



Pharmacology of Endogenous Opioids, Opiates and Their Receptors

12

Mohammed Noorladeen Al-Qattan, Nirupam Das,
and Rati Kailash Prasad Tripathi

Abstract

The search for analgesia is the main drive for the discovery of opioid receptors and their endogenous ligand peptides. Although opioid peptides are very similar in their *N*-terminal sequence, they are classified into different types according to their precursor proteins. The peptides activate opioid receptors by binding to orthosteric-binding sites to mediate intracellular second messengers. Biased signaling and allosteric modulation is a new approach to obtain receptor subtype selectivity and separates the desirable from a myriad of unwanted pharmacological effects. This chapter describes endogenous opioids and opiates with an emphasis on structure, origin and processing, receptors, physiological roles, and potential involvement in therapeutic interventions. Further, it also provides a brief discussion on the effect of opioids on various ion channels and recent developments of established and investigational opioid molecules.

Keywords

Opioids receptors · Endorphin · Enkephalin · Dynorphin · GPCR · Neurological disorders · Ion channels

Abbreviations

ACTH Adrenocorticotropin
BACE1 Beta-site APP cleaving enzyme 1

M. N. Al-Qattan
Department of Pharmacy, Al-Noor University College, Mosul, Iraq

N. Das (✉) · R. K. P. Tripathi
Department of Pharmaceutical Science, Sushruta School of Medical and Paramedical Sciences,
Assam University, Silchar, Assam, India

CLIP	Corticotropin-like intermediate peptide
DOR	Delta opioid receptor
GPCR	G protein-coupled receptor
KOR	Kappa opioid receptor
MOR	Mu opioid receptor
MSH	Melanotopins or melanocyte-stimulating hormone
N/OFQ	Nociceptin/orphanin FQ
OP	Opioid peptide
OR	Opioid receptor
ORL-1	Opioid receptor-like-1
POMC	Proopiomelanocortin

12.1 Introduction

The inscription of the ancient Sumerian clay tablet insinuates the use of the natural extract of poppy plant (*Papaver somniferum*) as analgesia and post-surgical pain (Norn et al. 2005; Brownstein 1993). The active ingredient of the extract is morphine, which necessitates the presence of receptors for binding inside the human body. The search for endogenous ligands acting on those opioid receptors (ORs) started in the 1960s and succeeded in a few years by using radiolabeled ligands (Pert and Snyder 1973). On the search for ligands for those ORs, several peptides were isolated. The early peptides were enkephalins (Hughes et al. 1975) which showed higher potency than morphine on opioid receptors (Kosterlitz and Hughes 1975). Soon the untriakontapeptide (endorphin) was discovered (Li and Chung 1976), then dynorphins (Cox et al. 1975) and endomorphins (Zadina et al. 1997).

Although opioid peptides (OPs) share almost similar *N*-terminal amino acid sequence, their precursor proteins are different. The processing of precursor proteins differs from one tissue to another as well as variation of the half-life of opioid peptides. Some endogenous opioid peptides like endomorphins act as a selective agonist for μ -opioid receptor (MOR) and thus have a good potential for clinical use. However, the low oral bioavailability of endomorphins restricts its use unless given as glycosylated derivatives (Varamini et al. 2012). Currently, three types of opioid receptors are known, namely Mu, Delta, and Kappa opioid receptors (MORs, DORs and KORs, respectively). The receptors can be activated by orthosteric ligands and cooperated with allosteric ligands to produce modulation of orthosteric affinity/efficacy as well as biasing the intracellular signaling.

12.2 Opioid Peptides

12.2.1 Classification, Synthesis, Storage, Release, and Removal

Endogenous opioid peptides are classified into three categories, namely endorphins, enkephalins, and dynorphins. The peptides are endogenously produced by peptidases digestion of larger precursor proteins, namely proopiomelanocortin,

proenkephalin and prodynorphin, respectively (Plevry 1991). Endomorphins is regarded as the fourth category of OPs; however, the precursor for endomorphins is not precisely known from the human genome (Terskiy et al. 2007) and may involve an oxidative transformation of guanine nucleotide from suspected gene leading to G → T transversion in the produced mRNA (messenger ribonucleic acid) (Matsushima et al. 2019). The proopiomelanocortin (POMC) precursor is processed to give rise to adrenocorticotropin (ACTH), melanotropins (MSH) and β -endorphin (Castro and Morrison 1997). The processing of precursor protein depends on a particular cell type. At anterior pituitary, POMC is processed in corticotrophs to generate mainly ACTH and β -lipotropin. When processed in melanotrophs, it produces MSH primarily (De Wied 1999). Both ACTH and β -lipotropin (1–91 aa) have no analgesic activity. The further processing of β -lipotropin gives rise to β -endorphin (61–91 aa), which in turn may yield [Met]-enkephalin (61–65) (Cox et al. 1976; Takeuchi 2001). In melanotrophs of pars intermediate of the pituitary, POMC is processed further to produce α -MSH (from N-terminal of POMC) which is attributed to different levels of specific peptidases in different regions of the pituitary (Day 2009). In addition to the pituitary, POMC is also detected in the arcuate nucleus of the hypothalamus, solitary tract of the medulla and several peripheral tissues (Papadimitriou and Priftis 2009).

Proenkephalin peptide contains six copies of [Met]-enkephalin and one copy of [Leu]-enkephalin, which are separated from each other by dibasic peptides as digestive site (Takahashi 2016). Although proenkephalin is produced by both the brain and adrenal gland, however, it is processed differently in each tissue (Geraciotti et al. 2009). Proenkephalin-producing neurons are more widespread in the brain than POMC neurons (Geraciotti et al. 2009). Prodynorphin is also called proenkephalin-B since it contains three copies of [Leu]-enkephalins in addition to dynorphin A, dynorphin B and α -neoeendorphin (Fig. 12.1 and Table 12.1).

In neuron soma, precursor neuropeptides are synthesized in the endoplasmic reticulum, and through Golgi apparatus, the peptides are processed by proteases and undergo functional group modification and then transported to axons as vesicles. At the nerve terminal, the vesicles may contain both precursor and opioid peptides (Hökfelt et al. 2000). The processing of precursor peptide is activated by potassium-induced depolarization of the neuron (Yakovleva et al. 2006). Moreover, the acidic pH of secretory vacuoles promotes digestion of precursor peptides such as POMC (Cawley et al. 2016). The opioid peptides vesicles appeared distinct from amino acid neurotransmitter vesicles, and the former appear denser under a microscope. The dense vesicles are distributed over neuronal cells but especially abundant in dendrites, cell body and axon varicosities and can be released from either site with machinery different from neurotransmitters (Russo 2017; Gu et al. 2017).

Post-translational modifications may take place on opioid peptides such as glycosylation, acetylation, methylation and phosphorylation which alter their biological activities (Froehlich 1997). While glycosylation improves the blood-brain barrier (BBB) penetration (Egleton et al. 2000), acetylation attenuates the activity of the peptide. Glycosylation may occur even on precursor proteins and thus controls the degree of digestion to active opioid peptides and protects against non-specific digestion (Hughes et al. 1980; Loh and Gainer 1978).

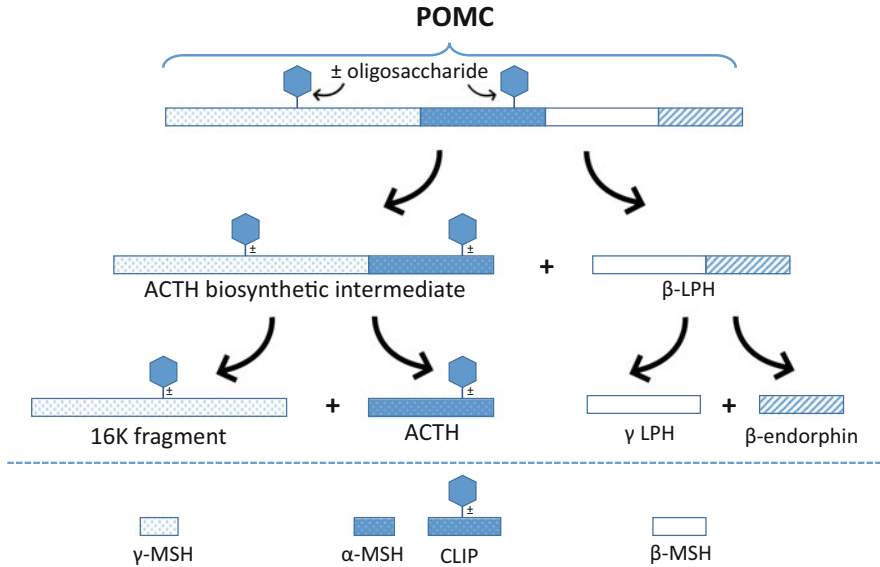


Fig. 12.1 The processing of POMC (Retrieved from Mains and Eipper 1981)

The neuropeptides (including opioids) may diffuse to distal sites (up to millimeters), unlike other neurotransmitters, which remain within the synaptic space. They exert their effect by interacting with opioid and other receptors and may have longer extracellular half-lives (van den Pol 2012; Russo 2017). Long-distance signaling within the brain through no synapses is called volume transmission (Agnati et al. 1995; Veening et al. 2012). Volume transmission is common with G protein-coupled receptors (GPCRs) that are more sensitive to neuropeptides where the nanomolar concentration of opioid peptides is sufficient to activate the receptors. This is unlike the ionotropic receptors like γ -aminobutyric acid (GABA), where micromolar concentration is needed (Ludwig and Leng 2006).

Another difference between opioid peptides and neurotransmitters is that opioid peptides lack reuptake mechanism by releasing neurons. Therefore, the inactivation mechanism is limited to digestion by peptidases extracellularly or after cellular internalization. Many peptidases degrade opioid peptides. The major peptidases involved in the degradation of opioid peptides are endopeptidases (those acting on *N*-terminal of peptides, the pharmacophoric domain). However, other peptidases also contribute such as an insulin-degrading enzyme, angiotensin-converting enzyme, neprilysin, serine peptidases and other dipeptidyl peptidases (Asvadi et al. 2014a; Malfroy et al. 1978; Turner 2004; Hersh and Rodgers 2008). Since the activity of these peptidases is affected by the pH of the tissue, the half-life of these peptidases increases in inflamed tissues that have acidic pH; therefore, opioid peptides are degraded more rapidly and differently (Asvadi et al. 2014b; Herath et al. 2012). The inhibition of the degrading enzymes by specific and non-specific inhibitors is effective in producing analgesia.

Table 12.1 Opioid peptides and their precursor proteins and receptor selectivity (Luca et al. 2007; Coward et al. 1998)

Endogenous peptide	Amino acid sequence	Receptor affinity	Precursor
β -Endorphin	YGGFMTSEKTSQIPLVTLFKNAIIKNA YKKGE	$\delta = \mu$	Proopiomelanocortin
[Met]-enkephalin	YGGFM (YGGFMRF, YGGFMRGL)	$\delta > > \mu$	Proenkephalin
[Leu]-enkephalin	YGGFL		
Metorphinamide	YGGFMRRV-NH ₂		
Dynorphin A	YGGFLRRIRPKLKWDNQ	$\kappa > > \delta = \mu$	Prodynorphin
Dynorphin A(1-8)	YGGFLRRI		
Dynorphin B	YGGFLRRQFKVVT		
α -neoendorphin	YGGFLRKYPK		
β -neoendorphin	YGGFLRKYP		
Nociceptin	FGFTGARKSARKLANQ	*ORL	Pronociceptin
Endomorphin-1	YPWF-NH ₂	μ	Unknown
Endomorphin-2	YPPF-NH ₂		

*ORL means orphan opioid-receptor-like

12.2.2 Physiological Role of Opioid Peptides

12.2.2.1 Sites of Action of Opioid Peptides

Opioids act at two sites on the neurons, *viz.* the presynaptic nerve terminal and the postsynaptic neuron. The postsynaptic actions of opioids are usually inhibitory, while presynaptically they inhibit the release of neurotransmitters. Thus, the central nervous system (CNS) effect of the opioids is an overall result of their actions at the multiple presynaptic sites of both inhibitory and excitatory neurons and also includes their actions at postsynaptic sites. As a consequence, the overall effect of opioids depends upon the location and density of opioid receptors on the neurons. For instance, presynaptic inhibition of neurotransmitter release leads to the excitatory effects in target neuron if the neurotransmitter produces an inhibitory effect. On the other hand, if the opioid also has a postsynaptic inhibitory effect on the target neuron, then excitatory effects may not occur (Winters et al. 2017; Fields and Margolis 2015). Morphine acts on μ -receptors and inhibits the discharge of numerous diverse neurotransmitters including acetylcholine, noradrenaline and the neuropeptide substance P (Kerage et al. 2019; Commons 2010; Dickenson 1994).

12.2.2.2 Opioid Peptides and Their Effect on Analgesic/Pain Pathways

Pain is usually considered to be associated with augmented activity in primary sensory neurons provoked either by strong thermal or mechanical stimuli or by the chemicals released by inflammation or tissue damage (Yam et al. 2018). The primary sensory neurons implicated in the pain sensation releases glutamate and substance P in the dorsal horn of the spinal cord principally (Zieglgänsberger 2018; Lembeck 2008). The nociceptive signal is then communicated *via* the spinothalamic tracts to the brain, thereby activating the descending pathways from the periaqueductal gray area in the midbrain which then exerts inhibitory control over the dorsal horn (Venkatraman et al. 2017; Mendell 2011).

Opioid receptors are located at various regions of the nervous system that are involved in pain transmission and control, including primary afferent neurons, spinal cord, midbrain, and thalamus. Though the physiological function of endogenous opioids toward the pain transmission is still unclear, under pathological conditions, the endogenous opioid system gets activated. The opioid drugs, however, produce analgesia by inhibition of neurotransmitter release from the primary afferent terminals in spinal cord and activation of descending inhibitory controls in the midbrain (Allouche et al. 2014; McDonald and Lambert 2005; Waldhoer et al. 2004).

Any modulation in the nociceptive pathways may result in profound alterations in levels of neurotransmitters in primary afferent neurons, thereby causing changes in sensitivity to opioid analgesia (Yam et al. 2018). Accordingly, neuropathic pain is related to reduced sensitivity to opioids; on the contrary, inflammatory pain is linked with increased opioid sensitivity. Moreover, the alterations that occur in pain sensitivity during the states of chronic pain have been ascribed to the activation of glutamate NMDA (*N*-Methyl-D-aspartate) receptor (Greenwald and Shafritz 2018; Latremoliere and Woolf 2009; Petrenko et al. 2003; Bennett 2000).

12.2.2.3 Activity of Opioid Peptides

Opioid peptides are produced by different tissues, including mainly pituitary and adrenal glands (Przewlocki 2013). Enkephalins, for instance, are also released by heart, skeletal muscle, kidney, and intestinal cells, thus playing an important role in behavior, pain, cardiac function, cellular growth, immunity, and ischemic tolerance (Denning et al. 2008). The expression of prodynorphin is mainly observed in the cerebral cortex and basal ganglia in addition to reproductive tissues of testis, uterus, and ovary (Collard et al. 1990; Douglass et al. 1987).

Opioid peptides are neuromodulators rather than neurotransmitters—that is, OPs can alter the release and neuronal response to neurotransmitter by changing the hyperpolarizing neuronal cell membrane (North and Williams 1983; Loose et al. 1990; Wu et al. 2007). The activation of ORs by endogenous peptides transmits or modulates signal transmission for other neurotransmitters. Therefore, the peptides have a wide range of activities that include central and peripheral antinociception, endocrine, immune, motor activity, feeding, sexual behavior, regulation of body temperature, respiration and cardiovascular and gastrointestinal functions (Przewlocki 2013).

The β -endorphin is regarded as the most important opioid peptide. The peptide is involved in interneuron communication through synapses as well as extracellular spaces and cerebrospinal fluid (volume transmission, VT) (Veening and Barendregt 2015). The peptide showed central and peripheral analgesic activities (Sprouse-Blum et al. 2010). Both ACTH and β -endorphin are stress hormones released during painful stimulations. The release of ACTH and β -endorphin from a culture of pituitary tumor cells can be evoked by epinephrine (Mains and Eipper 1981). Therefore β -endorphin is responsible for stress-induced analgesia (Rubinstein et al. 1996). The *N*-terminal amino acids of β -endorphin are likely accountable for analgesic activity, while the *C*-terminal amino acids are more related to potency. The removal of eight amino acids from *N*-terminal abolishes the analgesic activity of β -endorphin (Deakin et al. 1980). Although the analgesic potency of β -endorphin is higher than [Met]-enkephalin, the potency is reduced by *N*- α -acetylation (Deakin et al. 1980).

Enkephalins are another group of opioid peptides that include mainly [Met]-enkephalin and [Leu]-enkephalin. The affinity of enkephalins for MOR is similar to that of morphine and is ten times lower than affinity for DOR. Besides analgesia, enkephalins may be involved in emotional and motivational behavior (Nieto et al. 2005) and sexual activities for male (Rodríguez-Manzo et al. 2002) and controls gastrointestinal motility and secretions (Mitznegg et al. 1977; Holzer 2009).

Dynorphin is an opioid peptide which is released at the level of the spinal cord and augments the afferent pain signal (Podvin et al. 2016), thus provoking allodynia, pain sensation for usually non-painful stimuli. Therefore, dynorphin has a significant role in the mediation of chronic pain (Podvin et al. 2016) as well as tolerance to antinociceptive opioids (Vanderah et al. 2001). Moreover, dynorphin promotes anxiety, stress, and dysphoria-driven cravings for another dose in addiction through its action on KORs (Chavkin and Koob 2016; Knoll and Carlezon 2010).

Endomorphins (1 and 2) are reported to be the only discovered opioid peptide that are selective for MOR (Zadina et al. 1997). The two peptides differ from other opioid peptides in having $\text{NH}_2\text{-Tyr-Pro-Trp(Phe)-Phe-CO-NH}_2$, that is, having carboxamide (aminocarbonyl) terminal. Some studies showed that endomorphin-1 produces higher analgesia and without reward effect compared to morphine (Wilson et al. 2000). Although the reward effect is known to be mediated by binding MOR, it is limited to a specific conformational set of receptors; thus it is ligand-dependent property. However, the potential of respiratory depression, urinary retention, tolerance, addiction and cardiac side effects and the low intrinsic bioavailability restrict the clinical use of endomorphins (Gu et al. 2017). Glycosylation through succinamic acid linker at the *N*-terminal enhances metabolic stability and membrane permeability of endomorphins while maintaining potency and efficacy (Varamini et al. 2012).

Nociceptin is structurally related to dynorphin; however, it binds to opioid receptor-like (ORL) while having no affinity for other ORs. Similar to dynorphin, nociceptin antagonizes the analgesic effect of other opioid ligands (Mika et al. 2011). Activation of ORL inhibits adenylyl cyclase and Ca^{+2} channels while activating K^+ channels like opioid receptors (Calo et al. 2000). However, the pharmacological behavior observed upon activation by nociceptin produces potent anti-analgesic action supra-spinally and analgesic action spinally (Mogil and Pasternak 2001).

12.3 Receptor (Types, Subtypes, Localization, Down Signaling Pathways, Functions, Agonist, Antagonist)

Endogenous opioids produce effects on the neurons by binding to three types of receptors located on neuronal cell membranes: namely, mu (μ), delta (δ) and kappa (κ) receptors. Other receptors such as opioid receptor-like-1 (ORL1) have sequence similarity to ORs, however not classified as ORs since being unresponsive to classical opioids ligands (Snyder 2004). Naturally occurring opioids, β -endorphins interact preferentially with μ -receptors, while the enkephalins and dynorphin interact respectively with δ -receptors and κ -receptors (Ghelardini et al. 2015; Al-Hasani and Bruchas 2011; Dhawan et al. 1996). The relative selectivity for endogenous and exogenous ORs ligands are listed in Table 12.2. The crystal structures for opioid receptors are available for mu, delta, and recently kappa receptors. The structures are for inactive conformations that are stabilized by bound antagonists (Manglik et al. 2012; Wu et al. 2012; Granier et al. 2012) and for active conformations that are stabilized by either bound agonist or conformation-specific nanobody or G-protein (Huang et al. 2015; Koehl et al. 2018; Che et al. 2018). The further sorting of main OR classes into subtypes is only approved from the anesthetic perspective in a way that $\mu 1$ is responsible for analgesia and dependence, $\mu 2$ is for euphoria, dependence and respiratory depression, and $\mu 3$ is for vasodilation. However, currently, there is no clear evidence for the existence of subtypes for these receptors (Dietis et al. 2011).

Table 12.2 Selectivity of naturally occurring endogenous opioids and opiates for opioid receptors

	μ -receptor	δ -receptor	κ -receptor
<i>Opioid peptides (endogenous opioids)</i>			
β -endorphin	+++	+++	+++
[Leu]-enkephalin	+	+++	–
[Met]-enkephalin	++	+++	–
Dynorphin	++	+	+++
<i>Opioid drugs (agonists)</i>			
Morphine	+++	+	++
Codeine	+	+	+
Pethidine	++	+	+
Fentanyl	+++	+	–
<i>Opioid drugs (partial/mixed agonists)</i>			
Pentazocine	+	+	$\pm \pm$
Buprenorphine	$\pm \pm \pm$	–	–
<i>Opioid drugs (antagonists)</i>			
Naloxone	+++	++	++
Naltrexone	+++	++	++

+ indicates agonist; \pm indicates partial agonist; number of + or \pm indicates potency

Opioid receptors are classical G protein-coupled receptors (Fig. 12.2), thus interacting with intracellular G-protein that has GTPase activity to mediate intracellular signaling. The G-protein is a heterotrimeric protein composed of $G\alpha$, $G\beta$, and $G\gamma$. The activation of ORs leads to the dissociation of G-protein. The $G\beta\gamma$ subunits may interact directly with the membrane ion channels, thereby causing inhibition of voltage-gated calcium channels (VGCCs) or rectifying potassium channels (Kir). The $G\beta\gamma$ subunits also contribute to the activation of inositol triphosphate kinase (IP3K), phospholipase C—phosphatidyl inositol triphosphate (PLC-IP3), or mitogen-activated protein kinase (MAPK) (Khan et al. 2013; Smrcka 2008). With respect to $G\alpha$ subunit, it is classified into four families: $G\alpha_s$ (activates adenylyl cyclase), $G\alpha_{i/o}$ (inhibits adenylyl cyclase), $G\alpha_{q/11}$ (activation of phospholipase C β) and $G\alpha_{12/13}$ (activation of GTPase activating protein for Ras)(Ras, a small GTP-binding protein named after Rat sarcoma) (Neves et al. 2002). The inhibition of adenylyl cyclase by $G\alpha_{i/o}$ subunit leads to reduced production of cyclic adenosine monophosphate (cAMP). Decreased concentration of cAMP further modulates membrane sodium or calcium channels (Law 2011; Catterall 2011; Scheuer 2011). Additionally, cAMP may also interact with and modulate the inward rectifying potassium channels (Li et al. 2013; Butt and Kalsi 2006; North 1993). Chronic consumption of opiates inhibits the production of cAMP; however, this inhibition is offset in the long run by other cAMP production mechanisms (Ramaswamy and Langford 2017; Al-Hasani and Bruchas 2011; Kosten and George 2002). When no opiates are available, this increased cAMP production capacity comes to the force and results in neural hyperactivity, thus causing a sensation of craving the drug (Al-Hasani and Bruchas 2011; Bie 2005; Kosten and George 2002).

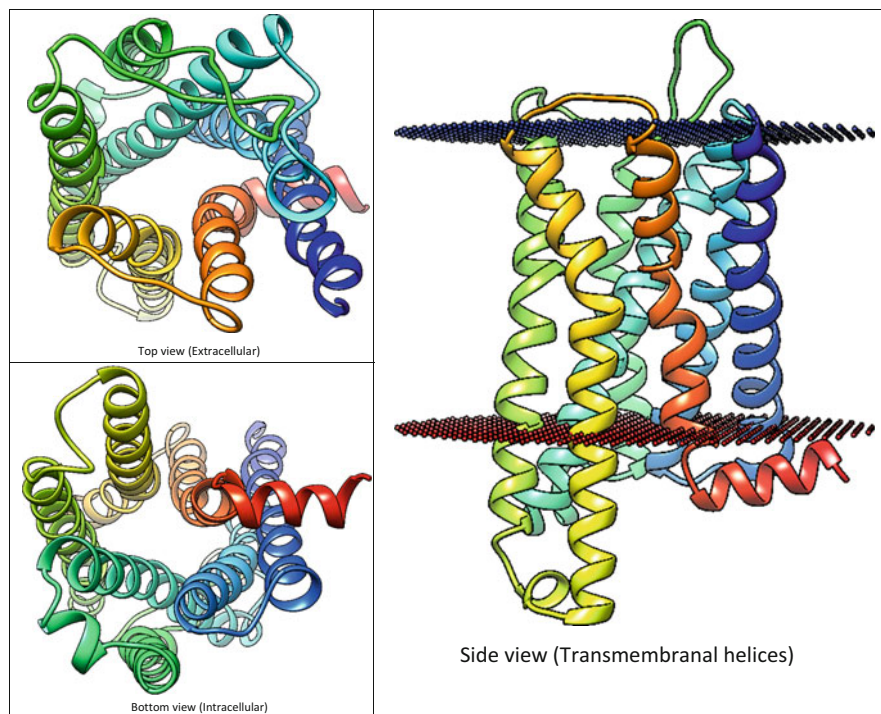


Fig. 12.2 Crystal structure of active conformation of Mu opioid receptors (PDB ID: 6DDE) with missing H8 being added from PDB 5C1M and the position within cell membrane is optimized using OPM database

Besides interaction with G-protein, ORs interact with other regulatory proteins and may end up with its sequestration or desensitization. Specific phosphorylation of ORs by GPCR kinases leads to interaction with β -arrestin and subsequent internalization of the receptor. Probably, $G_{i/o}$ mediates the antinociceptive effect produced by activation of MOR (Lamberts et al. 2011; Connor and Christie 1999), while respiratory depression and tolerance is mediated by activation of β -arrestin. Specific phosphorylation of ORs by GPCR kinases leads to interaction with β -arrestin and subsequent internalization of the receptor further causes tolerance to morphine analgesia (Bohn et al. 1999). Avoiding activation of β -arrestin-2 pathway reduces internalization of MOR and consequently tolerance; however, it has little effect on the up-regulation of adenylyl cyclase activity that is correlated to physical dependence (Bohn et al. 2000).

Opioid receptors may exist as homo- or heteromers to provide a cellular specific response for the same ligand. The ORL-1 or nociceptin/orphanin FQ (N/OFQ) receptor undergoes heteromerization to form heteromers with μ -opioid receptor (MOR), δ -opioid receptor (DOR), and κ -opioid receptor (KOR) (Evans et al. 2010). Additionally, MOR may also form heteromers with DOR in small dorsal

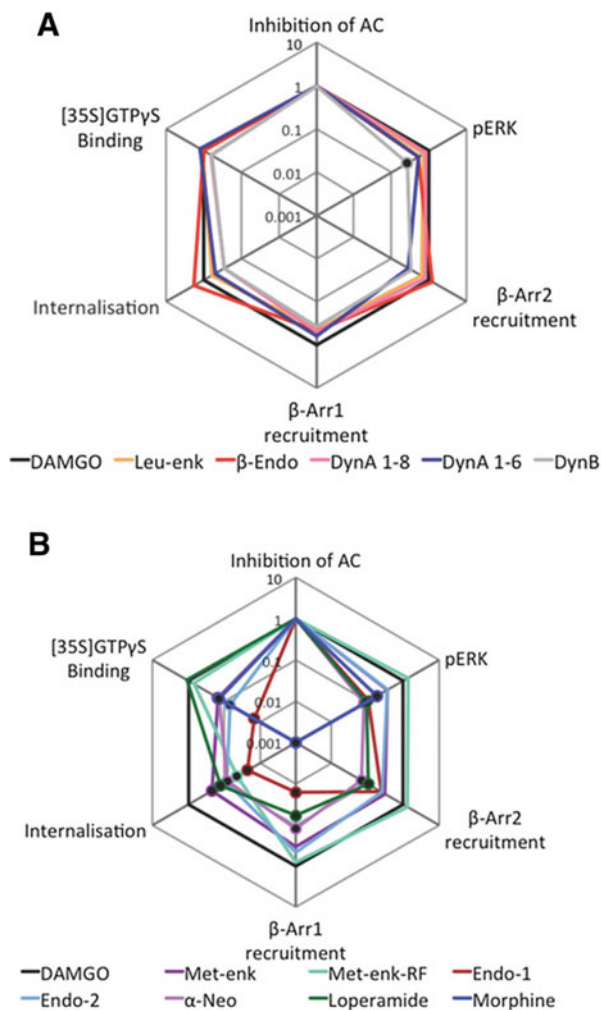
root ganglion neurons (DRGn) and in homologous expression systems (Gendron et al. 2015).

All three ORs produce analgesia when opioid binds to them; however, it is higher for μ than κ and δ subtypes and is associated with euphoria, dysphoria, and anxiolytic effects, respectively (Lutz and Kieffer 2013). Activation of κ -receptors produces comparatively less physical dependence as activation of μ -receptors (Machelska and Celik 2018; Feng et al. 2012). Each of these receptors is coupled to intracellular mechanisms *via* G-protein and mediates its effect through the second messengers, thereby influencing the probability of opening of the ion channels, which in some instances may reduce the excitability of the neurons (Machelska and Celik 2018, Feng et al. 2012). This reduced excitability contributes as a potential source of the euphoric effect of opiates and probably appears to be mediated by μ - and δ -receptors (Machelska and Celik 2018; Reisine and Bell 1993).

Like many GPCRs, ORs have binding sites for orthosteric and allosteric ligands. While orthosteric ligand can modulate receptor conformation independently, allosteric ligand depends on orthosteric ligand for modulating receptor conformation. The orthosteric active opioid peptides (Livingston and Traynor 2018) share a common *N*-terminal sequence of Tyr-Gly-Gly-Phe that starts with an ionized amine. The peptides are proposed to have two pharmacophores named as message and address pharmacophores. In other words, the address leads the peptide for opioid receptor subtype, while the message stabilizes particular receptor conformations to mediate specific set of intracellular signals. This proposal has been used to design highly potent and selective non-peptide DOR antagonists (Portoghese et al. 1988). The extracellular loops of opioid receptors have long been thought to involve in controlling ligand selectivity to receptor subtypes by interaction with the address part of the ligand (Metzger and Ferguson 1995). On the other hand, allosteric ligands (or modulators) are classified as positive (PAM), negative (NAM) and neutral/silent allosteric (SAM) modulators, which increases, decreases, and has no effect on orthosteric ligand activity, respectively (Mahmod Al-Qattan and Mordi 2019). Most of the allosteric ligands require bound orthosteric ligand in order to exert activity.

The conformational ensembles stabilized by orthosteric \pm allosteric ligand(s) modulate the downstream signaling of the receptor (i.e. interaction with intracellular second messengers). Therefore, any ligand binding to ORs has a 2D fingerprint of intracellular signaling paradigm as affinities versus efficacies. Accordingly, some ligands are biased toward activating a particular group of second messengers over others, which is referred to as biased agonism. Biased signaling (also known as differential efficacy or functional selectivity) is the way of separating wanted pharmacological effect from other unwanted effects mediated by the same receptor (Bologna et al. 2017). The biased activation of ORs was observed among endogenous as well as exogenous ligands. Unlike other endogenous OPs, α -neoendorphin, [Met]-enkephalin, [Met]-enkephalin-RF, endomorphin-1 and endomorphin-2- showed biased agonistic activities compared to reference DAMGO (a synthetic peptide with high μ -opioid receptor specificity and its structure is H-Tyr-D-Ala-Gly-N-MePhe-Gly-ol) across multiple signaling pathways as shown in Fig. 12.3 (Thompson et al. 2015).

Fig. 12.3 Webs of bias of endogenous opioid peptides and reference ligands at the MOP. **(a)** Ligands with profiles similar to DAMGO. **(b)** Ligands with profiles that differ from that of DAMGO. The score of τ/K_A (or efficacy/affinity) values was normalized to the reference ligand DAMGO and to the cAMP assay. Statistically significant differences ($P \leq 0.05$) are denoted by black circles as determined by two-tailed t test. For the purposes of visualization only, a τ/K_A for Met-enk-RF in the internalization assay was estimated using the incomplete concentration response curve for plasma membrane marker (Thompson et al. 2015)



With respect to exogenous ligands, biased activation of MOR toward G-protein was observed for a fungal peptide, which is opposite to the biased activity produced by endogenous endomorphin-2 that is toward β -arrestin-2 (Dekan et al. 2019). Biased agonist at MOR opens a new opportunity of agonists with lower side effects (Madariaga-MazÓN et al. 2017). Oliceridine is a biased agonist at MOR, which provides lower β -arrestin binding leading to lower receptor internalization and thus lower respiratory depression and tolerance compared to unbiased morphine agonist (Dewire et al. 2013). Similar natural products such as mitragynine and 7-hydroxymitragynine (Kruegel et al. 2016) and salvinorin-analogues (Groer et al. 2007) also showed a biased effect with lower β -arrestin binding. The PZM21, a molecule obtained by structure-based screening and design, showed selective MOR agonistic effect with minimal β -arrestin activation (Manglik et al. 2016). Biased agonist at KOR was developed to produce antinociception and anti-itching activities

without the concomitant dysphoria and sedation usually associated with this receptor (Brust et al. 2016). Several biased agonists at KOR were successfully developed, and some are being used clinically (Mores et al. 2019; Brust et al. 2016). Despite the previous successes, there are unresolved experimental limitations of measuring differences in biased factor among various ligands in addition to the implications of cellular environments (Mores et al. 2019; Ho et al. 2018). Although beneficial results observed by using biased agonists to stabilize receptor conformational ensembles that said to activate intracellular G α i over β -arrestin-2 recruitment (Ranjan et al. 2017), such simplification of the story might not be precise (Conibear and Kelly 2019; Bermudez et al. 2019).

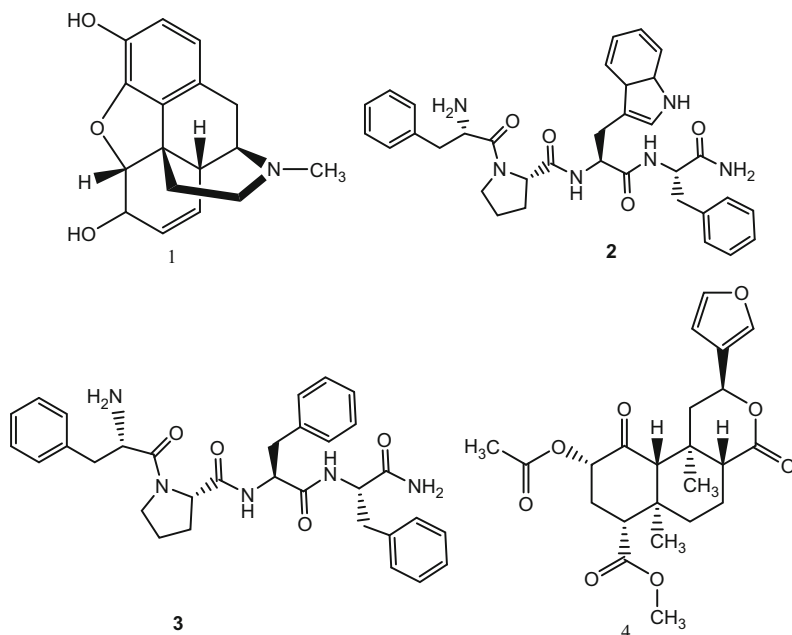
Like other GPCRs, the allosteric sites of ORs are therapeutically promising to get biased signaling (Livingston and Traynor 2018; Livingston et al. 2018). Moreover, allosteric ligands can enhance efficacy/affinity for endogenous opioid peptides as well as provide selectivity toward particular OR type, which usually is difficult to achieve using orthosteric ligands (Livingston et al. 2018). Interestingly, some endogenous compounds act as PAM for ORs, for example, oxytocin which is a peptide hormone principally involved in labor and lactation act as PAM for orthosteric endomorphin-1, β -endorphin, and morphine by enhancing efficacy and not affinity toward MOR (Meguro et al. 2018). The tuber of the species *Aconitum* is traditionally used in Japan to relieve pain, currently shown to have ignavine, that has selectivity for MOR over KOR and acts as PAM for endomorphin-1 and morphine (Ohbuchi et al. 2016). Biased signaling can also be produced using allosteric modulators. The BMS-986187, a synthetic compound discovered by high-throughput screening (HTS), can function as PAM selectively for DOR (Burford et al. 2015) and produce biased signaling toward G-protein over β -arrestin-2, which is elicited by lower receptor internalization (Stanczyk et al. 2019).

12.4 Opioid System as Potential Target for Neurological Disorders

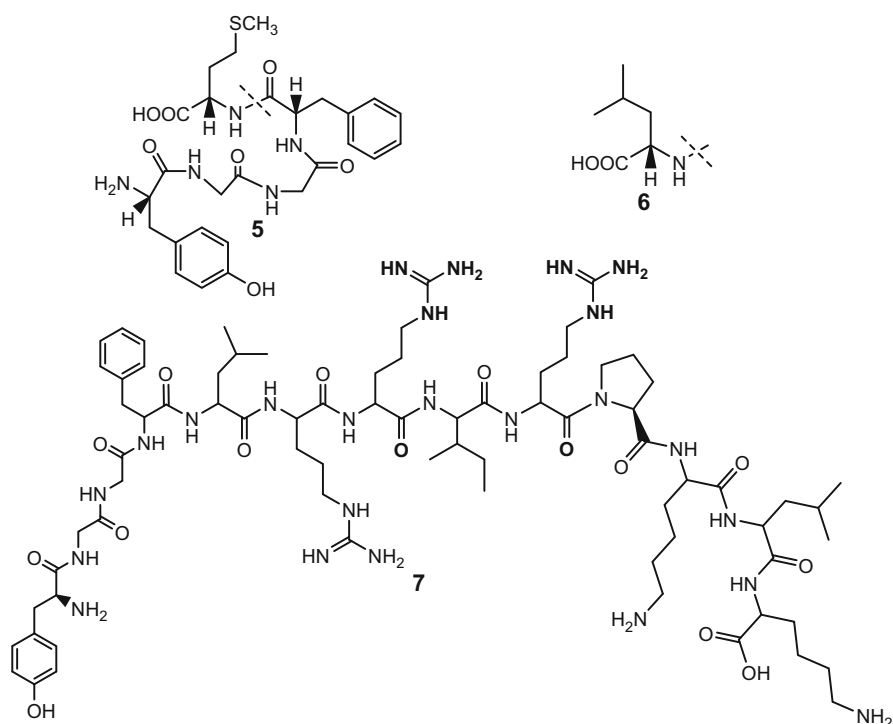
The ongoing developments, including a plethora of available drugs and various promising investigational molecules, suggest the potentiality of opioid receptors as viable drug targets for therapeutic interventions of numerous disorders. Targeting and modulating these molecular switches mediates the alleviation of Alzheimer's disease (AD) (Torres-Berrio and Nava-Mesa 2019), mood disorder (Lutz and Kieffer 2013; Lalanne et al. 2014; Ehrich et al. 2014; Browne and Lucki 2019) and psychiatric disorders (Tejeda et al. 2011; Carlezon and Krystal 2016; Guerrero et al. 2019) and treatment of alcohol and opioid use disorder (AUD and OUD) (Niciu and Arias 2013; Schuckit 2016). Besides, the target-specific treatment intervention related to bowel disorders (Lacy et al. 2016; Corsetti et al. 2019), chronic pain (Günther et al. 2017; Markman et al. 2019), ischemic stroke (Wang and Subedi 2020) and respiratory disorders (Zebraski et al. 2000) have also been demonstrated amongst others. This section focuses on the current updates on targeting the opioidergic system in an effort to treat neurological disorders.

12.4.1 Alzheimer's Disease (AD)

Numerous evidential researches suggest the possible association of the opioidergic system and pathogenesis of AD. The opioid receptors are innervated extensively in the specific region of the central nervous system that is vital for cognition and memory and includes hippocampus and cortex. Any abnormality within the opioid system homeostasis may result in the hyperphosphorylation of *tau* proteins and subsequent generation of amyloid-beta ($A\beta$) proteins. This event cause neuroinflammation followed by cholinergic neurons' deterioration and impairment of cognitive functions (Mathieu-Kia et al. 2001). Alterations in the level of G-protein coupled opioid receptors (μ , δ , and κ) indicate a significant association of the endogenous opioid in AD pathology (Nandhu et al. 2010). The ORs can modulate the imbalance of various neurotransmitters of cholinergic, adrenergic, GABAergic, serotonergic, and glutaminergic systems that are involved with the progression of AD (Cai and Ratka 2012). Studies on morphine (1) and endogenous opioids such as endomorphin-1 (2) and endomorphin-2 (3) showed substantial protection against intracellular $A\beta$ toxicity in both human and rat brains *in vivo*. Morphine mediated its activity by stimulating estradiol release in the hippocampus and increases the activity of P450 cytochrome aromatase. This chain of events stimulates the heat shock protein 70 (Hsp70) and confers protection against neurodegeneration (Cui et al. 2011). Furthermore, morphine attenuates the $A\beta$ induced neurotoxicity by activation of MOR and upregulates the mammalian target of rapamycin (mTOR) signaling as evaluated by cell viability and neurite outgrowth assay (Wang et al. 2014).



Unlike morphine, salvinorin A (**4**), a novel non-nitrogenous hallucinogenic neoclerodane diterpene isolated from *Salvia divinorum* (Roth et al. 2002), exhibited potent hallucinogenic property by selective agonism at κ -opioid receptor (KOR). This activity opens up new avenues and insights for the development of selective KOR antagonists. Targeting and blocking KOR will reverse the hallucinosis and altered perception as observed in AD and Pick's and Huntington's diseases (Cunningham et al. 2011; Sheffler and Roth 2003). Besides, salvinorin A also displayed modulatory activity on cholinergic systems and may signify a valuable molecular entity for alleviating the progression of AD (Motel et al. 2013). Research on an elevated level of dynorphin A (**5**) (Yakovleva et al. 2007) and enkephalins, namely [Met]-enkephalin (**6**) and [Leu]-enkephalin (**7**), observed that they induce neurodegeneration in transgenic mouse models and AD patients (Meilandt et al. 2008).



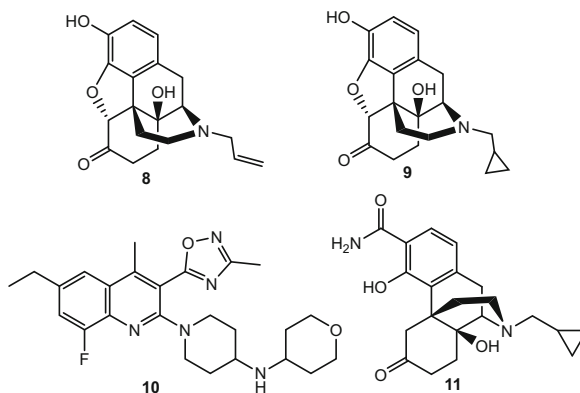
The dynorphins were profoundly expressed during the aging process and AD and have been reported to stimulate the KOR and thought to induce stress-related memory impairments. Additionally, they also affect glutamate neurotransmission and perturb their function of synaptic plasticity essential for memory (Ménard et al. 2013).

The rate-determining step in the production of A β is the proteolytic breakdown of Amyloid Precursor Protein (APP) by β -secretase (BACE1). The activation of DOR

by an agonist has been shown to increase the secretase activity probably through the post-translational mechanism and causing an increased formation of A β (Sarajarvi et al. 2015). The cascade of reaction makes DOR and BACE1 as prospective targets in the design of DOR antagonist (Zhao et al. 2015) and BACE1 inhibitors (Coimbra et al. 2018) for ameliorating the neurodegeneration underlying AD. The hypermethylation of opioid receptor δ 1 (OPRD1) promoter was also linked with the risk of AD (Ji et al. 2017). Subsequent studies suggest that in addition to OPRD1, elevated methylation of opioid receptor κ 1 and opioid receptor μ 1 genes are also involved in the progression of AD. Therefore, the genes of the opioid receptors could serve as potential biomarkers for AD diagnosis (Xu et al. 2018).

12.4.2 Schizophrenia

Schizophrenia is a multifaceted, diverse mental disorder that affects the behavior and cognitive function and has genetic or environmental predisposition, or both. Antipsychotics, along with psychological therapies, are the primary line of management available to alleviate the disorder. In recent years, much research to gain insight into the pathophysiology of schizophrenia has been undertaken (Owen et al. 2016; Patel and Shulman 2015) and identification of novel targets is under investigation (Gill et al. 2018; Yang and Tsai 2017). The cardinal features of schizophrenia are negative symptoms (reduced enthusiasm and withdrawal from society), cognitive symptoms (disruption of attentiveness and dementia), and positive symptoms (hallucinations accompanied by delusions). KOR agonists could elicit these specific symptoms and drugs blocking this receptor might result in a fruitful therapeutic outcome. Antipsychotics are capable of effectively combating the positive symptoms; however, presently, efficient therapeutic managements for controlling the symptoms (negative or cognitive) of schizophrenia are not available. The potentiality of a pan-opioid antagonist either naloxone (**8**) or naltrexone (**9**) could make KOR a promising target option for overall treatment benefits in schizophrenia (Clark and Abi-Dargham 2019; Shekhar 2019). Alongside, contemporary research on the discovery of novel KOR antagonist by Guerrero et al. has shown encouraging results. The promising drug candidate BTRX-335140/CYM-53093 (**10**) exhibited potent (IC_{50} -0.8 nM) and selective KOR antagonistic activity with a favorable pharmacokinetic profile. Currently, the compound is undergoing phase I clinical trials for the possible treatment of various psychiatric disorders (Guerrero et al. 2019). A recent double-blind phase II study found that a combination of olanzapine and fixed dose of MOR antagonist samidorphan (**11**) demonstrated clinically and statistically significant reduction of weight gain and adverse metabolic effect of olanzapine without compromising the antipsychotic efficacy of olanzapine. The combination was considerably tolerated and comparable to that of olanzapine-placebo in terms of safety (Chaudhary et al. 2019; Martin et al. 2019).

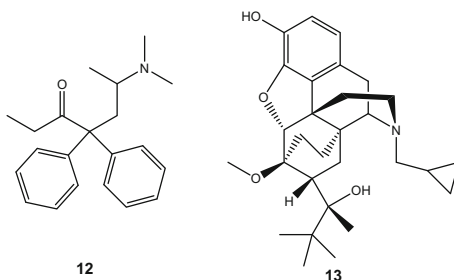


12.4.3 Post-traumatic Stress Disorder (PTSD), Opioid Use Disorder (OUD), and Alcohol Use Disorder (AUD)

Post-traumatic stress disorder (PTSD) is a psychiatric disorder that may arise after a single encounter or exposures to life-threatening chronic events. PTSD deteriorates physical health and is mostly accompanied by cardiorespiratory, musculoskeletal, gastrointestinal, immunological, endocrine, and metabolic problems. It is also associated with psychiatric comorbidity and an increased suicidal tendency (Bisson et al. 2015; Yehuda et al. 2015). Existing approaches in the treatment of PTSD involve cognitive behavioral therapy and the use of anxiolytic/antidepressant agents to ameliorate the symptoms (Shalev et al. 2017). Interestingly, studies suggest that endogenous opioid peptides exhibited a placebo effect in PTSD, and the mood-enhancing effects of the peptides may be initiated by exercise and light therapy to relieve the stress. It is suggested that the interaction between dopaminergic pathways and the endogenous opioids may be responsible for the placebo effect (Sher 2004), although it would not be a stand-alone option and may be concomitantly utilized along with standard drugs. As discussed earlier, the dynorphin mediates its action *via* KOR and the dynorphin/KOR interrelationship is associated in several brain disorders (De Lanerolle et al. 1997; Mathieu-Kia et al. 2001; Mello and Negus 2006). Various works of the literature suggest that in PTSD, there is a substantial expression of the KOR and mediate the symptoms of anxiety. Therefore, in line of the evidence, targeting the KOR might be a viable option in the management of PTSD (Bailey et al. 2013). The opioid analgesics prescribed in PTSD often result in comorbidity between PTSD and OUD, and they are frequently considered as two sides of the same coin (Elman and Borsook 2019; Hassan et al. 2017).

Centrally acting competitive MOR antagonist opioid receptor antagonists such as naloxone is the ideal choice in emergencies related to opioid overdose. On the contrary, naltrexone, which mediates its action *via* KOR antagonism, is employed mainly in OUD and AUD for maintaining abstinence by decreasing the cravings

(Theriot et al. 2019). However, both opioid agonist and antagonist are utilized in substance abuse therapies to combat the withdrawal syndromes and for the inhibition of return usage. Agonists such as morphine and methadone (**12**), partial agonist buprenorphine (**13**) and opioid antagonist such as extended-release injectable naltrexone are recommended for overall treatment and tackling the relapsing of OUD. The mechanism by which opioid antagonist maintain abstinence in OUD and AUD is by reducing the mesolimbic dopaminergic neurotransmission (McCarty et al. 2018; Williams et al. 2008).

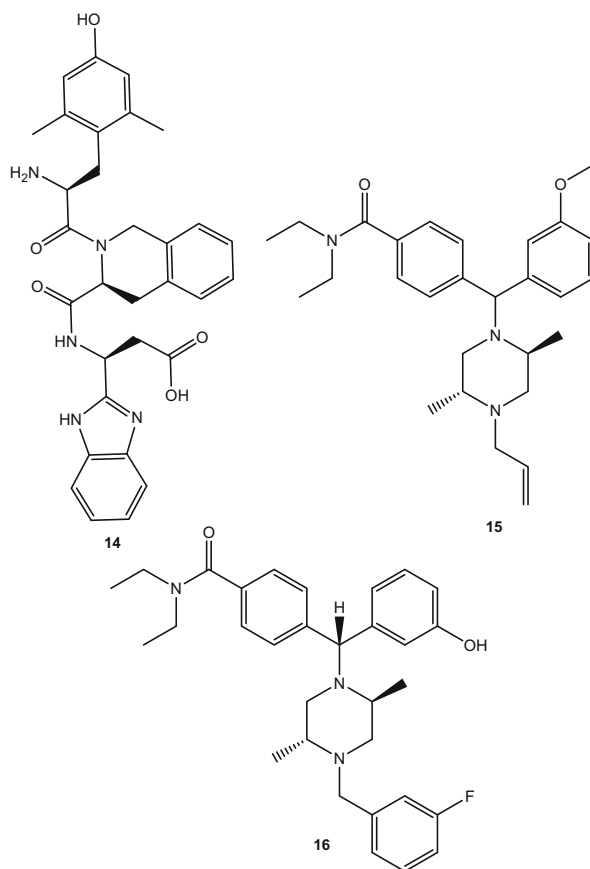


12.4.4 Parkinson's Disease (PD)

It is a chronic neurodegenerative disease having marked motor (rigidity, tremor, bradykinesia, firmness, and defective gait) and nonmotor features (hyposmia, sleep disorders, depression, etc.) (Schapira et al. 2017; Xia and Mao 2012). The disease is related to formation of Lewy bodies and dopaminergic neuronal damage in the substantia nigra. The main challenge in the treatment of Parkinson's disease (PD) is the failure to make a conclusive diagnosis at the earliest stages and difficulties in late-stage management of symptoms. Presently, there are no effective treatments for slowing down the neurodegenerative process and involve significant physical and mental co-morbidity (Demaagd and Philip 2015; Kalia and Lang 2015). The currently available pharmacotherapeutic options in PD are dopamine precursor (first line), dopamine agonist and monoamine oxidase B inhibitors (second line), the antiviral drug amantadine (third line) and a newer United States Food and Drug Administration (USFDA)-approved second-generation antipsychotic pimavanserin (Young and Mendoza 2018). The increasing perspective of the opioid receptors as a promising target in numerous brain disorders is well established. The treatment with levodopa is almost always associated with dyskinesia. Recent data suggest that selective opioid antagonists (MOR and DOR) can efficiently improve the dyskinesic side effect in animal models (Pan and Cai 2017; Sgroi and Tonini 2018).

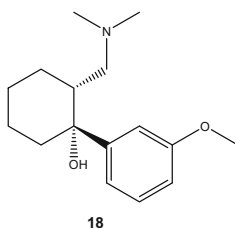
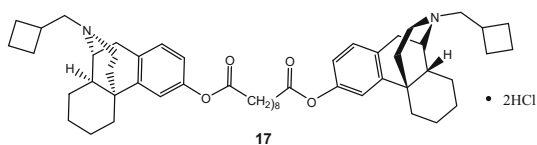
Nevertheless, a study on DOR agonist UFP-512 to mitigate motor insufficiencies in hemilesioned rodents resulted in a mixed response. At a low dose involving rotarod test, it largely improved the gait, whereas at high dose UFP-512 (**14**) was found inefficacious or to exacerbate the symptoms of Parkinsonism. Further, when

locally microinjected in the globus pallidus (GP), increased akinesia was observed, and vice versa, when injected in the substantia nigra pars reticulata. Adverse effects such as convulsions may restrict the use of DOR agonist in Parkinsonism. The convulsion was avoided by a synergistic combination of a DOR agonist, SNC-80, (**15**) and J-113397, an N/OFQ antagonist (Mabrouk et al. 2009; Mabrouk et al. 2014). Conversely, UF-512 showed encouraging activities as anxiolytic/antidepressant and treatment against chronic and neuropathic pain (Polo et al. 2019; Vergura et al. 2008). Recently, a new mixed DOR agonist/MOR antagonist, DPI-289 (**16**), in combination with levodopa elicited improved activity without increasing dyskinesia, and it was superior when compared to high dose levodopa (Johnston et al. 2018).



12.4.5 Mood Disorder

Mental illness such as bipolar disorder (BD) and major depressive disorder (MDD) are recalcitrant to treat due to the chronic nature and due to inter-individual variation (Jeon et al. 2016). Studies found that opioid analgesic demonstrated potent mood-elevating effect on patients with bipolar disorder and involve positive interaction between the opioid and the dopaminergic systems (Schaffer et al. 2007). Preclinical evaluations revealed that the dynorphin system is related to mood, motor, cognitive, and endocrine functionality and subjects with MDD and BD showed a decreased level of prodynorphin mRNA expression (Hurd 2002). Besides, a novel KOR antagonist MCL-144B (17) displayed antidepressant activity in the forced swim test (Reindl et al. 2008). Berrocoso et al. reported that a combination of a selective serotonin reuptake inhibitor (SSRI) with a weak MOR agonist, (+)-tramadol (18), produced better antidepressant activity than SSRI alone (Berrocoso and Mico 2009). As previously discussed, UF-512, a DOR agonist, showed anxiolytic/antidepressant properties (Polo et al. 2019). However, a combination of opioid-based samidorphan (MOR antagonist) and buprenorphine (ALKS 5461) in phase III trials was recently rejected by USFDA because of inadequate data to prove its effectiveness.



12.5 Effect of Opioids on Various Ion Channels

The body of human beings normally generates substances similar to opiates and utilizes them as neuromodulators. These opiate-like substances comprise of β -endorphins, dynorphins, and enkephalins, and are frequently conjointly called as opioid peptides or endogenous opioids (Corder et al. 2018; Li et al. 2012; Pathan and Williams 2012). Opioid peptides are implicated in the regulation and/or control of various body functions such as tolerance and drug dependency, stress and pain,

cognition, immunological, muscle-related, cardiovascular (CVS) and endocrine. Not only these, endogenous opioids also play a vital role in monitoring various sensory functions (Bodnar 2018). Opioids are enormously expressed in several parts of the brain including non-neuronal tissues as well, namely central nervous system (CNS) and peripheral nervous system (PNS). Within the CNS, opioids exert their actions in spinal cord; while in PNS, they have been found to act not only in myenteric but also in submucous plexus situated within the stomach wall and are accountable for producing vigorous constipation. Besides, opioids have been entailed in reduction of pain stimuli and inflammation in several peripheral tissues like joints (Iwaszkiewicz et al. 2013).

12.5.1 Calcium Channels

The entry of Ca^{2+} ions through the voltage-gated Ca^{2+} channels (VGCCs) results in depolarization of nerve terminals that further causes the discharge of neurotransmitters from the nerve cells. Three types of voltage-gated Ca^{2+} channels are reported, namely, T-type channels which are capable of showing small conductance, N-type channels which are inept to demonstrate intermediate conductance, and L-type channels which illustrate large conductance. Opioids act through the inhibition of N-type voltage-gated Ca^{2+} channels and reduce the passage of Ca^{2+} ions inside the cell, thereby inhibiting the release of neurotransmitters (Zamponi et al. 2015; Seseña et al. 2014; Catterall et al. 2013; Zamponi and Currie 2013). However, this action of opioids exclusively is not accountable for the total cumulative effect of opioids on neurotransmitter release (Chiang and Bekkers 2001).

12.5.2 GABA Channels

The euphoric effect of opioids may be due to another mechanism in which the GABA inhibitory interneurons of the ventral tegmental area (VTA) are involved (Listos et al. 2019; Creed et al. 2014; Xi 2002). By attaching to the μ -receptors, the exogenous opioids like morphine and heroine decrease the amount of GABA (a neurotransmitter) released. More often than not, GABA reduces the amount of dopamine released in the nucleus accumbens (NAcc). Hence, by inhibiting GABA, the opiates eventually increase the concentration of dopamine produced and consequently the amount of pleasure felt (Dubhashi 2018; Nuechterlein 2016; Shirayama and Chaki 2006). Opiates also have dopamine-independent effects within the NAcc, which play an important role in opiate reward (Ting-A-Kee and Van Der Kooy 2012; Tomkins and Sellers 2001; O'malley et al. 1992; Shippenberg and Elmer 1998; Koob and Bloom 1988).

Besides this, the periaqueductal gray (PAG) in the midbrain region being rich in endogenous opioids and opioid receptors is a major target of analgesic action in CNS (Tsagareli et al. 2012; Pathan and Williams 2012; Mansour et al. 1995). The analgesic action of opioids on PAG is exerted by the suppression of inhibitory

influence of neurotransmitter GABA on neurons that form part of a descending antinociceptive pathway (Tsagareli et al. 2012; Basbaum and Fields 1984). Opioids inhibit GABA-mediated (GABAergic) synaptic transmission in the PAG and other brain regions by reducing the probability of presynaptic neurotransmitter release (Wilson-Poe et al. 2017; Vaughan and Christie 1997), but the mechanisms involved remain uncertain. Literatures have reported that opioid inhibition of GABAergic synaptic currents in the PAG is controlled by a presynaptic voltage-dependent potassium conductance. Opioid receptors of μ -type in GABAergic presynaptic terminals are specifically coupled to this potassium conductance by a pathway involving phospholipase A₂, arachidonic acid and 12-lipoxygenase. Additionally, opioid inhibition of GABAergic synaptic transmission is also found to be potentiated by inhibitors of the enzymes cyclooxygenase and 5-lipoxygenase, presumably because more arachidonic acid is available for conversion to 12-lipoxygenase products (Zhang and Pan 2011; Heinke et al. 2011; Finnegan et al. 2006; Ingram et al. 1998). These mechanisms account for the analgesic action of cyclooxygenase inhibitors in the PAG and their synergism with opioids (Leith et al. 2007; Vaughan 1998; Vaughan et al. 1997; Tortorici and Vanegas 1995; Maves et al. 1994).

12.5.3 Sodium Channels

Voltage-gated sodium channel plays a critical role in nociception by interacting with the δ -opioid receptor. The dorsal root ganglia (DRG) neurons being rich in voltage-gated sodium channels (Wang et al. 2011; Rush et al. 2007; Wang and Wessendorf 2001; Zhang et al. 1997) can be correlated with the emergence of pain-related behavior characteristic of painful diabetic neuropathy (PDN). Activation of presynaptic δ -opioid receptor by enkephalin prevents the increase in neuronal Na⁺ in DRG through inhibition of protein kinase C (PKC) and p38 mitogen-activated protein kinase. This can be implicating presynaptic receptors of primary sensory afferents in modulating the amount of voltage-gated sodium channels and can be a useful therapy for PDN (Chattopadhyay et al. 2008).

12.6 Conclusion

Opioid peptides are endogenous ligands for opioid receptors. Proteolytic processing of larger precursor proteins generates the peptides. The peptides are stored in dense vesicles within neurons and released upon activation. The release is not restricted to synaptic space; thus peptides may signal other neurons by volume transmission. Opioid peptides inhibit the release of neurotransmitters by the affected neurons, thus modulating their signal propagations. The action of opioid peptides is mediated by binding GPCR group of receptors (opioid receptors) by binding to the orthosteric binding site. However, the affinity/efficacy of orthosteric ligand is affected by cooperation with ligand at allosteric site. Interestingly, many orthosteric and allosteric ligands show biased activation of intracellular second messengers, which

provide an opportunity of separating the desired pharmacological properties from a myriad of unwanted effects. Due to the interaction between opioid receptors and other types of receptors and ion channels, opioid peptides find a wide range of applications in managing several types of neurological disorders besides their primary use as analgesics.

12.7 Remark

The authors declare that theory of biological evolution and its related terms mentioned in this chapter and in references are not considered per se by them.

References

- Agnati LF, Bjelke B, Fuxe K (1995) Volume versus wiring transmission in the brain: a new theoretical frame for neuropsychopharmacology. *Med Res Rev* 15:33–45
- Al-Hasani R, Bruchas MR (2011) Molecular mechanisms of opioid receptor-dependent signaling and behavior. *Anesthesiology* 115:1363–1381
- Allouche SP, Noble F, Marie N (2014) Opioid receptor desensitization: mechanisms and its link to tolerance. *Front Pharmacol* 5:280
- Asvadi NH, Morgan M, Herath HM, Hewavitharana AK, Shaw PN, Cabot PJ (2014a) Beta-endorphin 1-31 biotransformation and cAMP modulation in inflammation. *PLoS One* 9:e90380
- Asvadi NH, Morgan M, Hewavitharana AK, Shaw PN, Cabot PJ (2014b) Biotransformation of beta-endorphin and possible therapeutic implications. *Front Pharmacol* 5:18–18
- Bailey CR, Cordell E, Sobin SM, Neumeister A (2013) Recent progress in understanding the pathophysiology of post-traumatic stress disorder. *CNS Drugs* 27:221–232
- Basbaum AI, Fields HL (1984) Endogenous pain control systems: brainstem spinal pathways and endorphin circuitry. *Annu Rev Neurosci* 7:309–338
- Bennett GJ (2000) Update on the neurophysiology of pain transmission and modulation: focus on the NMDA-receptor. *J Pain Symptom Manage* 19:2–6
- Bermudez M, Nguyen TN, Omieczynski C, Wolber G (2019) Strategies for the discovery of biased GPCR ligands. *Drug Discov Today* 24:1031–1037
- Berrococo E, Mico J-A (2009) Cooperative opioid and serotonergic mechanisms generate superior antidepressant-like effects in a mice model of depression. *Int J Neuropsychopharmacol* 12:1033–1044
- Bie B (2005) cAMP-mediated mechanisms for pain sensitization during opioid withdrawal. *J Neurosci* 25:3824–3832
- Bisson JI, Cosgrove S, Lewis C, Roberts NP (2015) Post-traumatic stress disorder. *BMJ* 351:h6161
- Bodnar RJ (2018) Endogenous opiates and behavior: 2016. *Peptides* 101:167–212
- Bohn LM, Lefkowitz RJ, Gainetdinov RR, Peppel K, Caron MG, Lin FT (1999) Enhanced morphine analgesia in mice lacking beta-arrestin 2. *Science* 286:2495–2498
- Bohn LM, Gainetdinov RR, Lin FT, Lefkowitz RJ, Caron MG (2000) Mu-opioid receptor desensitization by beta-arrestin-2 determines morphine tolerance but not dependence. *Nature* 408:720–723
- Bologna Z, Teoh JP, Bayoumi AS, Tang Y, Kim IM (2017) Biased G protein-coupled receptor signaling: new player in modulating physiology and pathology. *Biomol Ther (Seoul)* 25:12–25
- Browne CA, Lucki I (2019) Targeting opioid dysregulation in depression for the development of novel therapeutics. *Pharmacol Ther*. <https://doi.org/10.1016/j.pharmthera.2019.04.009>
- Brownstein MJ (1993) A brief history of opiates, opioid peptides, and opioid receptors. *Proc Natl Acad Sci U S A* 90:5391–5393

- Brust TF, Morgenweck J, Kim SA, Rose JH, Locke JL, Schmid CL, Zhou L, Stahl EL, Cameron MD, Scarry SM, AubÉ J, Jones SR, Martin TJ, Bohn LM (2016) Biased agonists of the kappa opioid receptor suppress pain and itch without causing sedation or dysphoria. *Sci Signal* 9:ra117
- Burford NT, Livingston KE, Canals M, Ryan MR, Budenholzer LM, Han Y, Shang Y, Herbst JJ, O'connell J, Banks M, Zhang L, Filizola M, Bassoni DL, Wehrman TS, Christopoulos A, Traynor JR, Gerritz SW, Alt A (2015) Discovery, synthesis, and molecular pharmacology of selective positive allosteric modulators of the delta-opioid receptor. *J Med Chem* 58:4220–4229
- Butt AM, Kalsi A (2006) Inwardly rectifying potassium channels (Kir) in central nervous system glia: a special role for Kir4.1 in glial functions. *J Cell Mol Med* 10:33–44
- Cai Z, Ratka A (2012) Opioid system and Alzheimer's disease. *Neuromol Med* 14:91–111
- Calo G, Guerrini R, Rizzi A, Salvadori S, Regoli D (2000) Pharmacology of nociceptin and its receptor: a novel therapeutic target. *Br J Pharmacol* 129:1261–1283
- Carlezon WA, Krystal AD (2016) Kappa-opioid antagonists for psychiatric disorders: from bench to clinical trials. *Depress Anxiety* 33:895–906
- Castro MG, Morrison E (1997) Post-translational processing of proopiomelanocortin in the pituitary and in the brain. *Crit Rev Neurobiol* 11:35–37
- Catterall WA (2011) Voltage-gated calcium channels. *Cold Spring Harb Perspect Biol* 3:a003947–a003947
- Catterall WA, Leal K, Nanou E (2013) Calcium channels and short-term synaptic plasticity. *J Biol Chem* 288:10742–10749
- Cawley NX, Li Z, Loh YP (2016) 60 years of POMC: biosynthesis, trafficking, and secretion of pro-opiomelanocortin-derived peptides. *J Mol Endocrinol* 56:T77–T97
- Chattopadhyay M, Mata M, Fink DJ (2008) Continuous δ -opioid receptor activation reduces neuronal voltage-gated sodium channel (NaV1.7) levels through activation of protein kinase C in painful diabetic neuropathy. *J Neurosci* 28:6652–6658
- Chaudhary AMD, Khan MF, Dhillon SS, Naveed S (2019) A review of samidorphan: a novel opioid antagonist. *Cureus*. <https://doi.org/10.7759/cureus.5139>
- Chavkin C, Koob GF (2016) Dynorphin, dysphoria, and dependence: the stress of addiction. *Neuropsychopharmacology* 41:373–374
- Che T, Majumdar S, Zaidi SA, Ondachi P, Mccorvy JD, Wang S, Mosier PD, Uprety R, Vardy E, Krumm BE, Han GW, Lee MY, Pardon E, Steyaert J, Huang XP, Strachan RT, Tribo AR, Pasternak GW, Carroll FI, Stevens RC, Cherezov V, Katritch V, Wacker D, Roth BL (2018) Structure of the nanobody-stabilized active state of the kappa opioid receptor. *Cell* 172:55–67. e15
- Chieng B, Bekkers JM (2001) Inhibition of calcium channels by opioid-and adenosine-receptor agonists in neurons of the nucleus accumbens. *Br J Pharmacol* 133:337–344
- Clark SD, Abi-Dargham A (2019) The role of dynorphin and the kappa opioid receptor in the symptomatology of schizophrenia: a review of the evidence. *Biol Psychiatry* 86:502–511
- Coimbra JRM, Marques DFF, Baptista SJ, Pereira CMF, Moreira PI, Dinis TCP, Santos AE, Salvador JAR (2018) Highlights in BACE1 inhibitors for Alzheimer's disease treatment. *Front Chem* 6:178
- Collard MW, Day R, Akil H, Uhler MD, Douglass JO (1990) Sertoli cells are the primary site of prodynorphin gene expression in rat testis: regulation of mRNA and secreted peptide levels by cyclic adenosine 3',5'-monophosphate analogs in cultured cells. *Mol Endocrinol* 4:1488–1496
- Commons KG (2010) Neuronal pathways linking substance P to drug addiction and stress. *Brain Res* 1314:175–182
- Conibear AE, Kelly E (2019) A biased view of mu-opioid receptors? *Mol Pharmacol* 96:542–549
- Connor M, Christie MJ (1999) Opioid receptor signalling mechanisms. *Clin Exp Pharmacol Physiol* 26:493–499
- Corder G, Castro DC, Bruchas MR, Scherrer G (2018) Endogenous and exogenous opioids in pain. *Annu Rev Neurosci*. <https://doi.org/10.1146/annurev-neuro-080317-061522>

- Corsetti M, Pannemans J, Whorwell P (2019) Targeting mu opioid receptors to modulate gastrointestinal function: what have we learnt so far from the studies in functional bowel disorders? *F1000Res* 8:257
- Coward P, Wada HG, Falk MS, Chan SDH, Meng F, Akil H, Conklin BR (1998) Controlling signaling with a specifically designed G_i-coupled receptor. *Proc Natl Acad Sci* 95:352–357
- Cox BM, Opheim KE, Teschemacher H, Goldstein A (1975) Purification and properties. *Life Sci* 16:1777–1782
- Cox BM, Goldstein A, Hi CH (1976) Opioid activity of a peptide, beta-lipotropin-(61-91), derived from beta-lipotropin. *Proc Natl Acad Sci U S A* 73:1821–1823
- Creed MC, Ntamati NR, Tan KR (2014) VTA GABA neurons modulate specific learning behaviors through the control of dopamine and cholinergic systems. *Front Behav Neurosci* 8:8
- Cui J, Wang Y, Dong Q, Wu S, Xiao X, Hu J, Chai Z, Zhang Y (2011) Morphine protects against intracellular amyloid toxicity by inducing estradiol release and upregulation of Hsp70. *J Neurosci* 31:16227–16240
- Cunningham CW, Rothman RB, Priszynano TE (2011) Neuropharmacology of the naturally occurring κ-opioid hallucinogen salvinorin A. *Pharmacol Rev* 63:316–347
- Day R (2009) Proopiomelanocortin. In: Squire LR (ed) *Encyclopedia of neuroscience*. Academic Press, Oxford
- De Lanerolle NC, Williamson A, Meredith C, Kim JH, Tabuteau H, Spencer DD, Brines ML (1997) Dynorphin and the kappa 1 ligand [3H]U69,593 binding in the human epileptogenic hippocampus. *Epilepsy Res* 28:189–205
- De Wied D (1999) Behavioral pharmacology of neuropeptides related to melanocortins and the neurohypophyseal hormones. *Eur J Pharmacol* 375:1–11
- Deakin JF, Doströvsky JO, Smyth DG (1980) Influence of N-terminal acetylation and C-terminal proteolysis on the analgesic activity of beta-endorphin. *Biochem J* 189:501–506
- Dekan Z, Sianati S, Yousuf A, Sutcliffe KJ, Gillis A, Mallet C, Singh P, Jin AH, Wang AM, Mohammadi SA, Stewart M, Ratnayake R, Fontaine F, Lacey E, Piggott AM, Du YP, Canals M, Sessions RB, Kelly E, Capon RJ, Alewood PF, Christie MJ (2019) A tetrapeptide class of biased analgesics from an Australian fungus targets the μ-opioid receptor. *Proc Natl Acad Sci U S A* 116:22353–22358
- Demaagd G, Philip A (2015) Parkinson's disease and its management: part 1: disease entity, risk factors, pathophysiology, clinical presentation, and diagnosis. *Pharm Ther* 40:504
- Denning GM, Ackermann LW, Barna TJ, Armstrong JG, Stoll LL, Weintraub NL, Dickson EW (2008) Proenkephalin expression and enkephalin release are widely observed in non-neuronal tissues. *Peptides* 29:83–92
- Dewire SM, Yamashita DS, Rominger DH, Liu G, Cowan CL, Graczyk TM, Chen XT, Pitis PM, Gotchev D, Yuan C, Koblisch M, Lark MW, Violin JD (2013) A G protein-biased ligand at the mu-opioid receptor is potently analgesic with reduced gastrointestinal and respiratory dysfunction compared with morphine. *J Pharmacol Exp Ther* 344:708–717
- Dhawan B, Cesselin F, Raghbir R, Reisine T, Bradley P, Portoghese PS, Hamon M (1996) International Union of Pharmacology. XII. Classification of opioid receptors. *Pharmacol Rev* 48:567–592
- Dickenson AH (1994). Where and how do opioids act?. In: Gebhart GF, Hammond DL, Jensen TS (eds) *Progress in pain research and management*. Vol. 2. Proceedings of the 7th World Congress on Pain, Seattle: IASP Press, Seattle
- Dietis N, Rowbotham DJ, Lambert DG (2011) Opioid receptor subtypes: fact or artifact? *Br J Anaesth* 107:8–18
- Douglass J, Cox B, Quinn B, Civelli O, Herbert E (1987) Expression of the prodynorphin gene in male and female mammalian reproductive tissues. *Endocrinology* 120:707–713
- Dubhashi J (2018) Analysis of the genetic and neurological components of opioid addiction, with public health perspectives of the opioid epidemic in the United States of America. *Discovery* 4:4

- Egleton RD, Mitchell SA, Huber JD, Janders J, Stropova D, Polt R, Yamamura HI, Hruby VJ, Davis TP (2000) Improved bioavailability to the brain of glycosylated Met-enkephalin analogs. *Brain Res* 881:37–46
- Ehrich E, Turncliff R, Du Y, Leigh-Pemberton R, Fernandez E, Jones R, Fava M (2014) Evaluation of opioid modulation in major depressive disorder. *Neuropsychopharmacology* 40:1448–1455
- Elman I, Borsook D (2019) The failing cascade: comorbid post traumatic stress- and opioid use disorders. *Neurosci Biobehav Rev* 103:374–383
- Evans RM, You H, Hameed S, Altier C, Mezghrani A, Bourinet E, Zamponi GW (2010) Heterodimerization of ORL1 and opioid receptors and its consequences for N-type calcium channel regulation. *J Biol Chem* 285:1032–1040
- Feng Y, He X, Yang Y, Chao D, Lazarus LH, Xia Y (2012) Current research on opioid receptor function. *Current Drug Targets* 13:230–246
- Fields HL, Margolis EB (2015) Understanding opioid reward. *Trends Neurosci* 38:217–225
- Finnegan TF, Chen S-R, Pan H-L (2006) μ opioid receptor activation inhibits GABAergic inputs to basolateral amygdala neurons through Kv1.1/1.2 channels. *J Neurophysiol* 95:2032–2041
- Froehlich JC (1997) Opioid peptides. *Alcohol health Res World* 21:132–135
- Gendron L, Mittal N, Beaudry H, Walwyn W (2015) Recent advances on the δ opioid receptor: from trafficking to function. *Br J Pharmacol* 172:403–419
- Geraciotti TD, Strawn JR, Ekhtor NN, Wortman M, Kasckow J (2009) 82 - Neuroregulatory peptides of central nervous system origin: from laboratory to clinic. In: PFAFF DW, Arnold AP, Etgen AM, Fahrbach SE, Rubin RT (eds) *Hormones, brain and behavior*, 2nd edn. Academic Press, San Diego
- Ghelardini C, Mannelli LDC, Bianchi E (2015) The pharmacological basis of opioids. *Clin Cases Miner Bone Metab* 12:219
- Gill BJA, Khan FA, Mckhann GM (2018) You're not hallucinating: potential new targets for schizophrenia treatment. *Neurosurgery* 84:E146–E147
- Granier S, Manglik A, Kruse AC, Kobilka TS, Thian FS, Weis WI, Kobilka BK (2012) Structure of the delta-opioid receptor bound to naltrindole. *Nature* 485:400–404
- Greenwald JD, Shafritz KM (2018) An integrative neuroscience framework for the treatment of chronic pain: from cellular alterations to behavior. *Front Integr Neurosci* 12:18
- Groer CE, Tidgewell K, Moyer RA, Harding WW, Rothman RB, Prisinzano TE, Bohn LM (2007) An opioid agonist that does not induce mu-opioid receptor – arrestin interactions or receptor internalization. *Mol Pharmacol* 71:549–557
- Gu ZH, Wang B, Kou ZZ, Bai Y, Chen T, Dong YL, Li H, Li YQ (2017) Endomorphins: promising endogenous opioid peptides for the development of novel analgesics. *Neurosignals* 25:98–116
- Guerrero M, Urbano M, Kim E-K, Gamo AM, Riley S, Abgaryan L, Leaf N, Van Orden LJ, Brown SJ, Xie JY, Porreca F, Cameron MD, Rosen H, Roberts E (2019) Design and synthesis of a novel and selective kappa opioid receptor (KOR) antagonist (Btrx-335140). *J Med Chem* 62:1761–1780
- Günther T, Dasgupta P, Mann A, Miess E, Kliewer A, Fritzwanker S, Steinborn R, Schulz S (2017) Targeting multiple opioid receptors – improved analgesics with reduced side effects? *Br J Pharmacol* 175:2857–2868
- Hassan AN, Foll BL, Intiaz S, Rehm J (2017) The effect of post-traumatic stress disorder on the risk of developing prescription opioid use disorder: results from the National Epidemiologic Survey on Alcohol and Related Conditions III. *Drug Alcohol Depend* 179:260–266
- Heinke B, Gingl E, Sandkühler J (2011) Multiple targets of μ -opioid receptor-mediated presynaptic inhibition at primary afferent A δ - and C-fibers. *J Neurosci* 31:1313–1322
- Herath HMDR, Cabot PJ, Shaw PN, Hewavitharana AK (2012) Study of beta endorphin metabolism in inflamed tissue, serum and trypsin solution by liquid chromatography–tandem mass spectrometric analysis. *Anal Bioanal Chem* 402:2089–2100
- Hersh LB, Rodgers DW (2008) Neprilysin and amyloid beta peptide degradation. *Curr Alzheimer Res* 5:225–231

- Ho JH, Stahl EL, Schmid CL, Scarry SM, Aube J, Bohn LM (2018) G protein signaling-biased agonism at the kappa-opioid receptor is maintained in striatal neurons. *Sci Signal* 11. <https://doi.org/10.1126/scisignal.aar4309>
- Hökfelt T, Broberger C, Xu Z-QD, Sergejev V, Ubink R, Diez M (2000) Neuropeptides — an overview. *Neuropharmacology* 39:1337–1356
- Holzer P (2009) Opioid receptors in the gastrointestinal tract. *Regul Pept* 155:11–17
- Huang W, Manglik A, Venkatakrisnan AJ, Laeremans T, Feinberg EN, Sanborn AL, Kato HE, Livingston KE, Thorsen TS, Kling RC, Granier S, Gmeiner P, Husbands SM, Traynor JR, Weis WI, Steyaert J, Dror RO, Kobilka BK (2015) Structural insights into micro-opioid receptor activation. *Nature* 524:315–321
- Hughes J, Smith T, Morgan B, Fothergill L (1975) Purification and properties of enkephalin — The possible endogenous ligand for the morphine receptor. *Life Sci* 16:1753–1758
- Hughes J, Beaumont A, Fuentes JA, Malfroy B, Unsworth C (1980) Opioid peptides: aspects of their origin, release and metabolism. *J Exp Biol* 89:239–255
- Hurd YL (2002) Subjects with major depression or bipolar disorder show reduction of prodynorphin mRNA expression in discrete nuclei of the amygdaloid complex. *Mol Psychiatry* 7:75–81
- Ingram SL, Vaughan CW, Bagley EE, Connor M, Christie MJ (1998) Enhanced Opioid Efficacy in Opioid Dependence Is Caused by an Altered Signal Transduction Pathway. *J Neurosci* 18:10269–10276
- Iwaszkiewicz KS, Schneider JJ, Hua S (2013) Targeting peripheral opioid receptors to promote analgesic and anti-inflammatory actions. *Front Pharmacol* 4:132
- Jeon HJ, Baek JH, Ahn Y-M, Kim SJ, Ha TH, Cha B, Moon E, Kang H-J, Ryu V, Cho C-H, Heo J-Y, Kim K, Lee H-J (2016) Review of cohort studies for mood disorders. *Psychiatry Investig* 13:265
- Ji H, Wang Y, Liu G, Chang L, Chen Z, Zhou D, Xu X, Cui W, Hong Q, Jiang L, Li J, Zhou X, Li Y, Guo Z, Zha Q, Niu Y, Weng Q, Duan S, Wang Q (2017) Elevated OPRD1 promoter methylation in Alzheimer's disease patients. *PLoS One* 12:e0172335
- Johnston TH, Versi E, Howson PA, Ravenscroft P, Fox SH, Hill MP, Reidenberg BE, Corey R, Brotchie JM (2018) DPI-289, a novel mixed delta opioid agonist / mu opioid antagonist (DAMA), has L-DOPA-sparing potential in Parkinson's disease. *Neuropharmacology* 131:116–127
- Kalia LV, Lang AE (2015) Parkinson's disease. *Lancet* 386:896–912
- Kerage D, Sloan EK, Mattarollo SR, McCombe PA (2019) Interaction of neurotransmitters and neurochemicals with lymphocytes. *J Neuroimmunol* 332:99–111
- Khan SM, Sleno R, Gora S, Zylbergold P, Laverdure J-P, Labbé J-C, Miller GJ, Hébert TE (2013) The expanding roles of Gβγ subunits in G protein-coupled receptor signaling and drug action. *Pharmacol Rev* 65:545–577
- Knoll AT, Carlezon WA (2010) Dynorphin, stress, and depression. *Brain Res* 1314:56–73
- Koehl A, Hu H, Maeda S, Zhang Y, Qu Q, Paggi JM, Latorraca NR, Hilger D, Dawson R, Matile H, Schertler GFX, Granier S, Weis WI, Dror RO, Manglik A, Skiniotis G, Kobilka BK (2018) Structure of the micro-opioid receptor-Gi protein complex. *Nature* 558:547–552
- Koob GF, Bloom FE (1988) Cellular and molecular mechanisms of drug dependence. *Science* 242:715–723
- Kosten T, George T (2002) The neurobiology of opioid dependence: implications for treatment. *Sci Pract Perspect* 1:13–20
- Kosterlitz HW, Hughes J (1975) Some thoughts on the significance of enkephalin, the endogenous ligand. *Life Sci* 17:91–96
- Kruegel AC, Gassaway MM, Kapoor A, VÁradi A, Majumdar S, Filizola M, Javitch JA, Sames D (2016) Synthetic and receptor signaling explorations of the mitragyna alkaloids: mitragynine as an atypical molecular framework for opioid receptor modulators. *J Am Chem Soc* 138:6754–6764

- Lacy BE, Mearin F, Chang L, Chey WD, Lembo AJ, Simren M, Spiller R (2016) Bowel disorders. *Gastroenterology* 150:1393–1407.e5
- Lalanne L, Ayranci G, Kieffer BL, Lutz P-E (2014) The kappa opioid receptor: from addiction to depression, and back. *Front Psychiatry* 5:170
- Lamberts JT, Jutkiewicz EM, Mortensen RM, Traynor JR (2011) mu-Opioid receptor coupling to Galpha(o) plays an important role in opioid antinociception. *Neuropsychopharmacology* 36:2041–2053
- Latremoliere A, Woolf CJ (2009) Central sensitization: a generator of pain hypersensitivity by central neural plasticity. *J Pain* 10:895–926
- Law PY (2011) Opioid receptor signal transduction mechanisms. In: Pasternak GW (ed) *The opiate receptors*. Humana Press, Totowa, NJ
- Leith JL, Wilson AW, Donaldson LF, Lumb BM (2007) Cyclooxygenase-1-derived prostaglandins in the periaqueductal gray differentially control C- versus A-fiber-evoked spinal nociception. *J Neurosci* 27:11296–11305
- Lembeck F (2008) The archeology of substance P. *Neuropeptides* 42:444–453
- Li CH, Chung D (1976) Isolation and structure of an untriakontapeptide with opiate activity from camel pituitary glands. *Proc Natl Acad Sci U S A* 73:1145–1148
- Li Y, Lefever MR, Muthu D, Bidlack JM, Bilsky EJ, Polt R (2012) Opioid glycopeptide analgesics derived from endogenous enkephalins and endorphins. *Future Med Chem* 4:205–226
- Li J, Blankenship ML, Bacceti ML (2013) Inward-rectifying potassium (Kir) channels regulate pacemaker activity in spinal nociceptive circuits during early life. *J Neurosci* 33:3352–3362
- Listos J, Łupina M, Talarek S, Mazur A, Orzelska-Górka J, Kotlińska J (2019) The mechanisms involved in morphine addiction: an overview. *Int J Mol Sci* 20:4302
- Livingston KE, Traynor JR (2018) Allostery at opioid receptors: modulation with small molecule ligands. *Br J Pharmacol* 175:2846–2856
- Livingston KE, Stanczyk MA, Burford NT, Alt A, Canals M, Traynor JR (2018) Pharmacologic evidence for a putative conserved allosteric site on opioid receptors. *Mol Pharmacol* 93:157–167
- Loh YP, Gainer H (1978) The role of glycosylation on the biosynthesis, degradation, and secretion of the ACTH-beta-lipotropin common precursor and its peptide products. *FEBS Lett* 96:269–272
- Loose M, Ronnekleiv O, Kelly M (1990) Membrane properties and response to opioids of identified dopamine neurons in the guinea pig hypothalamus. *J Neurosci* 10:3627–3634
- Luca G, Federico S, Roberto A (2007) Re-discussion of the importance of ionic interactions in stabilizing ligand-opioid receptor complex and in activating signal transduction. *Curr Drug Targets* 8:185–196
- Ludwig M, Leng G (2006) Dendritic peptide release and peptide-dependent behaviours. *Nat Rev Neurosci* 7:126–136
- Lutz PE, Kieffer BL (2013) Opioid receptors: distinct roles in mood disorders. *Trends Neurosci* 36:195–206
- Mabrouk OS, Marti M, Salvadori S, Morari M (2009) The novel delta opioid receptor agonist UFP-512 dually modulates motor activity in hemiparkinsonian rats via control of the nigro-thalamic pathway. *Neuroscience* 164:360–369
- Mabrouk OS, Viaro R, Volta M, Ledonne A, Mercuri N, Morari M (2014) stimulation of opioid receptor and blockade of nociceptin/orphanin FQ receptor synergistically attenuate Parkinsonism. *J Neurosci* 34:12953–12962
- Machelska H, Celik MÖ (2018) Advances in achieving opioid analgesia without side effects. *Front Pharmacol* 9. <https://doi.org/10.3389/fphar.2018.01388>
- Madariaga-MazÓN A, Marmolejo-Valencia AF, Li Y, Toll L, Houghten RA, Martinez-Mayorga K (2017) Mu-opioid receptor biased ligands: a safer and painless discovery of analgesics? *Drug Discov Today* 22:1719–1729
- Mahmod Al-Qattan MN, Mordi MN (2019) Molecular basis of modulating adenosine receptors activities. *Curr Pharm Design* 25:817–831

- Mains RE, Eipper BA (1981) Coordinate, equimolar secretion of smaller peptide products derived from pro-ACTH/endorphin by mouse pituitary tumor cells. *J Cell Biol* 89:21–28
- Malfroy B, Swerts JP, Guyon A, Roques BP, Schwartz JC (1978) High-affinity enkephalin-degrading peptidase in brain is increased after morphine. *Nature* 276:523–526
- Manglik A, Kruse AC, Kobilka TS, Thian FS, Mathiesen JM, Sunahara RK, Pardo L, Weis WI, Kobilka BK, Granier S (2012) Crystal structure of the micro-opioid receptor bound to a morphinan antagonist. *Nature* 485:321–326
- Manglik A, Lin H, Aryal DK, Mccorvy JD, Dengler D, Corder G, Levit A, Kling RC, Bernat V, Hubner H, Huang XP, Sassano MF, Giguere PM, Lober S, Da D, Scherrer G, Kobilka BK, Gmeiner P, Roth BL, Shoichet BK (2016) Structure-based discovery of opioid analgesics with reduced side effects. *Nature* 537:185–190
- Mansour A, Fox CA, Akil H, Watson SJ (1995) Opioid-receptor mRNA expression in the rat CNS: anatomical and functional implications. *Trends Neurosci* 18:22–29
- Markman J, Gudin J, Rauck R, Argoff C, Rowbotham M, Agaiby E, Gimbel J, Katz N, Doberstein SK, Tagliaferri M, Lu L, Siddhanti S, Hale M (2019) SUMMIT-07. *Pain* 160:1374–1382
- Martin WF, Correll CU, Weiden PJ, Jiang Y, Pathak S, Dipetrillo L, Silverman BL, Ehrlich EW (2019) Mitigation of olanzapine-induced weight gain with samidorphan, an opioid antagonist: a randomized double-blind phase 2 study in patients with schizophrenia. *Am J Psychiatry* 176:457–467
- Mathieu-Kia A-M, Fan L-Q, Kreek MJ, Simon EJ, Hiller JM (2001) μ -, δ - and κ -opioid receptor populations are differentially altered in distinct areas of postmortem brains of Alzheimer's disease patients. *Brain Res* 893:121–134
- Matsushima A, Sese J, Koyanagi KO (2019) Biosynthetic short neuropeptides: a rational theory based on experimental results for the missing pain-relief opioid endomorphin precursor gene. *Chembiochem* 20:2054–2058
- Maves TJ, Pechman PS, Meller ST, Gebhart G (1994) Ketorolac potentiates morphine antinociception during visceral nociception in the rat. *Anesthesiology* 80:1094–1101
- McCarty D, Priest KC, Korthuis PT (2018) Treatment and prevention of opioid use disorder: challenges and opportunities. *Annu Rev Public Health* 39:525–541
- McDonald J, Lambert D (2005) Opioid receptors. *Contin Educ Anaesth Crit Care Pain* 5:22–25
- Meguro Y, Miyano K, Hirayama S, Yoshida Y, Ishibashi N, Ogino T, Fujii Y, Manabe S, Eto M, Nonaka M, Fujii H, Ueta Y, Narita M, Sata N, Yada T, Uezono Y (2018) Neuropeptide oxytocin enhances μ opioid receptor signaling as a positive allosteric modulator. *J Pharmacol Sci* 137:67–75
- Meilandt WJ, Yu G-Q, Chin J, Roberson ED, Palop JJ, Wu T, Scarce-Levie K, Mucke L (2008) Enkephalin elevations contribute to neuronal and behavioral impairments in a transgenic mouse model of Alzheimer's disease. *J Neurosci* 28:5007–5017
- Mello NK, Negus SS (2006) Interactions between kappa opioid agonists and cocaine: preclinical studies. *Annal N Y Acad Sci* 909:104–132
- Ménard C, Herzog H, Schwarzer C, Quirion R (2013) Possible role of dynorphins in Alzheimer's disease and age-related cognitive deficits. *Neurodegener Dis* 13:82–85
- Mendell LM (2011) Computational functions of neurons and circuits signaling injury: relationship to pain behavior. *Proc Natl Acad Sci U S A* 108:15596–15601
- Metzger TG, Ferguson DM (1995) On the role of extracellular loops of opioid receptors in conferring ligand selectivity. *FEBS Lett* 375:1–4
- Mika J, Obara I, Przewlocka B (2011) The role of nociceptin and dynorphin in chronic pain: implications of neuro-glial interaction. *Neuropeptides* 45:247–261
- Mitznegg P, Domschke W, Sprugel W, Domschke S, Subramanian N, Wunsch E, Moroder L, Demling L (1977) Enkephalins inhibit intestinal motility: mode of action. *Acta Hepatogastroenterol (Stuttg)* 24:119–120
- Mogil JS, Pasternak GW (2001) The molecular and behavioral pharmacology of the orphanin Fq/nociceptin peptide and receptor family. *Pharmacol Rev* 53:381–415

- Mores KL, Cummins BR, Cassell RJ, Van Rijn RM (2019) A review of the therapeutic potential of recently developed G protein-biased kappa agonists. *Front Pharmacol* 10
- Motel CW, Coop A, Cunningham CW (2013) Cholinergic modulation by opioid receptor ligands: potential application to Alzheimer's disease. *Mini Rev Med Chem* 13:456–466
- Nandhu MS, Naijil G, Smijin S, Jayanarayanan S, Paulose CS (2010) Opioid system functional regulation in neurological disease management. *J Neurosci Res* 88:3215–3221
- Neves SR, Ram PT, Iyengar R (2002) G protein pathways. *Science* 296:1636–1639
- Niciu MJ, Arias AJ (2013) Targeted opioid receptor antagonists in the treatment of alcohol use disorders. *CNS Drugs* 27:777–787
- Nieto MM, Guen SL, Kieffer BL, Roques BP, Noble F (2005) Physiological control of emotion-related behaviors by endogenous enkephalins involves essentially the delta opioid receptors. *Neuroscience* 135:305–313
- Norn S, Kruse PR, Kruse E (2005) History of opium poppy and morphine. *Dan Medicinhist Arbog* 33:171–184
- North R (1993) Opioid actions on membrane ion channels. In: *Opioids*. Springer, New York, NY
- North RA, Williams JT (1983) How do opiates inhibit neurotransmitter release? *Trends Neurosci* 6:337–339
- Nuechterlein EB (2016) Alterations in endogenous opioid neurotransmission associated with acute and long-term use of drugs of abuse. PhD Thesis, University of Michigan, Ann Arbor, USA. https://deepblue.lib.umich.edu/bitstream/handle/2027.42/120787/emilybn_1.pdf?sequence=1
- O'malley SS, Jaffe AJ, Chang G, Schottenfeld RS, Meyer RE, Rounsaville B (1992) Naltrexone and coping skills therapy for alcohol dependence: a controlled study. *Arch Gen Psychiatry* 49:881–887
- Ohbuchi K, Miyagi C, Suzuki Y, Mizuhara Y, Mizuno K, Omiya Y, Yamamoto M, Warabi E, Sudo Y, Yokoyama A, Miyano K, Hirokawa T, Uezono Y (2016) Ignavine: a novel allosteric modulator of the μ opioid receptor. *Sci Rep* 6:31748
- Owen MJ, Sawa A, Mortensen PB (2016) Schizophrenia. *Lancet* 388:86–97
- Pan J, Cai H (2017) Opioid system in L-Dopa-induced dyskinesia. *Transl Neurodegener.* https://deepblue.lib.umich.edu/bitstream/handle/2027.42/120787/emilybn_1.pdf?sequence=1
- Papadimitriou A, Priftis KN (2009) Regulation of the hypothalamic-pituitary-adrenal axis. *Neuroimmunomodulation* 16:265–271
- Patel RM, Shulman ST (2015) Kawasaki disease: a comprehensive review of treatment options. *J Clin Pharm Ther* 40:620–625
- Pathan H, Williams J (2012) Basic opioid pharmacology: an update. *Br J Pain* 6:11–16
- Pert CB, Snyder SH (1973) Properties of opiate-receptor binding in rat brain. *Proc Natl Acad Sci U S A* 70:2243–2247
- Petrenko AB, Yamakura T, Baba H, Shimoji K (2003) The role of N-methyl-D-aspartate (NMDA) receptors in pain: a review. *Anesth Analg* 97:1108–1116
- Plevuvry BJ (1991) Opioid receptors and their ligands: natural and unnatural. *Br J Anaesth* 66:370–380
- Podvin S, Yaksh T, Hook V (2016) The emerging role of spinal dynorphin in chronic pain: a therapeutic perspective. *Annu Rev Pharmacol Toxicol* 56:511–533
- Polo S, Díaz AF, Gallardo N, LeÁñez S, Balboni G, Pol O (2019) Treatment with the delta opioid agonist Ufp-512 alleviates chronic inflammatory and neuropathic pain: mechanisms implicated. *Front Pharmacol* 10:283
- Portoghese PS, Sultana M, Nagase H, Takemori AE (1988) Application of the message-address concept in the design of highly potent and selective non-peptide .delta. opioid receptor antagonists. *J Med Chem* 31:281–282
- Przewlocki R (2013) Opioid peptides. In: Pfaff DW (ed) *Neuroscience in the 21st century: from basic to clinical*. New York, NY, Springer
- Ramaswamy S, Langford R (2017) Antinociceptive and immunosuppressive effect of opioids in an acute postoperative setting: an evidence-based review. *BJA Educ* 17:105–110

- Ranjan R, Pandey S, Shukla AK (2017) Biased opioid receptor ligands: gain without pain. *Trends Endocrinol Metab* 28:247–249
- Reindl JD, Rowan K, Carey AN, Peng X, Neumeyer JL, McLaughlin JP (2008) Antidepressant-like effects of the novel kappa opioid antagonist MCL-144B in the forced-swim test. *Pharmacology* 81:229–235
- Reisine T, Bell GI (1993) Molecular biology of opioid receptors. *Trends Neurosci* 16:506–510
- Rodríguez-Manzo G, Asai M, Fernández-Guasti A (2002) Evidence for changes in brain enkephalin contents associated to male rat sexual activity. *Behav Brain Res* 131:47–55
- Roth BL, Baner K, Westkaemper R, Siebert D, Rice KC, Steinberg S, Ernsberger P, Rothman RB (2002) Salvinorin A: a potent naturally occurring nonnitrogenous opioid selective agonist. *Proc Natl Acad Sci* 99:11934–11939
- Rubinstein M, Mogil JS, Japón M, Chan EC, Allen RG, Low MJ (1996) Absence of opioid stress-induced analgesia in mice lacking beta-endorphin by site-directed mutagenesis. *Proc Natl Acad Sci* 93:3995–4000
- Rush AM, Cummins TR, Waxman SG (2007) Multiple sodium channels and their roles in electrogenesis within dorsal root ganglion neurons. *J Physiol* 579:1–14
- Russo AF (2017) Overview of neuropeptides: awakening the senses? *Headache* 57(Suppl 2):37–46
- Sarajarvi T, Marttinen M, Natunen T, Kauppinen T, Mäkinen P, Helisalmi S, Laitinen M, Rauramaa T, Leinonen V, Petäjä-Repo U, Soininen H, Haapasalo A, Hiltunen M (2015) Genetic variation in δ -opioid receptor associates with increased β - and γ -secretase activity in the late stages of Alzheimer's disease. *J Alzheimer's Dis* 48:507–516
- Schaffer CB, Nordahl TE, Schaffer LC, Howe J (2007) Mood-elevating effects of opioid analgesics in patients with bipolar disorder. *J Neuropsychiatry* 19:449–452
- Schapira AHV, Chaudhuri KR, Jenner P (2017) Non-motor features of Parkinson disease. *Nat Rev Neurosci* 18:435–450
- Scheuer T (2011) Regulation of sodium channel activity by phosphorylation. *Semin Cell Dev Biol* 22:160–165
- Schuckit MA (2016) Treatment of opioid-use disorders. *N Engl J Med* 375:357–368
- Seseña E, Vega R, Soto E (2014) Activation of μ -opioid receptors inhibits calcium-currents in the vestibular afferent neurons of the rat through a cAMP dependent mechanism. *Front Cell Neurosci* 8:90
- Sgroi S, Tonini R (2018) Opioidergic modulation of striatal circuits, implications in Parkinson's disease and levodopa induced dyskinesia. *Front Neurol* 9:524
- Shalev A, Liberzon I, Marmar C (2017) Post-traumatic stress disorder. *N Engl J Med* 376:2459–2469
- Sheffler DJ, Roth BL (2003) Salvinorin A: the 'magic mint' hallucinogen finds a molecular target in the kappa opioid receptor. *Trends Pharmacol Sci* 24:107–109
- Shekhar A (2019) Role of kappa opioid receptors in symptoms of schizophrenia: what is the neurobiology? *Biol Psychiatry* 86:494–496
- Sher L (2004) The role of endogenous opioids in the placebo effect in post-traumatic stress disorder. *Complement Med Res* 11:354–359
- Shippenberg TS, Elmer GI (1998) The neurobiology of opiate reinforcement. *Crit Rev Neurobiol* 12:267–303
- Shirayama Y, Chaki S (2006) Neurochemistry of the nucleus accumbens and its relevance to depression and antidepressant action in rodents. *Curr Neuropharmacol* 4:277–291
- Smrcka A (2008) G protein $\beta\gamma$ subunits: central mediators of G protein-coupled receptor signaling. *Cell Mol Life Sci* 65:2191–2214
- Snyder SH (2004) Opiate receptors and beyond: 30 years of neural signaling research. *Neuropharmacology* 47:274–285
- Sprouse-Blum AS, Smith G, Sugai D, Parsa FD (2010) Understanding endorphins and their importance in pain management. *Hawaii Med J* 69:70–71

- Stanczyk MA, Livingston KE, Chang L, Weinberg ZY, Puthenveedu MA, Traynor JR (2019) The delta-opioid receptor positive allosteric modulator BMS 986187 is a G-protein-biased allosteric agonist. *Br J Pharmacol* 176:1649–1663
- Takahashi A (2016) Subchapter 7A - enkephalin. In: TAKEI Y, Ando H, Tsutsui K (eds) *Handbook of hormones*. Academic Press, San Diego
- Takeuchi M (2001) The mammalian pars intermedia—structure and function. *Zoolog Sci* 18 (133-144):12
- Tejada HA, Shippenberg TS, Henriksson R (2011) The dynorphin/k-opioid receptor system and its role in psychiatric disorders. *Cell Mol Life Sci* 69:857–896
- Terskiy A, Wannemacher KM, Yadav PN, Tsai M, Tian B, Howells RD (2007) Search of the human proteome for endomorphin-1 and endomorphin-2 precursor proteins. *Life Sci* 81:1593–1601
- Theriot J, Azadfar M, Kum B (2019) *Opioid antagonists*. In: StatPearls. StatPearls Publishing, Treasure Island (FL)
- Thompson GL, Lane JR, Coudrat T, Sexton PM, Christopoulos A, Canals M (2015) Biased agonism of endogenous opioid peptides at the mu-opioid receptor. *Mol Pharmacol* 88:335–346
- Ting-A-Kee R, Van Der Kooy D (2012) The neurobiology of opiate motivation. *Cold Spring Harb Perspect Med* 2:a012096–a012096
- Tomkins DM, Sellers EM (2001) Addiction and the brain: the role of neurotransmitters in the cause and treatment of drug dependence. *CMAJ* 164:817–821
- Torres-Berrio A, Nava-Mesa MO (2019) The opioid system in stress-induced memory disorders: from basic mechanisms to clinical implications in post-traumatic stress disorder and Alzheimer's disease. *Prog Neuropsychopharmacol Biol Psychiatry* 88:327–338
- Tortorici V, Vanegas H (1995) Anti-nociception induced by systemic or PAG-microinjected lysine-acetylsalicylate in rats. Effects on tail-flick related activity of medullary off-and on-cells. *Eur J Neurosci* 7:1857–1865
- Tsagareli MG, Tsiklauri N, Nozadze I, Gurtskaia G (2012) Tolerance effects of non-steroidal anti-inflammatory drugs microinjected into central amygdala, periaqueductal grey, and nucleus raphe. *Neural Regen Res* 7:1029–1039
- Turner AJ (2004) 108 - Neprilysin. In: Barrett AJ, Rawlings ND, Woessner JF (eds) *Handbook of proteolytic enzymes*, 2nd edn. Academic Press, London
- van den Pol AN (2012) Neuropeptide transmission in brain circuits. *Neuron* 76:98–115
- Vanderah TW, Ossipov MH, Lai J, Malan PJ, Porreca F (2001) Mechanisms of opioid-induced pain and antinociceptive tolerance: descending facilitation and spinal dynorphin. *Pain* 92:5–9
- Varamini P, Mansfeld FM, Blanchfield JT, Wyse BD, Smith MT, Toth I (2012) Synthesis and biological evaluation of an orally active glycosylated endomorphin-1. *J Med Chem* 55:5859–5867
- Vaughan CW (1998) Enhancement of opioid inhibition of GABAergic synaptic transmission by cyclo-oxygenase inhibitors in rat periaqueductal grey neurones. *Br J Pharmacol* 123:1479–1481
- Vaughan C, Christie M (1997) Presynaptic inhibitory action of opioids on synaptic transmission in the rat periaqueductal grey in vitro. *J Physiol* 498:463–472
- Vaughan CW, Ingram SL, Connor MA, Christie MJ (1997) How opioids inhibit GABA-mediated neurotransmission. *Nature* 390:611–614
- Veening JG, Barendregt HP (2015) The effects of beta-endorphin: state change modification. *Fluids Barriers CNS* 12:3–3
- Veening JG, Gerrits PO, Barendregt HP (2012) Volume transmission of beta-endorphin via the cerebrospinal fluid; a review. *Fluids Barriers CNS* 9:16–16
- Venkatraman A, Edlow BL, Immordino-Yang MH (2017) The brainstem in emotion: a review. *Front Neuroanat* 11:15
- Vergura R, Balboni G, Spagnolo B, Gavioli E, Lambert DG, McDonald J, Trapella C, Lazarus LH, Regoli D, Guerrini R, Salvadori S, Caló G (2008) Anxiolytic- and antidepressant-like activities of H-Dmt-Tic-Nh-CH(CH₂-COOH)-Bid (UFP-512), a novel selective delta opioid receptor agonist. *Peptides* 29:93–103

- Waldhoer M, Bartlett SE, Whistler JL (2004) Opioid receptors. *Annu Rev Biochem* 73:953–990
- Wang H, Subedi K (2020) δ -Opioid receptor as a potential therapeutic target for ischemic stroke. *Neural Regen Res* 15:20
- Wang H, Wessendorf MW (2001) Equal proportions of small and large DRG neurons express opioid receptor mRNAs. *J Comp Neurol* 429:590–600
- Wang W, Gu J, Li Y-Q, Tao Y-X (2011) Are voltage-gated sodium channels on the dorsal root ganglion involved in the development of neuropathic pain? *Mol Pain* 7:16
- Wang Y, Wang Y-X, Liu T, Law P-Y, Loh HH, Qiu Y, Chen H-Z (2014) μ -opioid receptor attenuates $\text{A}\beta$ oligomers-induced neurotoxicity through mTOR signaling. *CNS Neurosci Ther* 21:8–14
- Williams WA, Grant JE, Winstanley CA, Potenza MN (2008) Current concepts in the classification, treatment, and modeling of pathological gambling and other impulse control disorders. In: McArthur RA, Borsini F (eds) *Animal and translational models for CNS drug discovery*, Academic Press, San Diego
- Wilson AM, Soignier RD, Zadina JE, Kastin AJ, Nores WL, Olson RD, Olson GA (2000) Dissociation of analgesic and rewarding effects of endomorphin-1 in rats. *Peptides* 21:1871–1874
- Wilson-Poe AR, Jeong H-J, Vaughan CW (2017) Chronic morphine reduces the readily releasable pool of GABA, a presynaptic mechanism of opioid tolerance. *J Physiol* 595:6541–6555
- Winters BL, Gregoriou GC, Kissiwa SA, Wells OA, Medagoda DI, Hermes SM, Burford NT, Alt A, Aicher SA, Bagley EE (2017) Endogenous opioids regulate moment-to-moment neuronal communication and excitability. *Nat Commun* 8:14611
- Wu ZQ, Chen J, Chi ZQ, Liu JG (2007) Involvement of dopamine system in regulation of Na^+ , K^+ -ATPase in the striatum upon activation of opioid receptors by morphine. *Mol Pharmacol* 71:519–530
- Wu H, Wacker D, Mileni M, Katritch V, Han GW, Vardy E, Liu W, Thompson AA, Huang XP, Carroll FI, Mascarella SW, Westkaemper RB, Mosier PD, Roth BL, Cherezov V, Stevens RC (2012) Structure of the human kappa-opioid receptor in complex with JDTic. *Nature* 485:327–332
- Xi Z-X (2002) Gabaergic mechanisms of opiate reinforcement. *Alcohol Alcohol* 37:485–494
- Xia R, Mao Z-H (2012) Progression of motor symptoms in Parkinson's disease. *Neurosci Bull* 28:39–48
- Xu C, Liu G, Ji H, Chen W, Dai D, Chen Z, Zhou D, Xu L, Hu H, Cui W, Chang L, Zha Q, Li L, Duan S, Wang Q (2018) Elevated methylation of *OPRM1* and *OPRL1* genes in Alzheimer's disease. *Mol Med Rep*. <https://doi.org/10.3892/mmr.2018.9424>
- Yakovleva T, Bazov I, Cebers G, Marinova Z, Hara Y, Ahmed A, Vlaskovska M, Johansson B, Hochgeschwender U, Singh IN, Bruce-Keller AJ, Hurd YL, Kaneko T, Terenius L, Ekström TJ, Hauser KF, Pickel VM, Bakalkin G (2006) Prodynorphin storage and processing in axon terminals and dendrites. *FASEB J* 20:2124–2126
- Yakovleva T, Marinova Z, Kuzmin A, Seidah NG, Haroutunian V, Terenius L, Bakalkin G (2007) Dysregulation of dynorphins in Alzheimer disease. *Neurobiol Aging* 28:1700–1708
- Yam M, Loh Y, Tan C, Khadijah Adam S, Abdul Manan N, Basir R (2018) General pathways of pain sensation and the major neurotransmitters involved in pain regulation. *Int J Mol Sci* 19:2164
- Yang A, Tsai S-J (2017) New targets for schizophrenia treatment beyond the dopamine hypothesis. *Int J Mol Sci* 18:1689
- Yehuda R, Hoge CW, McFarlane AC, Vermetten E, Lanius RA, Nievergelt CM, Hobfoll SE, Koenen KC, Neylan TC, Hyman SE (2015) Post-traumatic stress disorder. *Nat Rev Dis Primers* 1:15057. <https://doi.org/10.1038/nrdp.2015.57>
- Young J, Mendoza M (2018) Parkinson's disease: A treatment guide. *J Fam Pract* 67:276–286
- Zadina JE, Hackler L, Ge LJ, Kastin AJ (1997) A potent and selective endogenous agonist for the μ -opioid receptor. *Nature* 386:499–502

- Zamponi GW, Currie KPM (2013) Regulation of CaV2 calcium channels by G protein coupled receptors. *Biochim Biophys Acta* 1828:1629–1643
- Zamponi GW, Striessnig J, Koschak A, Dolphin AC (2015) The physiology, pathology, and pharmacology of voltage-gated calcium channels and their future therapeutic Potential. *Pharmacol Rev* 67:821–870
- Zebraski SE, Kochenash SM, Raffa RB (2000) Lung opioid receptors: pharmacology and possible target for nebulized morphine in dyspnea. *Life Sci* 66:2221–2231
- Zhang Z, Pan ZZ (2011) Signaling cascades for δ -opioid receptor-mediated inhibition of GABA synaptic transmission and behavioral antinociception. *Mol Pharmacol* 81:375–383
- Zhang X, Bao L, Arvidsson U, Elde R, Hökfelt T (1997) Localization and regulation of the delta-opioid receptor in dorsal root ganglia and spinal cord of the rat and monkey: evidence for association with the membrane of large dense-core vesicles. *Neuroscience* 82:1225–1242
- Zhao J, Li X, Li B, Chen L, Pei G (2015) A new delta opioid receptor antagonist as a novel drug against Alzheimer's disease. *Alzheimer's Dement* 11:P371
- Zieglgänsberger W (2018) Substance P and pain chronicity. *Cell Tissue Res* 375:227–241