nanocarriers with multiple and versatile functions. The nanoparticles are opening their way to be used significantly in cancer immunotherapy. The BTO nanoparticles have been functionalized with cell membrane to deliver PD-L1 antibody. The engineered membranes are sensitive to MMP-2 enzyme in tumors and undergo cleavage and by increasing ROS generation, they promote CTLs infiltration and increase potential of PD-L1 immunotherapy [60]. Due to immunosuppression activity of tumor cells, application of nanoparticle-mediated immunotherapy is increasing. The PLGA nanoparticles with biodegradable feature are able to stimulate cGAS/STING axis to mediate IL-15-induced NK cell induction and enhancing cancer immunotherapy [61]. Furthermore, nanoparticles can be employed for purpose of vaccine development and improving immune system [62]. The selenium nanomaterials are able to modulate innate immune system through direct interaction with macrophages, dendritic and NK cells, and such immunotherapy can increase chemotherapy potential in cancer suppression [63]. The silver nanostructures can be stabilized by PVP and they decrease IL-6 secretion in impairing prostate cancer [64]. The targeted delivery of drugs by nanocarriers is the most common use in cancer therapy; doxorubicin has been loaded on hybrid magnetic nanostructures and then, they have been functionalized with chitosan and folic acid. These nanoparticles increase cell internalization of doxorubicin and release drug in response to pH-sensitive manner for tumor suppression [65]. The polymeric nanoparticles can co-deliver both elesclomol (ES) and Cu, and release cargo in response to ROS. When the drugs and cargo are released, they mediate cuproptosis to reduce viability of tumor cells and can stimulate anti-cancer immune responses. Moreover, they reprogram tumor microenvironment and reduce bladder cancer progression [66]. Interestingly, the identification and determination of anti-cancer drugs by nanostructures can also be performed [67, 68]. The nanoparticles are able to deeply penetrate into cancer cells and they are promising in tumor disruption [69]. In the field of chemotherapy, the application of nanoparticles is highly suggested due to their potential in improving cellular uptake of drugs [70]. Furthermore, nanoparticles can stimulate phototherapy in tumor ablation [71]. In addition, self-assembled peptide nanomaterials can target tumor microenvironment and prevent immunosuppression [72]. Hence, nanoparticles are extensively used in cancer therapy [73, 74]. However, the toxicity and biocompatibility of nanoparticles should also be considered; for instance, high concentration of glycol chitosan nanoparticles mediate cardiotoxicity [75]. **Table 1** displays application of nanoparticles in cancer therapy.

Table 1: Nanoparticles as promising tools in cancer therapy.

Nanoparticles	Aim of application	Reference
Albumin-based nanocarriers	High biocompatibility and biosafety Disrupting tumor vasculature	[76]

	Photothermal-mediated tumor ablation	
Polymeric nanoparticles	Increase in cellular uptake Oxidative damage HSP70 down-regulation	[77]
Mesoporous silica nanoparticles	pH-sensitive release of drug Increasing ROS generation Stimulation of chemodynamic therapy	[78]
Organosilica nanostructures and Au nanoparticles	Proliferation suppression Cell death induction	[79]
Lipid nanoparticles	Development of mRNA vaccines for cancer immunotherapy	[80]
Chondroitin sulfate-based prodrug nanomaterials	Targeting Golgi apparatus to mediate photodynamic immunotherapy	[81]
Core-shell nanoparticles	Development of biocompatible nanocarriers with chlorin e6-polydopamine in cancer phototherapy	[82]
PEGylated iron oxide nanoparticle-loaded liposomes	Ferroptosis induction Delivery of doxorubicin and chemotherapy	[83]
Polydopamine-coated gold nanoparticles	ROS overgeneration in photodynamic therapy Apoptosis induction	[84]
Hydroxyapatite nanomaterials	Delivery of altretamine and apoptosis induction	[85]
Metal-organic frameworks	Modification of nanocomposites with GST-Afb to protect the surface and preventing the interactions in biological media	[86]
Worm-like nanoparticles	Reduction-responsive system Delivery of curcumin and also, stimulation of phototherapy by IR820 delivery	[87]
Gold nanoparticles	Gadolinium chelate-coated nanoparticles to impair metastasis of glioblastoma cells	[88]

5. Nanoparticles in regulation of glycolysis in cancer

Based on the previous discussions, it was revealed that glycolysis induction in tumor cells promotes their growth. Therefore, inhibition of glycolysis using pharmacological compounds or gene therapy methods can prevent carcinogenesis. Notably, nanoparticles have been emerged as new tools in glycolysis regulation. The delivery of zoledronic acid by nanoparticles impairs isoprenoid synthesis and prevents upregulation of HIF-1 α through modulation of Ras/ERK1/2 and finally, suppresses glycolysis enzymes and glucose flux in reducing tumorigenesis [89]. Such nanoparticles have been extensively used for treatment of various human cancers and they are not specific to a certain kind of tumor. A recent experiment has revealed that albumin nanostructures containing bevacizumab can suppress glycolysis and reduce metabolism in tumor in a higher capacity compared to drug alone in suppression of colorectal tumor [90]. Since biological aspect of glycolysis and molecular pathways regulating this mechanism have been identified, gene delivery by nanoparticles can be also beneficial. A study on breast tumor has mentioned that RNAi-loaded iron oxide nanocarriers can reduce MCT4 expression to disrupt glycolysis and mediate acidosis in cancer cells to enhance oxidative damage [91]. Notably, phototherapy of tumor can also direct the aim for regulation of glycolysis and tumor metabolism.

The polypyrrole nanostructures can deliver dl-Menthol and can function as a diclofenac nanoplatfrom stimulate photothermal therapy that suppresses glycolysis in cancer [92]. Importantly, the charge of cancer cells is distinct ogf normal cells. The stimulation of glycolysis can change the charge of cell membrane to negative charge and therefore, positively charged nanocarriers have been developed in cancer therapy [93]. The glycolysis regulation by nanoparticles can also improve cancer immunotherapy; the nanogels have been loaded with doxorubicin and after intracellular accumulation, such nanocarriers release doxorubicin in response to GSH and by impairing glycolysis metabolism, they reduce ATP generation. Moreover, they can stimulate immunogenic cell death and promote maturation of DC cells [94]. One of the regulators of glycolysis in cancer is HIF-1 α that increases RNF146 expression in glycolysis stimulation and elevating proliferation [95]. HIF-1 down-regulation by cryptolepine impairs glycolysis in breast cancer [96]. A recent experiment has focused on development of a nanococktail comprised of dendrimer and polymer that is redox-responsive and by reducing HIF-1 α suppression, it disrupts glycolysis in tumor cells [97]. **Table 2** summarizes the function of nanoparticles in glycolysis regulation.

Table 2: Affecting the metabolism of cancer cells through nanoparticles.

Nanoparticle	Mechanism of action	Reference
Zinc-Carnosine Metallodrug Network	Cu-sensitive drug release Inducing Cu depletion Suppressing glycolysis and oxidative phosphorylation	[98]
JX06 nanostructures	A combination of nanoparticles and metformin decrease PDK1 expression in disrupting metabolic plasticity	[99]
Liposome hybrid nanoparticles	Surface modification with cancer cell membrane pH-responsive release of siRNA and docetaxel Glycolysis inhibition	[100]
Liposomes	Loading EDTA and ICG in nanoparticles Deep tumor penetration and glycolysis inhibition	[101]
Iron oxide nanoparticles	Glycolysis mediates negative charge of cancer cells and positively charged nanocarriers suppress tumorigenesis	[102]
Self-assembled nanoparticles	Co-application of berberine and BACH1 inhibitor hermin in preparation of nanoparticles Glycolysis inhibition	[103]
BPTES nanoparticles	A combination of metformin and nanostructures suppress glutaminase After glutaminase inhibition, tumor cells rely on glycolysis for survival	[104]
MOF nanoparticles	Delivery of dihydroartemisinin and down-regulation of PI3K/Akt in glycolysis suppression	[105]
Two-in-One nanoformulation	Delivery of miR-125 and tyrosine kinase inhibitor in glycolysis inhibition	[106]
Bioresponsive nanotherapeutics	Apoptosis and autophagy induction Glycolysis inhibition Targeted delivery of quercetin	[107]

6. CircRNAs induce glycolysis mechanism

The ability of circRNAs in regulating glycolysis mechanism has resulted in a significant impact on progression of tumor cells. Due to two distinct functions of circRNAs including tumor-promoter and tumor-suppressor functions, circRNAs can induce/suppress glycolysis. Circ-DONSON promotes expression level of SOX4 transcription factor to elevate gastric cancer progression [108]. Furthermore, circ-0001829 enhances SMAD2 expression via miR-155-5p sponging to avoid apoptosis in gastric tumor [109]. Besides, circ-0023409 stimulates PI3K/Akt thereby increasing gastric cancer malignancy [110]. Circ-UBE2Q2 stimulates autophagy and glycolysis thereby enhancing gastric cancer progression and to this end, circ-UBE2Q2 triggers STAT3 signaling through miR-370-3p down-regulation [111]. The expression level of oncogenic circRNAs increase in gastric cancer. Circ-0000592 has been positively associated with growth, invasion, and glycolysis. Circ-0000592 suppresses miR-1179 to elevate ANXA4 expression thereby triggering glycolysis and promoting malignancy [112].

Another life-threatening tumor malignancy is bladder cancer. Development of bladder cancer and its malignancy depend on alterations in genetic and epigenetic factors as well as environmental exposures. Despite various therapeutics for bladder cancer, prognosis of patients remains poor [113]. Epigenetic factors are key players in the development of bladder cancer [114-116]. Circ-SEMA5A is an inducer of bladder cancer progression associated with triggering glycolysis. Circ-SEMA5A reduces miR-330-5p to elevate levels of ENO1 and SMEA5A as downstream targets to accelerate tumor progression partially via inducing glycolysis [117]. One of the factors that significantly enhances progression of bladder tumor is hypoxia in tumor microenvironment. Presence of hypoxia can lead to activation of autophagy in bladder cancer and trigger cisplatin resistance [118]. Notably, expression level of circRNAs in bladder cancer can be affected by hypoxia to enhance drug resistance [119]. Hypoxia can result in upregulation of LDHA as one of the enzymes of glycolysis to increase tumor progression. Hypoxia enhances expression level of circ-403658 through HIF-1 α -dependent manner and increases LDHA expression which leads to glycolysis induction and bladder tumor progression [120]. In addition to bladder tumor, prostate tumor is also a leading cause of death among men and is associated with a decrease in life quality of patients. Chemotherapy, radiotherapy, surgery, and targeted delivery systems are currently applied in treatment of prostate tumor [121, 122]. Similarly, ncRNAs modulate prostate tumor hallmarks such as proliferation and metastasis as well as response to therapy [123-125]. Circ-MID1 is involved in evoking glycolysis and elevating the progression. Upregulation of IGF1R and YTHDC2 is essential for carcinogenesis. Circ-MID1 enhances the expression level of IGF1R and YTHDC2 via miR-330-3p sponging which leads to glycolysis induction in prostate cancer [126]. Another malignancy of urogenital tract is renal cancer. Circ-VAMP3 can trigger glycolysis in renal cancer. The induction of glycolysis in renal cancer which enhances cancer growth through LDHA overexpression. Circ-VAMP3 is involved in inducing the phosphorylation of LDHA at tyrosine 10 and is mediated by interacting with FGFR1 [127].

Colorectal tumor is a common cancer of digestive system and is still a major healthcare problem worldwide [128]. Colorectal cancer has the third highest incidence rate among cancers and is the

second leading cause of death among patients [129]. Radiotherapy, surgery, chemotherapy, and adjuvant therapies are used for colorectal cancer suppression but prognosis remains poor [130]. Circ-PLCE1 accounts for increasing the progression and malignancy. Circ-PLCE1 functions as a sponge for miR-485-5p to upregulate ACTG1 which induces glycolysis, increases glucose uptake and metabolism, and promotes lactic acid and pyruvate production [131]. The upregulation of circRNAs such as circ-NOX4 induce glycolysis and mediate poor prognosis in colorectal tumor. Circ-NOX4 promotes progression and proliferation of colorectal via inducing glycolysis. To do this, circ-NOX4 promotes CDC28 level via miR-485-5p inhibition [132].

7. CircRNAs suppress glycolysis mechanism

In the previous section, the function of oncogenic circRNAs in inducing glycolysis and promoting malignancy of tumor cells was discussed. However, there are also several circRNAs that can suppress glycolysis and reduce carcinogenesis. Although the focus of experiments is on oncogenic circRNAs, this section aims in highlighting the function of tumor-suppressor circRNAs in glycolysis inhibition. CircRNF13 is an inhibitor of tumor progression with reduced expression levels in lung cancer. Restoring circRNF13 expression leads to sponging miR-93-5p which leads to its downregulation and interference in lung tumor progression [133]. In nasopharyngeal tumor, circRNF13 is considered as an inhibitor of glycolysis. At first, circRNF13 binds to 3'-UTR of *SUMO2* gene to increase protein levels of SUMO2. Then, SUMO2 enhances SUMOylation and degradation of GLUT1 thereby suppressing glycolysis [134]. Circ-0086414 is another pivotal player in tumors and its level is decreased in oral cancer [135]. Circ-0086414 disrupts tumorigenesis via inhibiting glycolysis. Circ-0086414 decreases miR-1290 expression via sponging and such reverse relationship between circ-0086414 and miR-1290 is necessary for upregulating SPARCL1 mediating glycolysis suppression and diminution in growth and metastasis of esophageal tumor [136].

One of the important regulators of glycolysis mechanism in cancer is PI3K/Akt. PER1 suppresses glycolysis mechanism in oral tumor via suppressing PI3K signaling [137]. Furthermore, PLOD1 induces PI3K/Akt signaling via SOX9 upregulation thereby triggering glycolysis and promoting gastric cancer progression [138]. On the other hand, increasing evidence suggests that PI3K/Akt signaling is an oncogenic pathway and a driver of ovarian cancer progression [139, 140]. PI3K/Akt signaling stimulates glycolysis to increase progression of ovarian cancer. However, circ-RHOBTB3 suppresses PI3K/Akt signaling thereby prevents glycolysis, minimizes proliferation and metastasis, and reduces ovarian cancer malignancy [141].

CircFADS2 has been considered as a driver of lung cancer progression via miR-498 sponging [142]. Furthermore, circFADS2 is used as a biomarker in colorectal cancer [143]. However, one of the complexities of circRNAs is their dual function in cancer. For instance, a previous experiment demonstrated the role of circFADS2 to increase lung cancer progression. However, another experiment shows that circFADS2 is an inhibitor of cutaneous cancer progression via glycolysis inhibition. CircFADS2 increases HOXA9 expression via miR-766-3p down-regulation thereby suppressing glycolysis-mediated cancer progression [144]. Notably, the capacity of circRNAs in regulating glycolysis and cancer progression has been confirmed *in vitro* and *in vivo*. Circ-0001777 down-regulates miR-942-5p expression to enhance PRICKLE2

expression, resulting in glycolysis inhibition and lung tumor reduction *in vitro* and *in vivo* [145]. Therefore, more studies are needed to delineate the role of circRNAs in suppressing glycolysis in cancer (**Figure 1**).

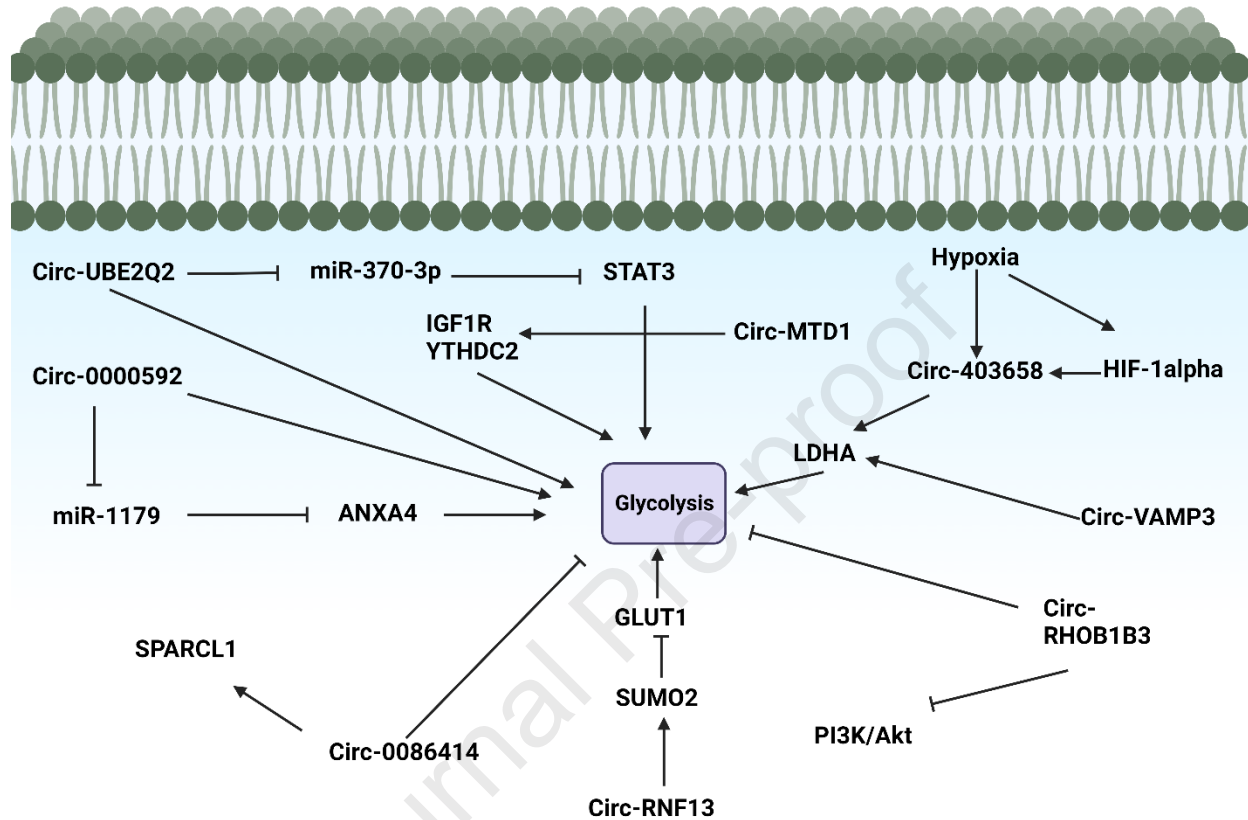


Figure 1: The role of CircRNAs in regulating glycolysis in cancer.

8. CircRNAs regulate drug resistance via glycolysis regulation

Accumulating data has shown that circRNAs are involved in two important aspects of cancer progression including metabolic reprogramming (e.g., Glycolysis) and mediating drug resistance. Circ-CUX1 can interact with EWSR1 to increase MAZ expression thereby triggering glycolysis and increasing proliferation [146]. Besides, circ-0008039 increases SKA2 expression via miR-140-3p sponging leading to glycolysis induction and tumorigenesis promotion in breast cancer [147]. Therefore, circRNAs are critical regulators of glycolysis in cancer [148]. On the other hand, increasing evidence has shown that circ-0,007,331 promotes ANLN expression via miR-200b-3p down-regulation to induce paclitaxel resistance in breast tumor [149]. CircNRP1 is suggested to be a key player in the process of developing resistance to 5-fluorouracil (5-FU) chemotherapy [150]. Although 5-FU is commonly used in treatment of gastric cancer, the development of hypoxia in tumor microenvironment can result in an increase in progression of gastric tumor due to activation of HIF-1 α signaling. After overexpression of HIF-1 α during hypoxia, glycolysis and glucose metabolism pave the way to increased proliferation of gastric tumor cells and 5-FU resistance. However, miR-138-5p is an inhibitor of HIF-1 α signaling

associated with suppression of gastric tumor progression. Notably, circNRIP1 stimulates HIF-1 α signaling via miR-138-5p sponging thereby triggering 5-FU resistance in gastric cancer [150]. Therefore, therapeutic targeting of circNRIP1/miR-138-5p/HIF-1 α axis is of importance in suppressing glycolysis and reversing 5-FU resistance in gastric cancer [150]. More importantly, 5-FU is utilized for the treatment of colorectal cancer and its anti-invasion activity is based on suppressing EMT mechanism [151]. However, overexpression of tumor-promoting factors and increase in stemness of colorectal cancer cells can result in development of 5-FU resistance [152]. On the other hand, glycolysis is implicated in increasing progression of colorectal cancer. LINRIS can increase stability of IGF2BP2 thereby triggering aerobic glycolysis and mediating colorectal cancer progression [153]. Besides, inhibition of glycolysis by fasting via suppression of Akt/mTOR/HIF-1 α axis results in significant reduction in growth of colorectal tumor cells [154]. CircSAMD4A has been implicated in developing 5-FU resistance in colorectal cancer. Upregulation of PFKFB3 induces glycolysis and increases proliferation of colorectal tumor cells thereby triggering 5-FU resistance. CircSAMD4A increases PFKFB3 expression via miR-545-3p sponging to induce glycolysis and 5-FU resistance in colorectal tumor [155].

Paclitaxel belongs to the family of taxanes and its mechanism of action in cancer therapy is based on disrupting the balance of microtubules via inhibiting depolymerization of microtubules to induce cell cycle arrest. Due to the development of paclitaxel resistance in cancer, nanoscale delivery systems have been contemplated on to increase paclitaxel potential for cancer suppression and reversing drug resistance [156]. Circ-RNF111 is involved in triggering paclitaxel resistance in breast tumor. miR-140-5p is capable of reducing expression level of E2F3 to inhibit paclitaxel insensitivity in breast tumor. However, circ-RNF111 promotes E2F3 expression via miR-140-5p down-regulation thereby promote progression and colony formation in breast cancer cells and induce paclitaxel resistance. Notably, the capacity of circ-RNF111 in developing paclitaxel resistance is partially mediated by glycolysis induction via affecting miR-140-5p/E2F3 axis [157]. Lung cancer is another malignant tumor in humans and non-small cell lung cancer (NSCLC) is the most prominent type [158, 159]. In spite of significant efforts in treatment of NSCLC, prognosis and survival rate of patients remain poor [160]. The gold standard of treatment for NSCLC is paclitaxel [161]. Upregulation of circ-0011298 in NSCLC can render paclitaxel resistance in tumor cells. miR-486-3p functions as a tumor-suppressor factor in NSCLC and increases paclitaxel sensitivity via CRABP2 inhibition. However, NSCLC cells show increased malignancy and glycolysis which reduce their sensitivity to paclitaxel chemotherapy. Circ-0011298 sponges miR-486-3p to upregulate CRABP2 thereby induce glycolysis and inhibit apoptosis to develop paclitaxel resistance in NSCLC cells [162].

miR-543 is a new emerging target in cancer therapy and accumulating data suggests its onco-suppressor function. Therefore, increasing miR-543 expression is key in cancer therapy. miR-543 reduces the expression levels of VCAN and UBE2T thereby suppress breast cancer progression [163, 164]. Besides, miR-543 induces apoptosis via suppressing STAT3 signaling in liver cancer [165]. However, a recent experiment has shown that miR-543 can induce cisplatin resistance in NSCLC. Notably, circ-FOXO3 reduces miR-543 expression to inhibit glycolysis and enhance cisplatin sensitivity of NSCLC cells [166]. CircRNAs induce glycolysis and prevent apoptosis to aid in developing drug resistance in cancer [167]. Therefore, future studies are

needed to investigate the regulation of apoptosis by circRNA/glycolysis axis in regulating therapy resistance in cancers.

Another important malignant tumor in humans is hepatocellular carcinoma. Hepatocellular carcinoma is a subtype of liver cancer and the second leading cause of cancer-related death [168, 169]. Despite advances in treatment of hepatocellular carcinoma, metastasis and recurrence are still major challenges [170, 171]. Furthermore, molecular pathways play a significant role in the progression of hepatocellular carcinoma [172-174]. Circ-UBE2D2 promotes the expression level of LDHA via reducing miR-889-3p which leads to glycolysis and sorafenib resistance [175]. In addition to regulating LDHA in glycolysis, circRNAs can also modulate expression level of HK2. Upregulation of HK2 results in glycolysis and mediates cisplatin resistance in NSCLC. Circ-0008928 promotes HK2 expression by miR-488 inhibition leading to glycolysis induction and cisplatin resistance in NSCLC [176]. More importantly, expression level of circRNAs can be regulated by astragalus IV in breast cancer. Astragalus IV is involved in reversing drug resistance in breast cancer. It promotes the expression levels of circ-0001982 which leads to down-regulation of miR-206/miR-613 to suppress drug resistance in breast cancer [177].

Small extracellular vesicles are known as exosomes. These minute structures are used for cell-to-cell communication. Bioactive molecules, RNA molecules, DNA, proteins, and lipids are among the factors that can be loaded in exosomes. Recently, much attention has been directed towards understanding the role of exosomes in cancer progression and therapy resistance [178-181]. Exosomal circRNAs have been considered as regulators of glycolysis that impact drug resistance. Exosomes can deliver circ-0005963 which induces glycolysis via increasing PKM2 expression. Additionally, exosomal circ-0005963 reduces miR-122 expression. Subsequently, PKM2 upregulation during glycolysis leads to drug resistance in colorectal cancer [182]. However, exosomal circ-0094343 increases drug sensitivity in colorectal cancer. Exosomal circ-0094343 promotes TRIM67 expression via miR-766-5p down-regulation to suppress glycolysis thereby promoting drug sensitivity in colorectal cancer [183].

miR-143 is a tumor-suppressor factor capable of inhibiting metastasis of tumor cells and impairing cancer invasion via EMT inhibition [184, 185]. CircRNAs are regulators of miR-143 in cancer progression [186, 187]. Down-regulation of miR-143 can induce glycolysis thereby trigger drug resistance in cancers. Exosomal circ-DLGAP4 can induce glycolysis and mediate chemoresistance in neuroblastoma. Exosomal circ-DLGAP4 increases HK2 expression by miR-143 down-regulation which triggers chemoresistance [188]. Therefore, exosomal circRNAs also regulate glycolysis in cancer therapy resistance [189]. One of the limitations of current studies is the lack of assessments of the role of circRNAs in modulating radio-resistance via glycolysis. **Table 3** summarizes circRNA/miRNA axis in cancer drug resistance.

Table 3: The role of circRNAs in regulating glycolysis and mediating drug resistance.

CircRNA	Molecular pathway	Cancer type	Drug	Remark	Ref
---------	-------------------	-------------	------	--------	-----

Circ-NRIP1	miR-138-5p/HIF-1 α	Gastric cancer	5-Flourouracil	miR-138-5p sponging by circ-NRIP1 Increase in HIF-1 α expression Glycolysis induction Drug resistance development	[150]
Circ-SAMD4A	miR-545-3p/PFKFB3	Colorectal cancer	5-Flourouracil	Increase in expression of PFKFB3 by circ-SAMD4A via miR-545-3p sponging Stimulating glycolysis and promoting drug resistance	[155]
Circ-RNF111	miR-140-5p/E2F3	Breast cancer	Paclitaxel	Circ-RNF111 increases E2F3 expression by miR-140-5p sponging in developing paclitaxel resistance	[157]
Circ-0011298	miR-486-3p/CRABP2	NSCLC	Taxol	Circ-0011298 increases CRABP2 expression by miR-486-3p down-regulation in taxol resistance	[162]
Circ-0080145	miR-326/PPFIA1	Leukemia	Imatinib	Circ-0080145 increases PPFIA1 expression via miR-326 down-regulation for inducing imatinib resistance	[167]
Circ-UBE2D2	miR-889-3p/LDHA	Hepatocellular carcinoma	Sorafenib	Circ-UBE2D2 promotes LDHA expression via miR-889-3p sponging for triggering drug resistance	[175]
Circ-0005963	miR-122/PKM2	Colorectal cancer	Oxaliplatin	Circ-0005963 increases PKM2 expression via miR-122 down-regulation for drug resistance development	[182]
Circ-0094343	miR-766-5p/TRIM67	Colorectal cancer	5-flourouracil	Circ-0094343 increases TRIM67 expression via miR-766-5p inhibition for glycolysis inhibition and mediating drug sensitivity	[183]

9. CircRNA/miRNA axis and glycolysis

Up to 75% of the human genome is transcribed to RNAs. There are two types of RNAs including protein-coding and non-coding RNAs (ncRNAs) [190]. microRNAs (miRNAs) are another type of ncRNAs with lengths of less than 24 nucleotides. MicroRNAs are pre-maturely produced in the nucleus and are transferred to the cytoplasm by exportin 5. In the cytoplasm, they undergo maturation and gain their function by loading into RNA-induced silencing complex (RISC) [191, 192]. miRNAs reduce the expression levels of target genes by binding to UTR regions to prevent translation or mediate degradation of mRNAs. Dysregulation of miRNAs is observed in cancers

and can increase/decrease tumor progression. Targeting miRNAs is key in regulating tumor malignancy and affecting cancer progression. miRNAs can regulate cancer hallmarks including proliferation, metastasis, and therapy resistance [193-196]. Furthermore, miRNAs have been considered as important regulators of glycolysis in cancer [197, 198]. CircRNA/miRNA axis can also regulate glycolysis in cancers. CircRNF20 is an inducer of Warburg effect in breast tumor. Upregulation of HIF-1 α can lead to an increase in progression of breast cancer and stimulation of glycolysis via enhancing expression level of HK2. However, miR-487a reduces HK2 expression by inhibiting HIF-1 α signaling thereby interfering with glycolysis in breast cancer. Notably, CircRNF20 stimulates HIF-1 α /HK2 axis by miR-487a sponging which triggers glycolysis and favors breast cancer progression [199]. Similarly, HIF-1 α signaling has been implicated in increasing the progression of gastric cancer. HIF-1 α upregulation by CTHRC1 is involved in increasing CXCR4 expression to promote invasion of gastric tumor cells [200]. Besides, PI3K/Akt signaling enhances HIF-1 α expression which aggravates gastric cancer progression [201]. miR-515-5p suppresses HIF-1 α signaling to inhibit glycolysis in gastric cancer. However, gastric tumor cells use molecular pathway switching which increases their progression via glycolysis induction. Gastric tumor cells increase expression level of circ-MAT2B which increases HIF-1 α expression via miR-515-5p sponging in cytoplasm leading to glycolysis-mediated gastric cancer progression [202].

miR-98 is another new emerging target in cancer which impairs tumor progression. Hence, restoring miR-98 expression is of importance for cancer therapy. miR-98 reduces the expression level of BCAT1 to reduce self-renewal capacity of cancer stem cells and avoid paclitaxel resistance in gastric cancer [203]. miR-98-5p reduces TGFBR1 and suppresses PI3K/Akt signaling which impair cancer proliferation and invasion [204, 205]. However, lung tumor cells prefer to reduce miR-98 expression in enhancing their progression. Circ-0006349 is involved in triggering glycolysis and promoting the progression of lung cancer. miR-98 reduces MKP1 expression which inhibits lung cancer progression. On the other hand, circ-0006349 increases MKP1 expression via miR-98 down-regulation leading to glycolysis induction and cancer progression [206]. Besides, circ-0091579 and CASC3 are increased in expression in hepatocellular carcinoma, while miR-490-5p is decreased. Knock-down of circ-0091579 is beneficial in preventing progression of tumor cells. Circ-0091579 sponges miR-490-5p to increase CASC3 expression. Then, glycolysis is induced and a significant increase is observed in progression of hepatocellular carcinoma cells [207].

The process of glycolysis is dependent on glucose delivery to cancer cells to be used as a valuable source of energy. Increasing evidence has shown that overexpression of GLUT1 as a cell surface transporter is vital for the process of cancer progression. S100A2 promotes GLUT1 expression to induce glycolysis in colorectal cancer [208]. SALL4 enhances GLUT1 expression via interacting with HP1 α to induce glycolysis [209]. Therefore, GLUT1 is an inducer of glycolysis and cancer acceleration. Circ-DENND4C is an inducer of glycolysis in colorectal tumors. Upregulation of GLUT1 has been considered as a pre-requisite of glycolysis induction in colorectal cancer. Circ-DENND4C promotes GLUT1 expression via miR-760 inhibition leading to glycolysis induction and cancer progression [210]. Therefore, glycolysis and proliferation of tumor cells are tightly regulated by circRNA/miRNA axis in cancers. For instance, circ-0001421

aggravates lung cancer progression by promoting CDCA3 expression via miR-4677-3p inhibition thereby triggering glycolysis and accelerating proliferation of lung tumor [211].

Hepatoma-derived growth factor (HDGF) is an oncogenic factor and its upregulation by ZEB1 can lead to increase in invasion of tumor cells [212]. Non-coding RNAs are regulators of HDGF which suppress carcinogenesis [213, 214]. Upregulation of HDGF can lead to glycolysis induction and lung tumor progression. Circ-MAGI3 is involved in facilitating glycolysis and increasing tumor progression in lung cancer. Circ-MAGI3 promotes HDGF expression via miR-515-5p sponging to increase lung cancer growth and glycolysis [215]. miR-106a is an oncogenic factor in different cancers, especially ovarian cancer [216, 217]. Upregulation of miR-106a can pave the way for growth and invasion of prostate tumor cells [218]. In a number of cancers such as colorectal tumor, miR-106a demonstrates tumor-suppressor activity [219] and modulates c-Jun signaling to impair cervical cancer progression [220]. Circ-ITCH is an inhibitor of ovarian cancer progression which reduces miR-106a expression to elevate CDH expression, resulting in glycolysis inhibition and reduction in progression of ovarian tumor cells [221]. Previously, it was mentioned that GLUT1 is involved in increasing the transportation of glucose into tumor cells and triggering glycolysis in cancer cells [222]. Notably, GLUT3 is also involved in the tumorigenesis process and its activation by YAP signaling promotes colorectal cancer progression [223]. Stimulation of tumor cells by IL-8 can lead to an increase in GLUT3 expression [224]. Besides, upregulation of GLUT3 increases glucose uptake and mediates drug resistance in cancer [225]. Circ-MYLK is involved in glycolysis via promoting glucose uptake in NSCLC cells. Silencing circ-MYLK impairs proliferation and glycolysis in NSCLC cells. Circ-MYLK enhances GLUT3 expression via miR-195-5p sponging thereby elevating glucose uptake and mediating glycolysis in tumor cells [226]. Based on these discussions, circRNAs are potential regulators of miRNAs that can impact glycolysis in tumor cells (**Figure 2, Table 4**).

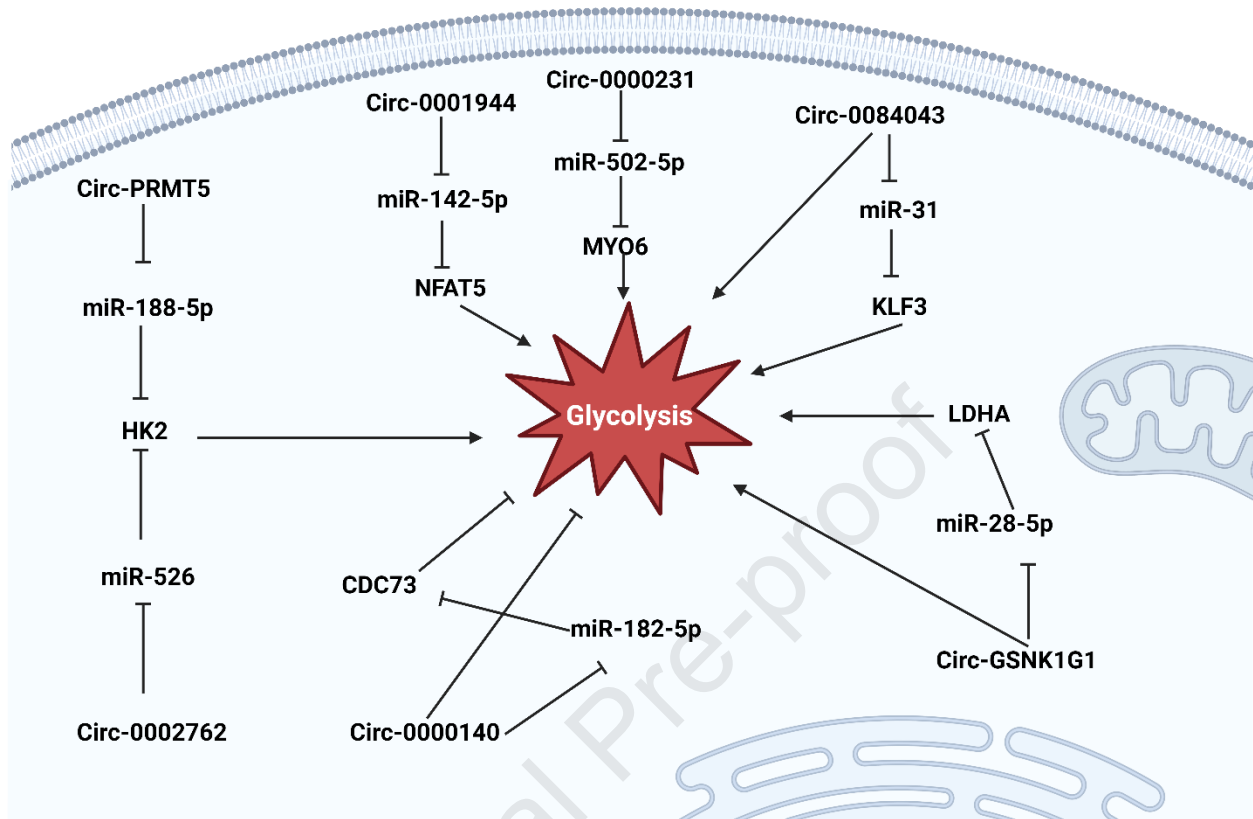


Figure 2: Role of CircRNA/miRNA axis in regulating glycolysis in cancer.

Table 4: An overview of circRNAs' role in regulating glycolysis in cancers.

CircRNA	Molecular pathway	Cancer type	Remark	Ref
Circ-0000231	miR-502-5p/MYO6	Colorectal cancer	Circ-0000231 increases MYO6 expression via miR-502-5p sponging for glycolysis induction	[227]
Circ-0001944	miR-142-5p/NFAT5	Lung cancer	Circ-0001944 increases NFAT5 expression via miR-142-5p inhibition for glycolysis induction	[228]
Circ-PRMT5	miR-188-5p/HK2	Hepatoma	Circ-PRMT5 increases HK2 expression by miR-188-5p down-regulation for glycolysis stimulation	[229]
Circ-0084043	miR-31/KLF3	Melanoma	Circ-0084043 increases KLF3 expression via miR-31 inhibition in increasing cancer progression Glycolysis induction	[230]
Circ-0002762	miR-526b-5p/HK2	Cervical cancer	Circ-0002762 increases HK2 expression via miR-526b-5p sponging for glycolysis induction	[231]

Circ-0000140	miR-182-5p/CDC73	Oral cancer	Circ-0000140 increases CDC73 expression via miR-182-5p sponging in glycolysis inhibition and preventing tumorigenesis	[232]
Circ-CSNK1G1	miR-28-5p/LDHA	Breast cancer	Circ-CSNK1G1 positively regulates LDHA expression via miR-28-5p down-regulation for glycolysis stimulation	[233]
Circ-0000735	miR-940/BMPER	Lung cancer	Circ-0000735 increases BMPER expression by miR-40 sponging for glycolysis induction	[234]
Circ-0009910	miR-361-3p/SNRPA	Gastric cancer	Circ-0009910 enhances SNRPA expression by miR-361-3p sponging Glycolysis induction Tumorigenesis acceleration	[235]
Circ-0000442	miR-1229-3p/ZBTB1	Breast cancer	Circ-0000442 promotes ZBTB1 expression via miR-1229-3p inhibition Suppressing carcinogenesis via glycolysis inhibition	[236]
Circ-0016760	miR-4295/E2F3	Lung cancer	Sponging miR-4295 by circ-0016760 Increasing E2F3 expression Inducing glycolysis	[237]
Circ-0047921	miR-1287-5p/LARP1	Lung cancer	Circ-0047921 increases LARP1 expression by miR-1287-5p sponging for carcinogenesis and glycolysis induction	[238]
Circ-0001982	miR-1287-5p/MUC19	Breast cancer	Circ-0001982 increases MUC19 expression via miR-1287-5p down-regulation for increasing cancer progression during hypoxia and inducing glycolysis	[239]
Circ-0099999	miR-330-5p/FSCN1	Pancreatic cancer	Circ-0099999 increases FSCN1 expression by miR-330-5p sponging for glycolysis induction and facilitating tumorigenesis	[240]
Circ-0136666	miR-383/CREB1	Colorectal cancer	Circ-0136666 promotes CREB1 expression via miR-383 sponging thereby triggering glycolysis and enhancing tumorigenesis	[241]
Circ-0039411	miR-423-5p/SOX4	Thyroid cancer	Circ-0039411 increases SOX4 expression via miR-423-5p sponging for glycolysis induction	[242]
Circ-0002346	miR-582-3p/STXBP6	Lung cancer	Circ-0002346 increases STXBP6 expression via miR-582-3p to suppress glycolysis	[243]

Circ-PITX1	miR-1248/CCND2	Lung cancer	Circ-PITX1 increases CCND2 expression via miR-1248 sponging leading to glycolysis induction	[244]
Circ-0078710	miR-431-5p/TXNDC5	Liver cancer	Circ-0078710 increases TXNDC5 expression by miR-431-5p inhibition Glycolysis induction Apoptosis inhibition	[245]
Circ-0072995	miR-149-5p/SHMT2	Breast cancer	Circ-0072995 increases SHMT2 expression by miR-149-5p down-regulation leading to glycolysis induction	[246]
Circ-TFF1	miR-338-3p/FGFR1	Breast cancer	Circ-TFF1 sponges miR-338-3p to increase FGFR1 expression Facilitating tumor progression via glycolysis induction	[247]
Circ-0001721	miR-372-3p/MAPK7	Osteosarcoma	Glycolysis induction Circ-0001721 promotes MAPK7 expression via miR-372-3p down-regulation	[248]
Circ-RPPH1	miR-328-3p/HMGA2	Breast cancer	Circ-RPPH1 increases HMGA2 expression via miR-328-3p sponging leading to glycolysis induction and increase in proliferation and invasion	[249]
Circ-0001495	miR-526b-3p/TMBIM6	Cervical cancer	Circ-0001495 induces glycolysis in cancer via increasing TMBIM6 expression through miR-526b-3p sponging	[250]
Circ-0026134	miR-3619-5p/CHAF1B	Lung cancer	Circ-0026134 increases CHAF1B expression via miR-3619-5p down-regulation to accelerate tumorigenesis via glycolysis induction	[251]
Circ-0000467	miR-330-5p/TYRO3	Colorectal cancer	Circ-0000467 sponges miR-330-5p to increase TYRO3 expression thereby triggering glycolysis	[252]
Circ-0045932	miR-873-5p/HK2	Colorectal cancer	Circ-0045932 sponges miR-873-5p in increasing HK2 expression and promoting tumorigenesis via glycolysis induction	[253]
Circ-0018289	miR-1294/ICMT	Cervical cancer	miR-1294 sponging by circ-0018289 to increase ICMT expression towards glycolysis induction	[254]
Circ-0001955	miR-1299/GLUT1	Breast cancer	Circ-0001955 decoys miR-1299 to increase GLUT1 expression thereby triggering glycolysis and increasing tumorigenesis	[255]

Circ-0008717	miR-217/FBX017	Renal cancer	Circ-0008717 enhances FBX017 expression via miR-217 sponging thereby glycolysis induction	[256]
Circ-0000376	miR-1182/NOVA2	Lung cancer	Circ-0000376 sponges miR-1182 to increase NOVA2 expression, stimulating glycolysis	[257]
Circ-0003221	miR-892b/DHCR24	Bladder cancer	Circ-0003221 increases DHCR24 expression via miR-892b sponging thereby triggering glycolysis and enhancing tumorigenesis	[258]

10. Anti-cancer agents regulating circRNA/glycolysis axis

One of the important aspects related to glycolysis is the ability of anti-tumor agents in impacting this mechanism. A question is if circRNAs can regulate glycolysis, can anti-tumor agents regulate glycolysis to affect tumorigenesis? Several studies have evaluated the role of anti-tumor agents in regulating glycolysis via affecting circRNAs. Bupivacaine is a local anesthetic agent that is commonly used in gastric cancer surgery [259]. Increasing evidence shows that bupivacaine is beneficial in cell death induction and limiting growth rate of cancer cells [260, 261]. Notably, bupivacaine is a regulator of circRNA expression for gastric cancer therapy. Bupivacaine decreases the expression level of circ-0000376 to pave the way for upregulation of miR-145-5p thereby suppressing glycolysis, increasing invasion of gastric tumor cells, triggering apoptosis, and reducing viability of cancer cells [262]. Propofol is another anesthetic agent that it is administered intravenously and is a short-acting agent effective for treating pathophysiological mechanisms of diseases [263, 264]. Propofol treatment of cancer is associated with STAT3 signaling suppression and induction of ferroptosis in tumor cells [265]. Furthermore, propofol inhibits SHH and PI3K/Akt molecular pathways thereby impairing cancer progression [266]. One of the important aspects related to propofol is its impact on circRNAs and regulation of glycolysis. Propofol administration results in a decrease in expression level of circ-ERBB2 which leads to upregulation of miR-7-5p. Subsequently, FOXM1 expression decreases in lung cancer, leading to glycolysis suppression [267].

Propofol has been increasingly used in the treatment of ovarian cancer. Propofol increases drug sensitivity of ovarian tumor cells and suppresses various molecular pathways such as STAT3 to impair ovarian cancer progression [268-270]. Propofol administration results in a decrease in expression level of circ-ZER. Then, expression level of miR-212-5p is enhanced to suppress SOD2, resulting in glycolysis inhibition in ovarian cancer [271]. The ability of propofol in decreasing tumor progression and suppressing glycolysis can be disrupted by increasing expression level of circ-MUC16 [272], suggesting that circRNAs are targets of propofol in cancer therapy. In addition, natural products are also promising candidates in treatment of cancer. Tanshinone I has been increasingly applied for the treatment of cancer and is capable of suppressing PI3K/Akt/mTOR axis in reducing ovarian cancer progression [273]. Furthermore, tanshinone I induces apoptosis and suppresses STAT3 signaling in cancer therapy [274, 275]. Tanshinone I has been beneficial in impairing the progression of osteosarcoma. For this purpose,

tanshinone I decreases circ-0000376 expression to upregulate miR-432-5p, resulting in Bcl-2 down-regulation and glycolysis and metastasis suppression [276]. Therefore, circRNA is a “druggable target” and its expression level can be regulated by anti-cancer agents in suppressing glycolysis for tumor treatment.

11. Conclusion and remarks

The tumor microenvironment is characterized by several features such as redox imbalance and low pH levels. The acidic pH of tumor microenvironment is because of high proliferation of cancer cells while colony formation can be attributed to glycolysis induction. There are several enzymes involved in glycolysis such as LDHA, HK2, and others, whose activity induces glycolysis and increase carcinogenesis. The regulation of glycolysis in cancer cells has been of interest and circRNAs are considered as potent modulators of glycolysis in cancer. Due to pleiotropic function of circRNAs and their capacity in modulating molecular pathways as well as their functions as epigenetic factors, increasing evidence has shown that aberrant expression of circRNAs can lead to dysregulation of glycolysis in cancer. The purpose of the current review is to show how circRNAs can affect glycolysis mechanism directly and indirectly in cancers. There are two types of circRNAs, those that induce glycolysis and those that suppress it. The circRNAs regulating glycolysis mechanism can be transferred in tumor microenvironment and among tumor cells via exosomes and therefore, modulating biogenesis of exosomes can control progression and glycolysis in cancer cells. The interaction of circRNAs with glycolysis can determine the response of tumor cells to chemotherapy. Furthermore, circRNAs primarily regulate glycolysis via affecting miRNAs. HK2, LDHA, and PKM2 are mainly regulated by circRNAs thereby affecting glycolysis. HMGA2, SOX4, and CREB1 are among the downstream targets indirectly affected by circRNAs in regulating glycolysis in cancer. More importantly, propofol, tanshinone I, and bupivacaine have been used as regulators of circRNA expression in suppressing glycolysis-mediated cancer progression.

Although significant efforts have been made in revealing the potential of circRNAs and downstream targets in glycolysis regulation, there are several gaps and limitations that should be considered in future studies. The first gap is that there is no experiment on the role of circRNA/glycolysis axis in radio-resistance development in cancer and since radiotherapy is one of the major arms in treatment of cancer, future studies should focus on this aspect. Additionally, two key entities, i.e., exosomal circRNA and tumor-suppressor circRNAs have been ignored for their role in regulating of glycolysis in cancer. Although there are experiments related to them, more studies on their role in tumor progression regulation are required. Another limitation is that only the efficacy of three anti-cancer agents including propofol, bupivacaine, and tanshinone I in regulating circRNA/glycolysis axis in cancer has been investigated and more anti-cancer agents need to be tested. Another major gap is the lack of studies related to using nanostructures for delivery of anti-tumor agents or genetic tools to manipulate levels of circRNAs and targeted therapy of cancer. Future studies can translate these findings into clinic for treatment of patients.

Although the molecular profile of glycolysis and its regulation by circRNA have been understood with details, it is vital to use agents for targeting glycolysis in human cancers. The section 10 of current paper displayed application of pharmacological compounds and anti-cancer

drugs in glycolysis suppression for tumor therapy, but they lack specific accumulation at cancer site. Therefore, nanoparticles for specific targeting and removal of tumor cells have been developed. In section 4, it was discussed that nanoparticles can suppress growth and metastasis of tumor cells. Nanoparticles can impair immunosuppression and induce tumor microenvironment remodeling. Nanoparticles are used for vaccine development and they can deliver both drugs and genes in cancer therapy. Moreover, nanoparticles induce phototherapy in cancer removal. Due to progresses in field of biology, nanoengineering and their combination, there have been efforts that focused on nanoparticle application in glycolysis inhibition. In section 5, it was discussed that nanoparticles can suppress pathways related to glycolysis such as HIF-1 α and enzymes of glycolysis pathway such as HK2. Moreover, nanoparticles can deliver drugs and genes suppressing glycolysis in cancer therapy. Therefore, targeted regulation of glycolysis in cancer therapy has been conducted and in order to increase specific targeting of cancer cells, the functionalization of nanoparticles with ligands has been conducted. Moreover, nanoparticles can mediate phototherapy for preventing glycolysis in cancer. However, we are still at the beginning way of using nanoparticles in cancer therapy and more advanced experiments regulating glycolysis in cancer should be conducted. Furthermore, the role of circRNA/glycolysis axis in human cancers was discussed properly and an important part is capacity of nanoparticles for delivery of circRNAs in cancer therapy [277]. Therefore, use of circRNA-loaded nanoparticles in glycolysis suppression in human cancers can be examined in future studies.

Conflict of interest:

The authors declare no conflict of interest.

Acknowledgement:

The figures were depicted by Biorender and authors are grateful for the helps of this software (Biorender.com).

Funding:

The authors express their gratitude to the Deanship of Scientific Research at King Khalid University for funding this work through the Large Research Group Project under grant number RGP.02/216/43.

References:

1. Zhou, J., et al., *Circular RNAs as novel rising stars with huge potentials in development and disease*. 2018. **22**(4): p. 597-610.
2. Capel, B., et al., *Circular transcripts of the testis-determining gene Sry in adult mouse testis*. 1993. **73**(5): p. 1019-1030.
3. Nigro, J.M., et al., *Scrambled exons*. 1991. **64**(3): p. 607-613.
4. Yuan, Y., et al., *The emerging roles of circular RNA-mediated autophagy in tumorigenesis and cancer progression*. *Cell Death Discov*, 2022. **8**(1): p. 385.
5. Guan, S., et al., *Circular RNA WHSC1 exerts oncogenic properties by regulating miR-7/TAB2 in lung cancer*. *J Cell Mol Med*, 2021. **25**(20): p. 9784-9795.
6. Chen, L.-L. and L.J.R.b. Yang, *Regulation of circRNA biogenesis*. 2015. **12**(4): p. 381-388.
7. Jin, X., et al., *CircRNA expression pattern and circRNA-miRNA-mRNA network in the pathogenesis of nonalcoholic steatohepatitis*. 2016. **7**(41): p. 66455.
8. Liang, D., J.E.J.G. Wilusz, and development, *Short intronic repeat sequences facilitate circular RNA production*. 2014. **28**(20): p. 2233-2247.
9. Jeck, W.R., et al., *Circular RNAs are abundant, conserved, and associated with ALU repeats*. 2013. **19**(2): p. 141-157.
10. Memczak, S., et al., *Circular RNAs are a large class of animal RNAs with regulatory potency*. 2013. **495**(7441): p. 333-338.
11. St-Pierre, P., et al., *Characterization of the siRNAs associated with peach latent mosaic viroid infection*. 2009. **383**(2): p. 178-182.
12. Lu, D. and A.-D.J.F.i.g. Xu, *Mini review: circular RNAs as potential clinical biomarkers for disorders in the central nervous system*. 2016. **7**: p. 53.
13. Zhang, H.-d., et al., *CircRNA: a novel type of biomarker for cancer*. 2018. **25**(1): p. 1-7.
14. Han, Y.-N., et al., *Circular RNAs: A novel type of biomarker and genetic tools in cancer*. 2017. **8**(38): p. 64551.
15. Zuo, L., et al., *Circulating Circular RNAs as Biomarkers for the Diagnosis and Prediction of Outcomes in Acute Ischemic Stroke*. *Stroke*, 2020. **51**(1): p. 319-323.
16. Wang, H.Y., et al., *Circular RNA is a popular molecule in tumors of the digestive system (Review)*. *Int J Oncol*, 2020. **57**(1): p. 21-42.
17. Jeck, W.R., et al., *Circular RNAs are abundant, conserved, and associated with ALU repeats*. *Rna*, 2013. **19**(2): p. 141-57.
18. Zhang, Y., et al., *Circular intronic long noncoding RNAs*. *Mol Cell*, 2013. **51**(6): p. 792-806.
19. Zhang, X.-O., et al., *Complementary sequence-mediated exon circularization*. 2014. **159**(1): p. 134-147.
20. Hsiao, K.Y., H.S. Sun, and S.J. Tsai, *Circular RNA - New member of noncoding RNA with novel functions*. *Exp Biol Med (Maywood)*, 2017. **242**(11): p. 1136-1141.
21. Vasilescu, C., et al., *From mobility to crosstalk. A model of intracellular miRNAs motion may explain the RNAs interaction mechanism on the basis of target subcellular localization*. 2016. **280**: p. 50-61.
22. Hansen, T.B., et al., *Natural RNA circles function as efficient microRNA sponges*. 2013. **495**(7441): p. 384-388.
23. Hansen, T.B., et al., *miRNA-dependent gene silencing involving Ago2-mediated cleavage of a circular antisense RNA*. 2011. **30**(21): p. 4414-4422.
24. Tran, A.M., et al., *A New World of Biomarkers and Therapeutics for Female Reproductive System and Breast Cancers: Circular RNAs*. *Front Cell Dev Biol*, 2020. **8**: p. 50.

25. Wen, J., et al., *Circular RNA HIPK3: A Key Circular RNA in a Variety of Human Cancers*. Front Oncol, 2020. **10**: p. 773.
26. Chen, Q., et al., *Biological functions, mechanisms, and clinical significance of circular RNA in pancreatic cancer: a promising rising star*. Cell Biosci, 2022. **12**(1): p. 97.
27. Hong, J., et al., *F. nucleatum targets lncRNA ENO1-IT1 to promote glycolysis and oncogenesis in colorectal cancer*. Gut, 2021. **70**(11): p. 2123-2137.
28. Guo, X., et al., *Circular RNA circSEPT9 Is Upregulated in Endometrial Cancer and Promotes Cell Invasion and Migration by Downregulating miR-186 through Methylation*. Ann Clin Lab Sci, 2022. **52**(3): p. 399-405.
29. Tao, W., et al., *Circular RNA circCPA4 promotes tumorigenesis by regulating miR-214-3p/TGIF2 in lung cancer*. Thorac Cancer, 2021. **12**(24): p. 3356-3369.
30. Yang, B.L., et al., *Circular RNA CUL2 regulates the development of colorectal cancer by modulating apoptosis and autophagy via miR-208a-3p/PPP6C*. Aging (Albany NY), 2022. **14**(1): p. 497-508.
31. Wang, J., et al., *Circular RNA circCSPP1 promotes the occurrence and development of colon cancer by sponging miR-431 and regulating ROCK1 and ZEB1*. J Transl Med, 2022. **20**(1): p. 58.
32. Kong, S., et al., *Circular RNA circPFKP suppresses the proliferation and metastasis of gastric cancer cell via sponging miR-644 and regulating ADAMTSL5 expression*. Bioengineered, 2022. **13**(5): p. 12326-12337.
33. Zhang, X., et al., *Circular RNA LONP2 regulates proliferation, invasion, and apoptosis of bladder cancer cells by sponging microRNA-584-5p*. Bioengineered, 2022. **13**(4): p. 8823-8835.
34. Huang, Y., et al., *Circular RNA circRPPH1 promotes breast cancer progression via circRPPH1-miR-512-5p-STAT1 axis*. Cell Death Discov, 2021. **7**(1): p. 376.
35. Liang, X., et al., *Circular RNA circRAB31 acts as a miR-885-5psponge to suppress gastric cancer progression via the PTEN/PI3K/AKT pathway*. Mol Ther Oncolytics, 2021. **23**: p. 501-514.
36. Yu, Z., et al., *Circular RNA hsa_circ_0002360 Promotes Proliferation and Invasion and Inhibits Oxidative Stress in Gastric Cancer by Sponging miR-629-3p and Regulating the PDLIM4 Expression*. Oxid Med Cell Longev, 2022. **2022**: p. 2775433.
37. Zhou, X., et al., *Circular RNA_0006014 promotes breast cancer progression through sponging miR-885-3p to regulate NTRK2 and PIK3/AKT pathway*. Aging (Albany NY), 2022. **14**(7): p. 3105-3128.
38. Pavlides, S., et al., *Warburg meets autophagy: cancer-associated fibroblasts accelerate tumor growth and metastasis via oxidative stress, mitophagy, and aerobic glycolysis*. Antioxid Redox Signal, 2012. **16**(11): p. 1264-84.
39. Chu, Y., et al., *Regulation of Autophagy by Glycolysis in Cancer*. Cancer Manag Res, 2020. **12**: p. 13259-13271.
40. Warburg, O.J.S., *On the origin of cancer cells*. 1956. **123**(3191): p. 309-314.
41. DeBerardinis, R.J., et al., *The biology of cancer: metabolic reprogramming fuels cell growth and proliferation*. Cell Metab, 2008. **7**(1): p. 11-20.
42. Kroemer, G. and J. Pouyssegur, *Tumor cell metabolism: cancer's Achilles' heel*. Cancer Cell, 2008. **13**(6): p. 472-82.
43. Vander Heiden, M.G., L.C. Cantley, and C.B. Thompson, *Understanding the Warburg effect: the metabolic requirements of cell proliferation*. Science, 2009. **324**(5930): p. 1029-33.
44. Feron, O., *Pyruvate into lactate and back: from the Warburg effect to symbiotic energy fuel exchange in cancer cells*. Radiother Oncol, 2009. **92**(3): p. 329-33.
45. Ganapathy-Kanniappan, S., *Taming Tumor Glycolysis and Potential Implications for Immunotherapy*. Front Oncol, 2017. **7**: p. 36.

46. Neugent, M.L., et al., *A New Perspective on the Heterogeneity of Cancer Glycolysis*. Biomol Ther (Seoul), 2018. **26**(1): p. 10-18.
47. Wang, J., et al., *Therapeutic targeting of SPIB/SPI1-facilitated interplay of cancer cells and neutrophils inhibits aerobic glycolysis and cancer progression*. Clin Transl Med, 2021. **11**(11): p. e588.
48. Zhang, P., Z. Li, and G. Yang, *Silencing of ISLR inhibits tumour progression and glycolysis by inactivating the IL-6/JAK/STAT3 pathway in non-small cell lung cancer*. Int J Mol Med, 2021. **48**(6).
49. Zhang, G., X. Zhao, and W. Liu, *NEDD4L inhibits glycolysis and proliferation of cancer cells in oral squamous cell carcinoma by inducing ENO1 ubiquitination and degradation*. Cancer Biol Ther, 2022. **23**(1): p. 243-253.
50. Fang, C., et al., *α -Hederin inhibits the growth of lung cancer A549 cells in vitro and in vivo by decreasing SIRT6 dependent glycolysis*. Pharm Biol, 2021. **59**(1): p. 11-20.
51. Zheng, C., et al., *Targeting PFKL with penfluridol inhibits glycolysis and suppresses esophageal cancer tumorigenesis in an AMPK/FOXO3a/BIM-dependent manner*. Acta Pharm Sin B, 2022. **12**(3): p. 1271-1287.
52. Qi, C.L., et al., *The IRF2/CENP-N/AKT signaling axis promotes proliferation, cell cycling and apoptosis resistance in nasopharyngeal carcinoma cells by increasing aerobic glycolysis*. J Exp Clin Cancer Res, 2021. **40**(1): p. 390.
53. Lin, J., et al., *The POU2F1-ALDOA axis promotes the proliferation and chemoresistance of colon cancer cells by enhancing glycolysis and the pentose phosphate pathway activity*. Oncogene, 2022. **41**(7): p. 1024-1039.
54. Huang, J., et al., *A feedback circuit comprising EHD1 and 14-3-3 ζ sustains β -catenin/c-Myc-mediated aerobic glycolysis and proliferation in non-small cell lung cancer*. Cancer Lett, 2021. **520**: p. 12-25.
55. Yang, Z., et al., *SETD5 Regulates Glycolysis in Breast Cancer Stem-Like Cells and Fuels Tumor Growth*. Am J Pathol, 2022. **192**(4): p. 712-721.
56. Hiatt, R.A. and N. Beyeler, *Cancer and climate change*. The Lancet Oncology, 2020. **21**(11): p. e519-e527.
57. Hoegh-Guldberg, O., et al., *Impacts of 1.5 C global warming on natural and human systems*. Global warming of 1.5° C., 2018.
58. Watts, N., et al., *The 2019 report of The Lancet Countdown on health and climate change: ensuring that the health of a child born today is not defined by a changing climate*. The Lancet, 2019. **394**(10211): p. 1836-1878.
59. Torre, L.A., et al., *Global Cancer Incidence and Mortality Rates and Trends—An Update*. Global Cancer Rates and Trends—An Update. Cancer epidemiology, biomarkers & prevention, 2016. **25**(1): p. 16-27.
60. Tang, Q., et al., *Genetically Engineering Cell Membrane-Coated BTO Nanoparticles for MMP2-Activated Piezocatalysis-Immunotherapy*. Adv Mater, 2023: p. e2300964.
61. Podojil, J.R., et al., *Biodegradable nanoparticles induce cGAS/STING-dependent reprogramming of myeloid cells to promote tumor immunotherapy*. Front Immunol, 2022. **13**: p. 887649.
62. Wang, N., et al., *Vaccination of TLR7/8 Agonist-Conjugated Antigen Nanoparticles for Cancer Immunotherapy*. Adv Healthc Mater, 2023: p. e2300249.
63. Chen, G., et al., *Immunomodulatory roles of selenium nanoparticles: Novel arts for potential immunotherapy strategy development*. Front Immunol, 2022. **13**: p. 956181.
64. Abdellatif, A.A.H., et al., *Silver Nanoparticles Stabilized by Poly (Vinyl Pyrrolidone) with Potential Anticancer Activity towards Prostate Cancer*. Bioinorg Chem Appl, 2022. **2022**: p. 6181448.

65. Amiryaghoubi, N., et al., *Smart chitosan-folate hybrid magnetic nanoparticles for targeted delivery of doxorubicin to osteosarcoma cells*. Colloids Surf B Biointerfaces, 2022. **220**: p. 112911.
66. Guo, B., et al., *Cuproptosis Induced by ROS Responsive Nanoparticles with Elesclomol and Copper Combined with α PD-L1 for Enhanced Cancer Immunotherapy*. Adv Mater, 2023: p. e2212267.
67. Karimi-Maleh, H., et al., *Guanine-Based DNA Biosensor Amplified with Pt/SWCNTs Nanocomposite as Analytical Tool for Nanomolar Determination of Daunorubicin as an Anticancer Drug: A Docking/Experimental Investigation*. Industrial & Engineering Chemistry Research, 2021. **60**(2): p. 816-823.
68. Karimi-Maleh, H., et al., *A green and sensitive guanine-based DNA biosensor for idarubicin anticancer monitoring in biological samples: A simple and fast strategy for control of health quality in chemotherapy procedure confirmed by docking investigation*. Chemosphere, 2022. **291**: p. 132928.
69. Gu, D., et al., *Size-Controllable DNA Origami-Stacked Gold Nanoparticles for Deep Tumor-Penetrating Therapy*. ACS Appl Mater Interfaces, 2022. **14**(33): p. 38048-38055.
70. Manayia, A.H., et al., *Photoreactive Mercury-Containing Metallosupramolecular Nanoparticles with Tailorable Properties That Promote Enhanced Cellular Uptake for Effective Cancer Chemotherapy*. Biomacromolecules, 2023. **24**(2): p. 943-956.
71. An, J., et al., *Naphthofluorescein-based organic nanoparticles with superior stability for near-infrared photothermal therapy*. Nanoscale, 2022. **14**(28): p. 10051-10059.
72. Zhang, M. and H. Xu, *Peptide-assembled nanoparticles targeting tumor cells and tumor microenvironment for cancer therapy*. Front Chem, 2023. **11**: p. 1115495.
73. Qaddoori, M.H. and H.S. Al-Shmgani, *Galangin-Loaded Gold Nanoparticles: Molecular Mechanisms of Antiangiogenesis Properties in Breast Cancer*. Int J Breast Cancer, 2023. **2023**: p. 3251211.
74. Choi, M.J., et al., *Tumor-Targeted Erythrocyte Membrane Nanoparticles for Theranostics of Triple-Negative Breast Cancer*. Pharmaceutics, 2023. **15**(2).
75. Chang, H., et al., *In vivo toxicity evaluation of tumor targeted glycol chitosan nanoparticles in healthy mice: repeated high-dose of glycol chitosan nanoparticles potentially induce cardiotoxicity*. J Nanobiotechnology, 2023. **21**(1): p. 82.
76. Du, B., et al., *J-aggregates albumin-based NIR-II fluorescent dye nanoparticles for cancer phototheranostics*. Mater Today Bio, 2022. **16**: p. 100366.
77. Mudigunda, S.V., et al., *Bioactive Polymeric Nanoparticles of Moringa oleifera Induced Phyto-Photothermal Sensitization for the Enhanced Therapy of Retinoblastoma*. Pharmaceutics, 2023. **15**(2).
78. Zhang, Y., et al., *Cu(2+)-Chelating Mesoporous Silica Nanoparticles for Synergistic Chemotherapy/Chemodynamic Therapy*. Pharmaceutics, 2022. **14**(6).
79. Mochizuki, C., et al., *Surface Functionalization of Organosilica Nanoparticles With Au Nanoparticles Inhibits Cell Proliferation and Induces Cell Death in 4T1 Mouse Mammary Tumor Cells for DNA and Mitochondrial-Synergized Damage in Radiotherapy*. Front Chem, 2022. **10**: p. 907642.
80. Sasaki, K., et al., *mRNA-Loaded Lipid Nanoparticles Targeting Dendritic Cells for Cancer Immunotherapy*. Pharmaceutics, 2022. **14**(8).
81. Li, H., et al., *Chondroitin sulfate-based prodrug nanoparticles enhance photodynamic immunotherapy via Golgi apparatus targeting*. Acta Biomater, 2022. **146**: p. 357-369.
82. Chen, S., et al., *Highly biocompatible chlorin e6-poly(dopamine) core-shell nanoparticles for enhanced cancer phototherapy*. Nanoscale Adv, 2022. **4**(21): p. 4617-4627.

83. Liu, Y., et al., *Liposomes embedded with PEGylated iron oxide nanoparticles enable ferroptosis and combination therapy in cancer*. Natl Sci Rev, 2023. **10**(1): p. nwac167.
84. Lee, M.S., et al., *Dual irradiation-triggered anticancer therapeutics composed of polydopamine-coated gold nanoparticles*. Biomater Adv, 2022. **136**: p. 212779.
85. Alghazwani, Y., et al., *The Combined Anti-Tumor Efficacy of Bioactive Hydroxyapatite Nanoparticles Loaded with Altretamine*. Pharmaceutics, 2023. **15**(1).
86. Oh, J.Y., et al., *Protein-Precoated Surface of Metal-Organic Framework Nanoparticles for Targeted Delivery*. Small, 2023: p. e2300218.
87. Hu, H., et al., *Reduction-responsive worm-like nanoparticles for synergistic cancer chemophotodynamic therapy*. Mater Today Bio, 2023. **18**: p. 100542.
88. Durand, M., et al., *Radiosensitization with Gadolinium Chelate-Coated Gold Nanoparticles Prevents Aggressiveness and Invasiveness in Glioblastoma*. Int J Nanomedicine, 2023. **18**: p. 243-261.
89. Kopecka, J., et al., *Self-assembling nanoparticles encapsulating zoledronic acid revert multidrug resistance in cancer cells*. Oncotarget, 2015. **6**(31): p. 31461-78.
90. Luis de Redín, I., et al., *In vivo efficacy of bevacizumab-loaded albumin nanoparticles in the treatment of colorectal cancer*. Drug Deliv Transl Res, 2020. **10**(3): p. 635-645.
91. Liu, Y., et al., *Engineering Multifunctional RNAi Nanomedicine To Concurrently Target Cancer Hallmarks for Combinatorial Therapy*. Angew Chem Int Ed Engl, 2018. **57**(6): p. 1510-1513.
92. Ma, Y., et al., *dl-Menthol Loaded Polypyrrole Nanoparticles as a Controlled Diclofenac Delivery Platform for Sensitizing Cancer Cells to Photothermal Therapy*. ACS Appl Bio Mater, 2019. **2**(2): p. 848-855.
93. Deng, Z., et al., *Dual Targeting with Cell Surface Electrical Charge and Folic Acid via Superparamagnetic Fe(3)O(4)@Cu(2-x)S for Photothermal Cancer Cell Killing*. Cancers (Basel), 2021. **13**(21).
94. Ma, X., et al., *Bioresponsive immune-booster-based prodrug nanogel for cancer immunotherapy*. Acta Pharm Sin B, 2022. **12**(1): p. 451-466.
95. Shen, G., et al., *HIF-1/2 α -Activated RNF146 Enhances the Proliferation and Glycolysis of Hepatocellular Carcinoma Cells via the PTEN/AKT/mTOR Pathway*. Front Cell Dev Biol, 2022. **10**: p. 893888.
96. Zheng, Z., et al., *Cryptolepine suppresses breast adenocarcinoma via inhibition of HIF-1 mediated glycolysis*. Biomed Pharmacother, 2022. **153**: p. 113319.
97. Huang, J., et al., *Targeted Drug/Gene/Photodynamic Therapy via a Stimuli-Responsive Dendritic-Polymer-Based Nanococktail for Treatment of EGFR-TKI-Resistant Non-Small-Cell Lung Cancer*. Adv Mater, 2022. **34**(27): p. e2201516.
98. Lei, L., et al., *Zinc-Carnosine Metallodrug Network as Dual Metabolism Inhibitor Overcoming Metabolic Reprogramming for Efficient Cancer Therapy*. Nano Lett, 2023.
99. Yang, X., et al., *Targeting Cancer Metabolism Plasticity with JX06 Nanoparticles via Inhibiting PDK1 Combined with Metformin for Endometrial Cancer Patients with Diabetes*. Adv Sci (Weinh), 2022. **9**(8): p. e2104472.
100. Zhang, W., et al., *Tumor microenvironment-activated cancer cell membrane-liposome hybrid nanoparticle-mediated synergistic metabolic therapy and chemotherapy for non-small cell lung cancer*. J Nanobiotechnology, 2021. **19**(1): p. 339.
101. Zhou, J., et al., *A deep tumor penetration nanoplatform for glycolysis inhibition and antimetastasis of breast cancer*. J Mater Chem B, 2022. **10**(22): p. 4306-4320.
102. Han, X., et al., *Biomarkerless targeting and photothermal cancer cell killing by surface-electrically-charged superparamagnetic Fe(3)O(4) composite nanoparticles*. Nanoscale, 2017. **9**(4): p. 1457-1465.

103. Yang, X., et al., *Coordinated regulation of BACH1 and mitochondrial metabolism through tumor-targeted self-assembled nanoparticles for effective triple negative breast cancer combination therapy*. Acta Pharm Sin B, 2022. **12**(10): p. 3934-3951.
104. Elgogary, A., et al., *Combination therapy with BPTES nanoparticles and metformin targets the metabolic heterogeneity of pancreatic cancer*. Proc Natl Acad Sci U S A, 2016. **113**(36): p. E5328-36.
105. Li, Y., et al., *MOF nanoparticles with encapsulated dihydroartemisinin as a controlled drug delivery system for enhanced cancer therapy and mechanism analysis*. J Mater Chem B, 2020. **8**(33): p. 7382-7389.
106. Lo, Y.L., et al., *Two-in-One Nanoparticle Formulation to Deliver a Tyrosine Kinase Inhibitor and microRNA for Targeting Metabolic Reprogramming and Mitochondrial Dysfunction in Gastric Cancer*. Pharmaceutics, 2022. **14**(9).
107. Zhang, X., et al., *Inhibition of growth and lung metastasis of breast cancer by tumor-homing triple-bioresponsive nanotherapeutics*. J Control Release, 2020. **328**: p. 454-469.
108. Ding, L., et al., *Circular RNA circ-DONSON facilitates gastric cancer growth and invasion via NURF complex dependent activation of transcription factor SOX4*. Mol Cancer, 2019. **18**(1): p. 45.
109. Niu, Q., et al., *Circular RNA hsa_circ_0001829 promotes gastric cancer progression through miR-155-5p/SMAD2 axis*. J Exp Clin Cancer Res, 2020. **39**(1): p. 280.
110. Li, J., et al., *hsa_circ_0023409 Accelerates Gastric Cancer Cell Growth and Metastasis Through Regulating the IRS4/PI3K/AKT Pathway*. Cell Transplant, 2021. **30**: p. 963689720975390.
111. Yang, J., et al., *Circular RNA UBE2Q2 promotes malignant progression of gastric cancer by regulating signal transducer and activator of transcription 3-mediated autophagy and glycolysis*. Cell Death Dis, 2021. **12**(10): p. 910.
112. Ji, Z., W. Diao, and J. Shang, *Circular RNA circ_0000592 elevates ANXA4 expression via sponging miR-1179 to facilitate tumor progression in gastric cancer*. Anticancer Drugs, 2022. **33**(1): p. e644-e654.
113. Ashrafizadeh, M., et al., *(Nano) platforms in bladder cancer therapy: Challenges and opportunities*. p. e10353.
114. Bi, J., et al., *Circ-BPTF promotes bladder cancer progression and recurrence through the miR-31-5p/RAB27A axis*. Aging (Albany NY), 2018. **10**(8): p. 1964-1976.
115. Cao, H.L., et al., *lncRNA-RMRP promotes proliferation, migration and invasion of bladder cancer via miR-206*. Eur Rev Med Pharmacol Sci, 2019. **23**(3): p. 1012-1021.
116. Ren, W., et al., *miR-616-5p Promotes Invasion and Migration of Bladder Cancer via Downregulating NR2C2 Expression*. Front Oncol, 2021. **11**: p. 762946.
117. Wang, L., et al., *Circular RNA circSEMA5A promotes bladder cancer progression by upregulating ENO1 and SEMA5A expression*. Aging (Albany NY), 2020. **12**(21): p. 21674-21686.
118. Mao, X., et al., *Hypoxia-Induced Autophagy Enhances Cisplatin Resistance in Human Bladder Cancer Cells by Targeting Hypoxia-Inducible Factor-1 α* . J Immunol Res, 2021. **2021**: p. 8887437.
119. Su, Y., et al., *Hypoxia-elevated circELP3 contributes to bladder cancer progression and cisplatin resistance*. Int J Biol Sci, 2019. **15**(2): p. 441-452.
120. Wei, Y., et al., *Hypoxia-induced circular RNA has_circRNA_403658 promotes bladder cancer cell growth through activation of LDHA*. Am J Transl Res, 2019. **11**(11): p. 6838-6849.
121. Ashrafizadeh, M., et al., *Targeting autophagy in prostate cancer: preclinical and clinical evidence for therapeutic response*. 2022. **41**(1): p. 1-37.
122. Ashrafizadeh, M., et al., *Nanotechnological approaches in prostate cancer therapy: Integration of engineering and biology*. 2022. **45**: p. 101532.
123. Liu, Y., et al., *MIR-629-5p Promotes Prostate Cancer Development and Metastasis by Targeting AKAP13*. Front Oncol, 2021. **11**: p. 754353.

124. Wu, M., et al., *LncRNA MEG3 inhibits the progression of prostate cancer by modulating miR-9-5p/QKI-5 axis*. J Cell Mol Med, 2019. **23**(1): p. 29-38.
125. Yu, Y.Z., et al., *Hsa_circ_0003258 promotes prostate cancer metastasis by complexing with IGF2BP3 and sponging miR-653-5p*. Mol Cancer, 2022. **21**(1): p. 12.
126. Ding, Y., M. Wang, and J. Yang, *Circular RNA midline-1 (circMID1) promotes proliferation, migration, invasion and glycolysis in prostate cancer*. Bioengineered, 2022. **13**(3): p. 6293-6308.
127. Li, J., et al., *Circular RNA circVAMP3 promotes aerobic glycolysis and proliferation by regulating LDHA in renal cell carcinoma*. Cell Death Dis, 2022. **13**(5): p. 443.
128. Schreuders, E.H., et al., *Colorectal cancer screening: a global overview of existing programmes*. Gut, 2015. **64**(10): p. 1637-49.
129. Brenner, H., M. Kloor, and C.P. Pox, *Colorectal cancer*. Lancet, 2014. **383**(9927): p. 1490-1502.
130. Kaminski, M.F., et al., *Optimizing the Quality of Colorectal Cancer Screening Worldwide*. Gastroenterology, 2020. **158**(2): p. 404-417.
131. Yi, B., et al., *Circular RNA PLCE1 promotes epithelial mesenchymal transformation, glycolysis in colorectal cancer and M2 polarization of tumor-associated macrophages*. Bioengineered, 2022. **13**(3): p. 6243-6256.
132. Wang, X., et al., *Circular RNA NOX4 promotes the development of colorectal cancer via the microRNA-485-5p/CKS1B axis*. Oncol Rep, 2020. **44**(5): p. 2009-2020.
133. Wang, L., et al., *CircRNF13 regulates the invasion and metastasis in lung adenocarcinoma by targeting miR-93-5p*. Gene, 2018. **671**: p. 170-177.
134. Mo, Y., et al., *Circular RNA circRNF13 inhibits proliferation and metastasis of nasopharyngeal carcinoma via SUMO2*. Mol Cancer, 2021. **20**(1): p. 112.
135. Li, L. and Z.T. Zhang, *Hsa_circ_0086414 Might Be a Diagnostic Biomarker of Oral Squamous Cell Carcinoma*. Med Sci Monit, 2020. **26**: p. e919383.
136. Jiang, Q., et al., *Circular_0086414 induces SPARC like 1 (SPARCL1) production to inhibit esophageal cancer cell proliferation, invasion and glycolysis and induce cell apoptosis by sponging miR-1290*. Bioengineered, 2022. **13**(5): p. 12099-12114.
137. Gong, X., H. Tang, and K. Yang, *PER1 suppresses glycolysis and cell proliferation in oral squamous cell carcinoma via the PER1/RACK1/PI3K signaling complex*. Cell Death Dis, 2021. **12**(3): p. 276.
138. Zhang, Y., Y. Wu, and X. Su, *PLOD1 promotes cell growth and aerobic glycolysis by regulating the SOX9/PI3K/Akt/mTOR signaling pathway in gastric cancer*. Front Biosci (Landmark Ed), 2021. **26**(8): p. 322-334.
139. Gao, T., et al., *SIK2 promotes reprogramming of glucose metabolism through PI3K/AKT/HIF-1 α pathway and Drp1-mediated mitochondrial fission in ovarian cancer*. Cancer Lett, 2020. **469**: p. 89-101.
140. Liu, L., et al., *TRPM7 promotes the epithelial-mesenchymal transition in ovarian cancer through the calcium-related PI3K / AKT oncogenic signaling*. J Exp Clin Cancer Res, 2019. **38**(1): p. 106.
141. Yalan, S., et al., *Circular RNA circRHOBTB3 inhibits ovarian cancer progression through PI3K/AKT signaling pathway*. Panminerva Med, 2020.
142. Zhao, F., et al., *circFADS2 regulates lung cancer cells proliferation and invasion via acting as a sponge of miR-498*. Biosci Rep, 2018. **38**(4).
143. Xiao, Y.S., et al., *CircFADS2: A potential prognostic biomarker of colorectal cancer*. Exp Biol Med (Maywood), 2020. **245**(14): p. 1233-1241.
144. Zhang, Z., et al., *Circular RNA circFADS2 inhibits the progression of cutaneous squamous cell carcinoma by regulating miR-766-3p/HOXA9 axis*. Histol Histopathol, 2022. **37**(4): p. 335-348.
145. Zhu, L., et al., *Circular RNA Circ_0001777 Suppresses Lung Adenocarcinoma Progression In Vitro and In Vivo*. Biochem Genet, 2022.

146. Li, H., et al., *Therapeutic targeting of circ-CUX1/EWSR1/MAZ axis inhibits glycolysis and neuroblastoma progression*. EMBO Mol Med, 2019. **11**(12): p. e10835.
147. Dou, D., et al., *Circ_0008039 supports breast cancer cell proliferation, migration, invasion, and glycolysis by regulating the miR-140-3p/SKA2 axis*. Mol Oncol, 2021. **15**(2): p. 697-709.
148. Song, H.M., et al., *circRNA hsa_circ_0005909 Predicts Poor Prognosis and Promotes the Growth, Metastasis, and Drug Resistance of Non-Small-Cell Lung Cancer via the miRNA-338-3p/SOX4 Pathway*. Dis Markers, 2021. **2021**: p. 8388512.
149. Yang, C., et al., *Circ_0,007,331 Promotes the PTX Resistance and Progression of Breast Cancer via miR-200b-3p/ANLN*. J Surg Res, 2022. **279**: p. 619-632.
150. Xu, G., et al., *Circular RNA circNRIP1 Sponges microRNA-138-5p to Maintain Hypoxia-Induced Resistance to 5-Fluorouracil Through HIF-1 α -Dependent Glucose Metabolism in Gastric Carcinoma*. Cancer Manag Res, 2020. **12**: p. 2789-2802.
151. Park, M., et al., *Synergistically Anti-metastatic Effect of 5-Flourouracil on Colorectal Cancer Cells via Calcium-mediated Focal Adhesion Kinase Proteolysis*. Anticancer Res, 2017. **37**(1): p. 103-114.
152. Ressa, A.L., et al., *Spinophilin expression determines cellular growth, cancer stemness and 5-flourouracil resistance in colorectal cancer*. Oncotarget, 2014. **5**(18): p. 8492-502.
153. Wang, Y., et al., *LncRNA LINRIS stabilizes IGF2BP2 and promotes the aerobic glycolysis in colorectal cancer*. Mol Cancer, 2019. **18**(1): p. 174.
154. Weng, M.L., et al., *Fasting inhibits aerobic glycolysis and proliferation in colorectal cancer via the Fdft1-mediated AKT/mTOR/HIF1 α pathway suppression*. Nat Commun, 2020. **11**(1): p. 1869.
155. Gao, Y., et al., *Circular RNA sterile alpha motif domain containing 4A contributes to cell 5-fluorouracil resistance in colorectal cancer by regulating the miR-545-3p/6-phosphofructo-2-kinase/fructose-2,6-bisphosphataseisotype 3 axis*. Anticancer Drugs, 2022. **33**(6): p. 553-563.
156. Ashrafizadeh, M., et al., *Chitosan-based advanced materials for docetaxel and paclitaxel delivery: Recent advances and future directions in cancer theranostics*. International Journal of Biological Macromolecules, 2020. **145**: p. 282-300.
157. Zang, H., et al., *Circ-RNF111 contributes to paclitaxel resistance in breast cancer by elevating E2F3 expression via miR-140-5p*. Thorac Cancer, 2020. **11**(7): p. 1891-1903.
158. Entezari, M., et al., *Long non-coding RNAs and exosomal lncRNAs: Potential functions in lung cancer progression, drug resistance and tumor microenvironment remodeling*. Biomedicine & Pharmacotherapy, 2022. **150**: p. 112963.
159. Ashrafizadeh, M., et al., *Therapeutic potential of AMPK signaling targeting in lung cancer: Advances, challenges and future prospects*. Life Sciences, 2021. **278**: p. 119649.
160. Hirsch, F.R., et al., *New and emerging targeted treatments in advanced non-small-cell lung cancer*. 2016. **388**(10048): p. 1012-1024.
161. Zhang, D., et al., *Nuclear Factor- κ B Inhibition by Parthenolide Potentiates the Efficacy of Taxol in Non-Small Cell Lung Cancer In vitro and In vivoParthenolide Potentiates Chemosensitization*. 2009. **7**(7): p. 1139-1149.
162. Wu, Y., et al., *Circular RNA hsa_circ_0011298 enhances Taxol resistance of non-small cell lung cancer by regulating miR-486-3p/CRABP2 axis*. J Clin Lab Anal, 2022. **36**(5): p. e24408.
163. Li, R., et al., *miR-543 impairs cell proliferation, migration, and invasion in breast cancer by suppressing VCAN*. Biochem Biophys Res Commun, 2021. **570**: p. 191-198.
164. Li, L. and Q. Li, *miR-543 impairs breast cancer cell phenotypes by targeting and suppressing ubiquitin-conjugating enzyme E2T (UBE2T)*. Bioengineered, 2021. **12**(2): p. 12394-12406.
165. Xiu, D., et al., *MicroRNA-543 suppresses liver cancer growth and induces apoptosis via the JAK2/STAT3 signaling pathway*. Oncol Lett, 2019. **17**(2): p. 2451-2456.

166. Zhang, Y., et al., *Circular RNA FOXO3 accelerates glycolysis and improves cisplatin sensitivity in lung cancer cells via the miR-543/Foxo3 axis*. *Oncol Lett*, 2021. **22**(6): p. 839.
167. Che, H., H. Ding, and X. Jia, *circ_0080145 Enhances Imatinib Resistance of Chronic Myeloid Leukemia by Regulating miR-326/PPFIA1 Axis*. *Cancer Biother Radiopharm*, 2020.
168. Kumari, R., et al., *Hepatocellular carcinoma treatment: hurdles, advances and prospects*. *Hepat Oncol*, 2018. **5**(2): p. Hep08.
169. Miguet, M., et al., *Multidisciplinary meetings specific to hepatocellular carcinoma: How to proceed?* *J Visc Surg*, 2019. **156**(3): p. 217-227.
170. Hiraoka, A., et al., *Newly Proposed ALBI Grade and ALBI-T Score as Tools for Assessment of Hepatic Function and Prognosis in Hepatocellular Carcinoma Patients*. *Liver Cancer*, 2019. **8**(5): p. 312-325.
171. McCarthy, C.M., M. McCarthy, and K. O'Donoghue, *Recurrent hepatocellular carcinoma in pregnancy: A case report and literature review*. *Obstet Med*, 2019. **12**(4): p. 202-204.
172. Lu, Y., et al., *Epigenetic regulation of ferroptosis via ETS1/miR-23a-3p/ACSL4 axis mediates sorafenib resistance in human hepatocellular carcinoma*. *J Exp Clin Cancer Res*, 2022. **41**(1): p. 3.
173. Chen, Y.T., et al., *Upregulation of lncRNA NIFK-AS1 in hepatocellular carcinoma by m(6)A methylation promotes disease progression and sorafenib resistance*. *Hum Cell*, 2021. **34**(6): p. 1800-1811.
174. Paskeh, M.D.A., et al., *Wnt/ β -Catenin Signaling as a Driver of Hepatocellular Carcinoma Progression: An Emphasis on Molecular Pathways*. 2021. **8**: p. 1415.
175. Huang, H., et al., *Circular RNA circUBE2D2 functions as an oncogenic factor in hepatocellular carcinoma sorafenib resistance and glycolysis*. *Am J Transl Res*, 2021. **13**(6): p. 6076-6086.
176. Shi, Q., et al., *Serum Exosomes-Based Biomarker circ_0008928 Regulates Cisplatin Sensitivity, Tumor Progression, and Glycolysis Metabolism by miR-488/HK2 Axis in Cisplatin-Resistant Non-small Cell Lung Carcinoma*. *Cancer Biother Radiopharm*, 2021.
177. Li, H., et al., *Astragalus IV Undermines Multi-Drug Resistance and Glycolysis of MDA-MB-231/ADR Cell Line by Depressing hsa_circ_0001982-miR-206/miR-613 Axis*. *Cancer Manag Res*, 2021. **13**: p. 5821-5833.
178. Paskeh, M.D.A., et al., *Emerging role of exosomes in cancer progression and tumor microenvironment remodeling*. *Journal of Hematology & Oncology*, 2022. **15**(1): p. 83.
179. Zhou, Y., et al., *The Role of Exosomes and Their Applications in Cancer*. *Int J Mol Sci*, 2021. **22**(22).
180. Li, C., et al., *Cancer associated-fibroblast-derived exosomes in cancer progression*. *Mol Cancer*, 2021. **20**(1): p. 154.
181. Thakur, A., et al., *Exosomes: Small vesicles with big roles in cancer, vaccine development, and therapeutics*. *Bioact Mater*, 2022. **10**: p. 281-294.
182. Wang, X., et al., *Exosome-delivered circRNA promotes glycolysis to induce chemoresistance through the miR-122-PKM2 axis in colorectal cancer*. *Mol Oncol*, 2020. **14**(3): p. 539-555.
183. Li, C. and X. Li, *Exosome-Derived Circ_0094343 Promotes Chemosensitivity of Colorectal Cancer Cells by Regulating Glycolysis via the miR-766-5p/TRIM67 Axis*. *Contrast Media Mol Imaging*, 2022. **2022**: p. 2878557.
184. Bajhan, E., et al., *MicroRNA-143 inhibits proliferation and migration of prostate cancer cells*. *Arch Physiol Biochem*, 2022. **128**(5): p. 1323-1329.
185. Asghariazar, V., et al., *Restoration of miR-143 reduces migration and proliferation of bladder cancer cells by regulating signaling pathways involved in EMT*. *Mol Cell Probes*, 2022. **61**: p. 101794.

186. Chen, J. and J. Gong, *CircMMP11 overexpression predicts the poor survival of non-small cell lung cancer and downregulates miR-143 through methylation to suppress cell proliferation*. J Cardiothorac Surg, 2021. **16**(1): p. 327.
187. Lin, G.R., et al., *Circular RNA circ_0006089 promotes the progression of gastric cancer by regulating the miR-143-3p/PTBP3 axis and PI3K/AKT signaling pathway*. J Dig Dis, 2022. **23**(7): p. 376-387.
188. Tan, W.Q., et al., *Exosome-delivered circular RNA DLGAP4 induces chemoresistance via miR-143-HK2 axis in neuroblastoma*. Cancer Biomark, 2022. **34**(3): p. 375-384.
189. Ma, J., G. Qi, and L. Li, *A Novel Serum Exosomes-Based Biomarker hsa_circ_0002130 Facilitates Osimertinib-Resistance in Non-Small Cell Lung Cancer by Sponging miR-498*. Onco Targets Ther, 2020. **13**: p. 5293-5307.
190. Dai, X., S. Zhang, and K.J.M.b.r. Zaleta-Rivera, *RNA: interactions drive functionalities*. 2020. **47**(2): p. 1413-1434.
191. Ashrafizadeh, M., et al., *Interplay between SOX9 transcription factor and microRNAs in cancer*. International Journal of Biological Macromolecules, 2021. **183**: p. 681-694.
192. Mirzaei, S., et al., *The role of microRNA-338-3p in cancer: growth, invasion, chemoresistance, and mediators*. Life Sciences, 2021. **268**: p. 119005.
193. Abadi, A.J., et al., *Small in size, but large in action: microRNAs as potential modulators of PTEN in breast and lung cancers*. 2021. **11**(2): p. 304.
194. Ashrafizadeh, M., et al., *MicroRNA-mediated autophagy regulation in cancer therapy: the role in chemoresistance/chemosensitivity*. 2021. **892**: p. 173660.
195. Ashrafizadeh, M., et al., *Lung cancer cells and their sensitivity/resistance to cisplatin chemotherapy: Role of microRNAs and upstream mediators*. 2021. **78**: p. 109871.
196. Mirzaei, S., et al., *MicroRNAs regulating SOX2 in cancer progression and therapy response*. 2021. **23**.
197. Wang, G., et al., *miR-99a-5p inhibits glycolysis and induces cell apoptosis in cervical cancer by targeting RRGD*. Oncol Lett, 2022. **24**(1): p. 228.
198. Arora, S., et al., *miR-16-5p regulates aerobic glycolysis and tumorigenesis of NSCLC cells via LDH-A/lactate/NF- κ B signaling*. Life Sci, 2022. **304**: p. 120722.
199. Cao, L., et al., *Circular RNA circRNF20 promotes breast cancer tumorigenesis and Warburg effect through miR-487a/HIF-1 α /HK2*. Cell Death Dis, 2020. **11**(2): p. 145.
200. Ding, X., et al., *CTHRC1 promotes gastric cancer metastasis via HIF-1 α /CXCR4 signaling pathway*. Biomed Pharmacother, 2020. **123**: p. 109742.
201. Gao, L., et al., *CD90 affects the biological behavior and energy metabolism level of gastric cancer cells by targeting the PI3K/AKT/HIF-1 α signaling pathway*. Oncol Lett, 2021. **21**(3): p. 191.
202. Liu, J., et al., *Circular RNA circ-MAT2B facilitates glycolysis and growth of gastric cancer through regulating the miR-515-5p/HIF-1 α axis*. Cancer Cell Int, 2020. **20**: p. 171.
203. Zhan, P., et al., *miR-98-5p inhibits gastric cancer cell stemness and chemoresistance by targeting branched-chain aminotransferases 1*. Life Sci, 2021. **276**: p. 119405.
204. Jiang, F., et al., *MicroRNA-98-5p inhibits proliferation and metastasis in non-small cell lung cancer by targeting TGFBR1*. Int J Oncol, 2019. **54**(1): p. 128-138.
205. Xiao, R., H. Wang, and B. Yang, *MicroRNA-98-5p modulates cervical cancer progression via controlling PI3K/AKT pathway*. Bioengineered, 2021. **12**(2): p. 10596-10607.
206. Qin, C., et al., *Circular RNA 0006349 Augments Glycolysis and Malignance of Non-small Cell Lung Cancer Cells Through the microRNA-98/MKP1 Axis*. Front Cell Dev Biol, 2021. **9**: p. 690307.
207. Liu, W., C. Yin, and Y. Liu, *Circular RNA circ_0091579 Promotes Hepatocellular Carcinoma Proliferation, Migration, Invasion, and Glycolysis Through miR-490-5p/CASC3 Axis*. Cancer Biother Radiopharm, 2021. **36**(10): p. 863-878.

208. Li, C., et al., *S100A2 promotes glycolysis and proliferation via GLUT1 regulation in colorectal cancer*. *Faseb j*, 2020. **34**(10): p. 13333-13344.
209. Kim, J., et al., *SALL4 promotes glycolysis and chromatin remodeling via modulating HP1 α -Glut1 pathway*. *Oncogene*, 2017. **36**(46): p. 6472-6479.
210. Zhang, Z.J., et al., *Circular RNA circDENND4C facilitates proliferation, migration and glycolysis of colorectal cancer cells through miR-760/GLUT1 axis*. *Eur Rev Med Pharmacol Sci*, 2020. **24**(5): p. 2387-2400.
211. Zhang, K., et al., *Circ_0001421 facilitates glycolysis and lung cancer development by regulating miR-4677-3p/CDCA3*. *Diagn Pathol*, 2020. **15**(1): p. 133.
212. Xiao, Y.Y., et al., *ZEB1 promotes invasion and metastasis of endometrial cancer by interacting with HDGF and inducing its transcription*. *Am J Cancer Res*, 2019. **9**(11): p. 2314-2330.
213. Sun, Y., J. Li, and S. Zheng, *MiR-769-5p, Which Targets HDGF, Inhibits Cell Proliferation and Invasion in Nonsmall Cell Lung Cancer*. *Cancer Biother Radiopharm*, 2021.
214. Zhang, Z., et al., *MicroRNA-139-5p inhibits cell viability, migration and invasion and suppresses tumor growth by targeting HDGF in non-small cell lung cancer*. *Oncol Lett*, 2020. **19**(3): p. 1806-1814.
215. Guo, F., et al., *Circular RNA circMAGI3 accelerates the glycolysis of non-small cell lung cancer through miR-515-5p/HDGF*. *Am J Transl Res*, 2020. **12**(7): p. 3953-3963.
216. Chen, L., et al., *MicroRNA-106a regulates phosphatase and tensin homologue expression and promotes the proliferation and invasion of ovarian cancer cells*. 2016. **36**(4): p. 2135-2141.
217. You, F., et al., *miRNA-106a promotes breast cancer cell proliferation, clonogenicity, migration, and invasion through inhibiting apoptosis and chemosensitivity*. 2019. **38**(2): p. 198-207.
218. Luo, B., et al., *Oncogene miR-106a promotes proliferation and metastasis of prostate cancer cells by directly targeting PTEN in vivo and in vitro*. 2017. **109**(1): p. 24-30.
219. Huang, Q. and Q.J.O.L. Ma, *MicroRNA-106a inhibits cell proliferation and induces apoptosis in colorectal cancer cells*. 2018. **15**(6): p. 8941-8944.
220. Lai, Y., et al., *LINC00116 enhances cervical cancer tumorigenesis through miR-106a/c-Jun pathway*. 2020. **121**(3): p. 2247-2257.
221. Lin, C., et al., *Circular RNA ITCH suppresses proliferation, invasion, and glycolysis of ovarian cancer cells by up-regulating CDH1 via sponging miR-106a*. *Cancer Cell Int*, 2020. **20**: p. 336.
222. Lin, Q., H. Jiang, and D. Lin, *Circular RNA ITCH downregulates GLUT1 and suppresses glucose uptake in melanoma to inhibit cancer cell proliferation*. *J Dermatolog Treat*, 2021. **32**(2): p. 231-235.
223. Jiang, L., et al., *YAP promotes the proliferation and migration of colorectal cancer cells through the Glut3/AMPK signaling pathway*. *Oncol Lett*, 2021. **21**(4): p. 312.
224. Shimizu, M. and N. Tanaka, *IL-8-induced O-GlcNAc modification via GLUT3 and GFAT regulates cancer stem cell-like properties in colon and lung cancer cells*. *Oncogene*, 2019. **38**(9): p. 1520-1533.
225. Li, X., et al., *Ajuba Overexpression Promotes Breast Cancer Chemoresistance and Glucose Uptake through TAZ-GLUT3/Survivin Pathway*. *Biomed Res Int*, 2022. **2022**: p. 3321409.
226. Xiong, S., et al., *Circular RNA MYLK Promotes Glycolysis and Proliferation of Non-Small Cell Lung Cancer Cells by Sponging miR-195-5p and Increasing Glucose Transporter Member 3 Expression*. *Cancer Manag Res*, 2020. **12**: p. 5469-5478.
227. Liu, Y., et al., *Hsa_circ_0000231 knockdown inhibits the glycolysis and progression of colorectal cancer cells by regulating miR-502-5p/MYO6 axis*. *World J Surg Oncol*, 2020. **18**(1): p. 255.
228. Dou, Y., et al., *Circ_0001944 Contributes to Glycolysis and Tumor Growth by Upregulating NFAT5 Through Acting as a Decoy for miR-142-5p in Non-Small Cell Lung Cancer*. *Cancer Manag Res*, 2021. **13**: p. 3775-3787.

229. Ding, Z., et al., *Circ-PRMT5 enhances the proliferation, migration and glycolysis of hepatoma cells by targeting miR-188-5p/HK2 axis*. Ann Hepatol, 2020. **19**(3): p. 269-279.
230. Wu, S., Y. Tang, and W. Liu, *Circ_0084043 promotes cell proliferation and glycolysis but blocks cell apoptosis in melanoma via circ_0084043-miR-31-KLF3 axis*. Open Life Sci, 2020. **15**(1): p. 774-786.
231. Zhao, N., L. Hu, and H. Chen, *Circ_0002762 accelerates glycolysis metabolism to promote cervical cancer progression via the miR-526b-5p/HK2 axis*. Gynecol Obstet Invest, 2022.
232. Guo, J., Y. Su, and M. Zhang, *Circ_0000140 restrains the proliferation, metastasis and glycolysis metabolism of oral squamous cell carcinoma through upregulating CDC73 via sponging miR-182-5p*. Cancer Cell Int, 2020. **20**: p. 407.
233. Zan, X., et al., *Circ-CSNK1G1 promotes cell proliferation, migration, invasion and glycolysis metabolism during triple-negative breast cancer progression by modulating the miR-28-5p/LDHA pathway*. Reprod Biol Endocrinol, 2022. **20**(1): p. 138.
234. Huang, W., et al., *Downregulation of Hsa_circ_0000735 Inhibits the Proliferation, Migration, Invasion, and Glycolysis in Non-small-cell Lung Cancer by Targeting miR-940/BMPER Axis*. Onco Targets Ther, 2020. **13**: p. 8427-8439.
235. Liu, J., et al., *Circ_0009910 Serves as miR-361-3p Sponge to Promote the Proliferation, Metastasis, and Glycolysis of Gastric Cancer via Regulating SNRPA*. Biochem Genet, 2022. **60**(5): p. 1809-1824.
236. Liu, J. and J. Liu, *Circ_0000442 functions as a tumor repressor in breast cancer by impacting miR-1229-3p and upregulating ZBTB1*. Mamm Genome, 2022. **33**(3): p. 543-554.
237. Yan, X., T. Wang, and J. Wang, *Circ_0016760 Acts as a Sponge of MicroRNA-4295 to Enhance E2F Transcription Factor 3 Expression and Facilitates Cell Proliferation and Glycolysis in Non-Small Cell Lung Cancer*. Cancer Biother Radiopharm, 2022. **37**(2): p. 147-158.
238. Xiao, Y., et al., *Circ_0047921 acts as the sponge of miR-1287-5p to stimulate lung cancer progression by regulating proliferation, migration, invasion, and glycolysis of lung cancer cells*. World J Surg Oncol, 2022. **20**(1): p. 108.
239. Qiu, Z., L. Wang, and H. Liu, *Hsa_circ_0001982 promotes the progression of breast cancer through miR-1287-5p/MUC19 axis under hypoxia*. World J Surg Oncol, 2021. **19**(1): p. 161.
240. Wang, Y., et al., *A novel circ_0099999/miR-330-5p/FSCN1 ceRNA crosstalk in pancreatic cancer*. Autoimmunity, 2021. **54**(7): p. 471-482.
241. Li, Y., et al., *circ_0136666 Facilitates the Progression of Colorectal Cancer via miR-383/CREB1 Axis*. Cancer Manag Res, 2020. **12**: p. 6795-6806.
242. Wen, X., J. Du, and X. Wang, *Circ_0039411 promotes papillary thyroid carcinoma development through mediating the miR-423-5p/SOX4 signaling*. Int J Biol Markers, 2021. **36**(4): p. 10-20.
243. Wang, W., et al., *circ_0002346 Suppresses Non-Small-Cell Lung Cancer Progression Depending on the Regulation of the miR-582-3p/STXBP6 Axis*. Int J Genomics, 2021. **2021**: p. 1565660.
244. Yue, Q., et al., *Circ-PITX1 Promotes the Progression of Non-Small Cell Lung Cancer Through Regulating the miR-1248/CCND2 Axis*. Onco Targets Ther, 2021. **14**: p. 1807-1819.
245. Lu, H., L. Gao, and J. Lv, *Circ_0078710 promotes the development of liver cancer by upregulating TXNDC5 via miR-431-5p*. Ann Hepatol, 2022. **27**(1): p. 100551.
246. Qi, C., et al., *Circ_0072995 Promotes Cell Carcinogenesis via Up-Regulating miR-149-5p-Mediated SHMT2 in Breast Cancer*. Cancer Manag Res, 2020. **12**: p. 11169-11181.
247. Wan, L., et al., *Circ-TFF1 Facilitates Breast Cancer Development via Regulation of miR-338-3p/FGFR1 Axis*. Biochem Genet, 2022. **60**(1): p. 315-335.
248. Gao, Y., et al., *CircRNA Circ_0001721 Promotes the Progression of Osteosarcoma Through miR-372-3p/MAPK7 Axis*. Cancer Manag Res, 2020. **12**: p. 8287-8302.

249. Li, J., Y. Li, and H. Cheng, *Circ-RPPH1 knockdown retards breast cancer progression via miR-328-3p-mediated suppression of HMGA2*. Clin Breast Cancer, 2022. **22**(3): p. e286-e295.
250. Zhang, X. and X. Zheng, *Hsa_circ_0001495 contributes to cervical cancer progression by targeting miR-526b-3p/TMBIM6/mTOR axis*. Reprod Biol, 2022. **22**(2): p. 100648.
251. Ge, L., et al., *Circ_0026134 promotes NSCLC progression by the miR-3619-5p/CHAF1B axis*. Thorac Cancer, 2022. **13**(4): p. 582-592.
252. Huang, Y., et al., *Circ_0000467 Exerts an Oncogenic Role in Colorectal Cancer via miR-330-5p-Dependent Regulation of TYRO3*. Biochem Genet, 2022. **60**(5): p. 1488-1510.
253. Hong, F., et al., *Hsa_circ_0045932 regulates the progression of colorectal cancer by regulating HK2 through sponging miR-873-5p*. J Clin Lab Anal, 2022. **36**(9): p. e24641.
254. Li, Y., et al., *CircRNA hsa_circ_0018289 exerts an oncogenic role in cervical cancer progression through miR-1294/ICMT axis*. J Clin Lab Anal, 2022. **36**(5): p. e24348.
255. Cheng, H., et al., *Circ_0001955 plays a carcinogenic role in breast cancer via positively regulating GLUT1 via decoying miR-1299*. Thorac Cancer, 2022. **13**(7): p. 913-924.
256. Shen, S., et al., *Circ_0008717 promotes renal cell carcinoma progression by upregulating FBXO17 via targeting miR-217*. J Gene Med, 2022: p. e3418.
257. Li, C., et al., *Circ_0000376, a Novel circRNA, Promotes the Progression of Non-Small Cell Lung Cancer Through Regulating the miR-1182/NOVA2 Network*. Cancer Manag Res, 2020. **12**: p. 7635-7647.
258. Lu, P., Y. Jiang, and Z. Xia, *Hsa_circ_0003221 facilitates the malignant development of bladder cancer cells via resulting in the upregulation of DHCR24 by targeting miR-892b*. Investig Clin Urol, 2022. **63**(5): p. 577-588.
259. Kim, B., et al., *The efficacy of topical bupivacaine and triamcinolone acetonide injection in the relief of pain after endoscopic submucosal dissection for gastric neoplasia: a randomized double-blind, placebo-controlled trial*. 2015. **29**(3): p. 714-722.
260. Bundscherer, A., et al., *Effects of ropivacaine, bupivacaine and sufentanil in colon and pancreatic cancer cells in vitro*. 2015. **95**: p. 126-131.
261. Xuan, W., et al., *Local anesthetic bupivacaine induced ovarian and prostate cancer apoptotic cell death and underlying mechanisms in vitro*. 2016. **6**(1): p. 1-12.
262. Ju, C., et al., *Bupivacaine suppresses the progression of gastric cancer through regulating circ_0000376/miR-145-5p axis*. BMC Anesthesiol, 2020. **20**(1): p. 275.
263. Irwin, M., et al., *Influence of propofol-based total intravenous anaesthesia on peri-operative outcome measures: a narrative review*. 2020. **75**: p. e90-e100.
264. Bateman, B.T. and A.S.J.D.D.T. Kesselheim, *Propofol as a transformative drug in anesthesia: insights from key early investigators*. 2015. **20**(8): p. 1012-1017.
265. Liu, Y.P., et al., *Propofol induces ferroptosis and inhibits malignant phenotypes of gastric cancer cells by regulating miR-125b-5p/STAT3 axis*. World J Gastrointest Oncol, 2021. **13**(12): p. 2114-2128.
266. Zhang, H., et al., *Propofol Inhibits Thyroid Cancer Cell Proliferation, Migration, and Invasion by Suppressing SHH and PI3K/AKT Signaling Pathways via the miR-141-3p/BRD4 Axis*. J Healthc Eng, 2021. **2021**: p. 2704753.
267. Gao, J., et al., *Propofol suppresses lung cancer tumorigenesis by modulating the circ-ERBB2/miR-7-5p/FOXO1 axis*. Thorac Cancer, 2021. **12**(6): p. 824-834.
268. Huang, X., et al., *Propofol inhibits invasion and growth of ovarian cancer cells via regulating miR-9/NF- κ B signal*. Braz J Med Biol Res, 2016. **49**(12): p. e5717.
269. Shen, X., et al., *Propofol inhibits proliferation, migration, invasion and promotes apoptosis by regulating HOST2/JAK2/STAT3 signaling pathway in ovarian cancer cells*. Cytotechnology, 2021. **73**(2): p. 243-252.

270. Sun, Y., et al., *Propofol inhibits proliferation and cisplatin resistance in ovarian cancer cells through regulating the microRNA-374a/forkhead box O1 signaling axis*. Mol Med Rep, 2020. **21**(3): p. 1471-1480.
271. Qu, D., X. Zou, and Z. Liu, *Propofol modulates glycolysis reprogramming of ovarian tumor via restraining circular RNA-zinc finger RNA-binding protein/microRNA-212-5p/superoxide dismutase 2 axis*. Bioengineered, 2022. **13**(5): p. 11881-11892.
272. Yang, H., et al., *Circ_MUC16 attenuates the effects of Propofol to promote the aggressive behaviors of ovarian cancer by mediating the miR-1182/S100B signaling pathway*. BMC Anesthesiol, 2021. **21**(1): p. 297.
273. Zhou, J., et al., *Tanshinone I attenuates the malignant biological properties of ovarian cancer by inducing apoptosis and autophagy via the inactivation of PI3K/AKT/mTOR pathway*. Cell Prolif, 2020. **53**(2): p. e12739.
274. Wang, W., et al., *Tanshinone I inhibits the growth and metastasis of osteosarcoma via suppressing JAK/STAT3 signalling pathway*. J Cell Mol Med, 2019. **23**(9): p. 6454-6465.
275. Jian, S., et al., *Tanshinone I induces apoptosis and protective autophagy in human glioblastoma cells via a reactive oxygen species-dependent pathway*. Int J Mol Med, 2020. **45**(4): p. 983-992.
276. Ye, B., et al., *Tanshinone I restrains osteosarcoma progression by regulating circ_0000376/miR-432-5p/BCL2 axis*. Mol Cell Biochem, 2022. **477**(1): p. 1-13.
277. Wang, Y., et al., *Nanoparticles loaded with circ_0086375 for suppressing the tumorigenesis of pancreatic cancer by targeting the miR-646/SLC4A4 axis*. Clin Exp Metastasis, 2023. **40**(1): p. 53-67.

Declaration of interests

☒ The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

☐ The authors declare the following financial interests/personal relationships which may be considered as potential competing interests: